



**International Journal of Biology, Pharmacy  
and Allied Sciences (IJBPAS)**

*'A Bridge Between Laboratory and Reader'*

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## ROLE OF MACROPHAGE DERIVED CYTOKINES IN ARTICULAR CARTILAGE DESTRUCTION IN RHEUMATOID ARTHRITIS

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Received 26<sup>th</sup> Nov. 2020; Revised 27<sup>th</sup> Dec. 2020; Accepted 12<sup>th</sup> Jan. 2021; Available online 1<sup>st</sup> Sept. 2021

<https://doi.org/10.31032/IJBPAS/2021/10.9.5683>

### ABSTRACT

In recent years variety of proinflammatory and anti-inflammatory cytokines have been identified which are involved in pathogenesis and progression of rheumatoid arthritis (RA) in various extent. Macrophage derived cytokines and T cell cytokines having crucial role in the pathogenesis of RA. It is well established that imbalance between proinflammatory and anti-inflammatory cytokines triggers induction of autoimmunity and RA. However exact cytokine pathways that responsible for induction of inflammation and cartilage destruction are not fully cleared. The aim of this review article is to provide a detailed information about the role of various macrophage derived cytokines in the pathogenesis and progression of RA.

**Keywords: Rheumatoid arthritis, cytokine, macrophage, cartilage**

### INTRODUCTION

Rheumatoid arthritis (RA) is an autoimmune & inflammatory disorder that can affect synovial membrane of the synovial joints [1]. The most important hallmark of RA is inflammation & destruction of synovial

membrane [2]. The most important Signs and symptoms are swelling of the joint, arthralgia, redness in the joint and reduction in the rate of movements if RA affects the knees [1]. It has been noted that early cases

of RA affect smaller joints first. Particularly the joints of the fingers of hands and feet are affected first. In the severe cases, the disease spread to the larger joints of knees, wrists, ankles, hips, elbows, and shoulders. The metacarpophalangeal (MCP) joints and proximal interphalangeal (PIP) joints of hands, the metatarsophalangeal (MTP) joints of the feet are the most commonly affected joints by RA. Interaction between synovial fibroblasts, T & B lymphocytes, macrophages and the inflamed synovial cells like plasma cells, macrophages and dendritic cells leads to synovial inflammation. These cells secrete various cytokines that responsible for interaction between these cells. During the inflammatory reactions, various proinflammatory cytokines like IL-1, IL-6, TNF- $\alpha$  are produced which responsible for inflammation of synovial membrane and cartilage destruction through the formation of hyperplastic fibroblast like synoviocytes (FLS) also known as pannus. Pannus releases proteolytic enzymes known as matrix metalloproteinases (MMPs) which destroys cartilage so that pannus invades inside the cartilage [119]. Apart from the matrix proteinase enzymes, mast cell proteases and cathepsin having important role in cartilage destruction [120, 121]. Fibroblastic pannus cell releases various factors that activate

osteoclast to contribute increased resorption of bones [122]. Inflammatory cytokines inhibit mineralization of bones and differentiation of osteoblast cells which results in prevention of osteoblast activity [123]. In some people; the condition can damage a wide variety of body systems, including the skin, eyes, lungs, heart and blood vessels [2]. It has been noted that about 20-30% peoples of RA having the airway problems like pulmonary fibrosis [3]. Lung diseases are more prevalent in males of RA who are smoker & previous history of lung disease [4]. A recent case study indicates that risk of cardiovascular diseases like vascular endothelium damage & early atherosclerosis are 2 times more in the patients of RA than the general population [5]. After 20 years of illness, RA patients have a 40 percent elevated risk of mortality relative to the general population [124]. Some persons with RA having ocular complications like fibrinoid necrosis, vascular occlusion & infiltration of neutrophils & macrophages which leads to deposition of immune complex, matrix metalloproteinases (MMPs) secretions, inflammatory cytokine formation & autoantibody formation [6].

## EPIDEMIOLOGY

1% of total world populations are affected by RA [2]. It prevalently affects women than the

men [1]. women have greater functional disability and pain than men [30]. According to global burden of disease study in the year 2010, it is estimated that 0.24 percent people worldwide effected by RA [26]. 0.5 to 1 percent peoples of united states of America & northern European countries are affected by RA [27, 28]. In united states & in northern European countries the annual incidence of RA is estimated to be 40 per 100,000 persons [29].

#### **ROLE OF MACROPHAGE DERIVED CYTOKINES IN RA**

Cytokines plays an important role in RA. For better understanding, cytokines are divided into two groups: Proinflammatory cytokines & anti-inflammatory cytokines. Balance between proinflammatory & anti-inflammatory cytokines plays an important role in prevention of cartilage destruction in RA. Some cytokines like interleukin (IL-1) and tumor necrosis factor (TNF)- $\alpha$  increases inflammatory responses and responsible for destruction of cartilage. IL-15, IL-17, RANKL, IL-12, and IL-18 etc are the T cell cytokine which have important role in inflammation of joints [8]. Other cytokines, such as IL-4, IL-13 and IL-10 act mainly as anti-inflammatory cytokines. In progressive RA the levels of anti-inflammatory cytokines are too low to neutralize the destructive

effects of proinflammatory cytokines though anti-inflammatory cytokines are also present in the joints of RA [31]. During inflammatory reactions, monocyte & T lymphocytes migrates at the synovium from blood capillaries. Activated macrophages releases variety of cytokines among which IL-1 & TNF- $\alpha$  [7]. It is believed that IL-1 & TNF- $\alpha$  plays a crucial role in cartilage destruction in RA. TNF $\alpha$  is an inflammatory mediator, whereas IL-1 is a crucial cytokine in destruction of articular cartilage [8]. TNF $\alpha$  alone is not so much destructive, but it has some synergistic way that can enhance the destructive behavior of IL-1 [8]. T cells produces IL-15, IL-17, RANKL, IL-12, and IL-18 which plays an important role as proinflammatory cytokines in RA [8].

#### **A. Role of IL-1 family cytokines in RA:**

Several members of the IL-1 superfamily have been implicated in the pathogenesis of RA. 11 cytokines constitutes IL-1 superfamily constitute IL-1 family that shows significant participation in the progression and pathogenesis of RA. The members of IL-1 superfamily are- IL-1 $\alpha$ , IL-1 $\beta$ , IL-1 receptor antagonist (IL-1ra), IL-18, IL-33, IL-36 ( $\alpha$ ,  $\beta$ ,  $\gamma$ , IL-36Ra), IL-37, IL-38 Among of which IL-1 $\alpha$ , IL-1 $\beta$ , IL-18, IL-33, IL-36( $\alpha$ ,  $\beta$ ,  $\gamma$ ) are proinflammatory cytokines involved in

various inflammatory disorders whereas IL-1 receptor antagonist (IL-1Ra), IL-36 receptor antagonist (IL-36Ra), IL-37, IL-38 having anti-inflammatory effects [13]. IL-1 $\alpha$ , IL-1 $\beta$  and natural IL-1 receptor antagonist (IL-1ra) are expressed widely in the synovium [32]. There are two types of IL-1 Receptors (IL-1R) are present. These are type I & type II IL-1 Receptors. IL-1R type I expressed in very low concentration in cells like fibroblast, endothelial cells & in T lymphocytes. IL-1R type I having the capacity of intracellular signaling because of long cytoplasmic tail [34]. IL-1R type II expressed in monocytes, B lymphocytes & in neutrophils. This type of IL-1R is not functionally active because of short cytoplasmic tail.

#### **A.1. Role of IL-1 $\alpha$ in RA:**

IL-1 $\alpha$  are proinflammatory cytokine which is initially available as 31kD precursor molecule known as pro-IL-1 $\alpha$ . Pro- IL-1 $\alpha$  is biologically active. IL-1  $\alpha$  is primarily bound to the membrane. Chondrocytes present in articular cartilage & responsible for the production and maintenance of the cartilage matrix. Pro-inflammatory cytokines like IL-1 $\alpha$  upregulate the production of MMPs and to inhibit proteoglycan synthesis which causes loss of cartilage [33].

#### **A.2. Role of IL-1 $\beta$ in RA:**

IL-1 $\beta$  independently as well as with other inflammatory mediators causes inflammatory & catabolic reactions in cartilage and various parts of joints. It is produced as precursor protein known as pro-IL-1 $\beta$  having 269 amino acids [35, 36]. Active form of IL-1 $\beta$  consist of 153 amino acid residues [37]. IL-1 $\beta$  produced from inactive precursor molecule pro-IL-1 $\beta$  by the enzyme Caspase 1 or IL-1 $\beta$  converting enzyme or ICE by intracellular proteolysis. IL-1 $\beta$  then released into extracellular space [37]. The synthesis of IL-1 $\beta$  in joints are regulated by various cells like osteoblasts, synovial membrane cells, mononuclear cells that infiltrated in the structure of joints during inflammation [38, 39]. IL-1 $\beta$  binds to IL-1R1 receptor leads to recruitment of additional IL-1R3 chain which forms complex with the help of its intracellular portion Toll-IL-1R which further recruits the adapter protein known as MyD88 [40]. This entire complex binds to serine/threonine kinase enzymatic domain of IRAK (IL-1 receptor associated kinase signaling) which further activates TRAF6 which leads to further binding of TAB1, TAK1 & TAB2(41). I $\kappa$ B kinase complex phosphorylation done by TAK1 which further causes activation of transcription factor NF- $\kappa$ B [41]. It also activates c-Jun N-terminal kinase (JNK) and p38MAPK. [41].

Activation of Transcriptional factors leads to expression of various genes encoding for cytokines, chemokines, enzymes, adhesion molecules & various inflammatory mediators [42]. IL-1 $\beta$  having Signiant role in the metabolism of extracellular matrix [43]. Many studies have been confirmed that IL-1 $\beta$  having numerous roles in the destruction of articular cartilage. Type-II collagen & aggrecan which are one of the important structural proteins of extracellular matrix are mainly produced by chondrocytes.IL-1 $\beta$  interfere the synthesis of type-II collagen & aggrecans so that formation of building blocks of extracellular matrix is inhibited [44]. Apart from the inhibition of the synthesis of building blocks of extracellular matrix, IL-1 $\beta$  increases the production of MMPs like MMP-1, MMP-3 & MMP-13 which mainly responsible for cartilage destruction [45, 46]. Apart from these MMPs, release of MMP-9 also stimulated by IL-1 $\beta$  [62].

MMPs are the zinc dependent endopeptidases which are capable of degradation of extracellular matrix [51]. These enzymes are highly catabolic in nature & physiological expressions of genes encoding for MMPs are regulated very strictly. There are 5 types of MMPs have been identified: 1) collagenases (MMP-1, MMP8 & MMP-13), 2) Gelatinases

A&B (MMP-2 & MMP-9 respectively), 3) stromelysins (MMP-3, MMP-10 & MMP-11) 4) membrane type MMPs (MMP-14, MMP-15, MMP-16, MMP-17, MMP24 & MMP-25), and 5) miscellaneous subgroup that includes MMP-7, MMP-11, MMP-12, MMP-20 & MMP-23. Genes encoding for MMPs are expressed mainly during wound healing& during development of embryo [52, 53]. Among these MMPs, MMP-1, MMP-13, MMP-2and MMP-14 are expressed constitutively in adult cartilage & having important role in tissue turnover [54]. Apart from these effects, MMPs also responsible for degradation of various extracellular components [156]. In normal joint, expression of collagenases, gelatinases A and B, stromelysins and membrane type MMPs are very low but their expression is increased in RA joints [10, 157].

MMP-1 is also known as interstitial collagenase. Under normal conditions, expression of MMP-1is low. The expression of MMP-1 occurs during endochondral bone formation, metaphysis & diaphysis of long bones. During metaphysis & diaphysis MMP-1 is mainly produced by osteoblast & hypertrophic chondrocytes [51]. Upregulation of MMP-1 expression occurs in RA & responsible for degradation of type I, type II & type III collagen [55]. However, it

has been noted that MMP-1 shows most destructive activity against type III collagen. The MMP-1 expression level is 10 times greater than the MMP-13 expression level [57].

MMP-3 is also known as stromelysin-1 & are normally absent in the joint tissues. MMP-3 levels increased in the joint tissue during osteoarthritis & RA which results in degradation of collagen type II, IV, III, IX and X, laminin, elastin etc. [51]. MMP-3 also acts as a transcription factor and upregulate the expression of other MMPs [58]. Apart from these effects, MMP-3 also activates proMMP-1 which ultimately responsible for increased MMP-1 formation [59].

Articular chondrocytes mainly express MMP-13 that breaks down type II collagen and shows five to ten times more destructive effects to type II collagen than MMP-1 [60]. Apart from destructive effects on type II collagen, MMP-13 also destroys other matrix components like proteoglycan, osteonectin, perlecan, type IV and type IX collagen [61].

MMP-9 levels are nearly undetectable in most of the cells but its expression can be increased in malignancy and inflammatory disorders like RA [63]. Expression of MMP-9 is stimulated by IL-1 $\beta$  and TNF- $\alpha$ . Apart from its degradation capacity of extra cellular

matrix it can responsible for processing of TNF- $\alpha$  and IL-1 $\beta$  [64, 65].

IL-1 $\beta$  increases the production of ADAMTS metalloproteinases by chondrocytes which causes proteolytic degradation of aggrecan molecules [47]. IL-1 stimulates prostaglandin E2 [48]. Synovial fibroblast cells & chondrocytes expressed prostaglandin E2. Prostaglandin H2 is converted to Prostaglandin E2 with the help of the enzyme prostaglandin E synthase (PGES) [66]. Three different isoenzymes of prostaglandin E2 synthase are present: Microsomal prostaglandin E synthase 1 (mPGES-1), Microsomal prostaglandin E synthase 2 (mPGES-2) & cytosolic prostaglandin E synthase (cPGES) [67]. During RA, inflammation in the synovium causes activation of cyclooxygenase-II (COX-II) enzyme which ultimately causes induction of microsomal prostaglandin E synthase 1 (mPGES-1) which catalyzes the formation of PGE2 [68]. PGE2 produces its variety of effects by acting through its receptors which are collectively known as EP receptors. EP receptors are rhodopsin type 7 alpha helical transmembrane G-protein coupled receptor [67]. Mainly 4 types of EP receptors are present: EP1, EP2, EP3 & EP4 among which EP3 & EP4 receptors are mostly present in human body & PGE2 having higher affinity

to these receptors than EP1 & EP2 receptors [69, 70]. It has been seen that PGE2 by acting through the EP4 receptors produces some proinflammatory responses in RA. PGE2 signaling through EP2 receptors shows some anti-inflammatory properties and also responsible for proliferation of chondrocytes & cartilage regeneration. PGE2 induces apoptosis of chondrocytes in synovium by binding to EP4 receptors [71]. PGE2 signaling through EP4 receptors shows some important catabolic effects in cartilage like activation of matrix metalloproteinase, degradation of type II collagen & proteoglycan biosynthesis suppression [72, 73].

IL-1 $\beta$  increases its own secretion by its autocrine activity as well as it increases the secretion of other cytokines like IL-6, TNF $\alpha$ , IL-8 [74]. During the progression of RA, reactive active species (ROS) production is stimulated by IL-1 $\beta$ . Reactive oxygen species (ROS) increases the generation of hydroxylated radicals, peroxides which ultimately responsible for cartilage destruction [75].

The prime functions of synovial membrane are to prevent friction of joints by releasing synovial fluids which lubricate the surface of the joints, provide nutrition to the cartilage & provide joint support [9]. The synovial

membrane consists of two layers: The intimal lining layer & sublining layer (subintima). The intimal layer is the superficial layer & faces towards the intra articular cavities. Synovial fluids are produced by the intimal layer & responsible for the lubricating properties to the joints. Macrophage like synoviocytes (Type A Synoviocytes) & fibroblast like synoviocytes or FLS (Type B Synoviocytes) are the two types of cells responsible for the production of intimal layer of synovial membrane [9]. Intimal layer having the thickness of 2-3 cells [9]. The sublining layer is made up of loose connective tissue, blood and lymph vessels, nerve fibers macrophages, mast cells, fibroblasts and T lymphocytes [49]. Hyperplasia of fibroblast like synoviocytes (FLS) is one of the crucial histopathological characterization of RA. IL-1  $\beta$  produced by the activated macrophages responsible for the hyperplasia of FLS [10]. Hyperplastic FLS cells lack of apoptotic pathway [11]. The hyperplastic FLS characterized by increased expression of the MDM 4 protein that inhibits the activity of tumor suppressor gene p53 which inhibits the activity of transcription factor p53 [50]. Hyperplastic FLS cells leads to the formation of pannus which causes bone erosion & cartilage destruction [12].

A.3. Role of IL-1 receptor antagonist (IL-1ra) in RA:

IL-1ra is a type I & type II IL-1 receptor antagonist that is produced in the body. There are two types of IL-1ra: intracellular form (icIL-1ra) & secreted form (sIL-1ra)[76]. Activated macrophages & monocytes secrete sIL-1ra & during inflammation its secretion is increased whereas icIL-1ra mostly present in the epithelial cells [77]. Monocytes induces the expression of icIL-1ra mRNA [77]. IL-1ra binds to both type I & type II IL-1 receptors thereby block the signal transduction of IL-1 $\alpha$  and IL-1 $\beta$ . Although homology in amino acid sequence of IL-1ra & IL-1 $\alpha$  is 30% and with IL-1 $\beta$  is 19%, IL-1ra binding to both type I & type II IL-1 receptors does not activate the receptors to induce signal transduction [78]. It has been seen that binding capacity of IL-1ra to type I IL-1 receptors are more than IL-1 whereas binding capacity of IL-1ra to type II IL-1 receptors are less than IL-1 [79].

Although IL-1ra are present in the RA synovium, their levels are significantly less than IL-1. Ratio range of IL-1 to IL-1ra of 3.6 to 1.2 is significantly less than the 10- to 100-fold excess in IL-1ra which is required to inhibit 50% biological activity of IL-1 [80]. Since activation of cellular response

requires only 5% IL-1 receptor occupation by IL-1, a large amount of IL-1ra would be required therapeutically to inhibit inflammation induced by IL-1.

A.4. Role of IL-18 in RA:

IL-18 is a proinflammatory cytokine which can be detected in the arthritic synovium and in synovial fluids. It is mainly synthesized as precursor molecule pro IL-18 which is having 192 amino acid residues [87]. Pro IL-18 is converted to active protein IL-18 with 157 amino acid residues [87] with the help of the enzyme IL-1 $\beta$  converting enzyme (Also known as caspase 1) [81]. Activated macrophages mainly secrete IL-18 [82]. Apart from macrophages, chondrocytes [83], osteoblasts [84], synovial fibroblasts [85] & keratinocytes [86] also secrete IL-18. IL-18 receptor having ligand binding chain known as IL-18R $\alpha$  which binds with IL-18 with low affinity. Cells which express the beta chain of IL-18 receptors (IL-18R $\beta$ ) as coreceptor forms a strong affinity complex with IL-18-IL-18R $\alpha$ . This complex act through toll/IL-1 receptor (TIR) domain and responsible for recruitment of MyD88, interleukin 1 receptor associated kinase (IRAKs), TNF receptor associated factor 6 (TRAF-6) and ultimately responsible for NF $\kappa$ B and other proinflammatory cytokine release. Binding capacity of IL-18 binding protein (IL-18BP)

to soluble IL-18 is much higher than IL-18R $\alpha$  and inhibit binding of IL-18 receptor with IL-18. Thus IL-18-BP suppress the activity of IL-18 [132].

IL-18 increases the secretion of TNF $\alpha$ , interferon gamma from Th1 cells & granulocyte-macrophage colony stimulating factor (GM-CSF) by natural killer cells (NK Cells) and T lymphocytes [88]. IL-18 expressed in the intimal lining layer & sublining layer (subintima) of the synovial membrane & activates macrophages produced by these lining layer. Activated macrophage releases various proinflammatory cytokines which is having pivotal role in articular cartilage destruction and inflammation. Synovial fibroblast & chondrocytes located proximity to the activated macrophages releases various inflammatory mediators & MMPs like MMP-13, MMP-3, MMP-1 [89]. IL-18 stimulates synovial cells & chondrocytes which results in increased production of iNOS, COX-2, PGE2, VEGF & IL-6 [90, 91].

#### **A.5. Role of IL-33 in RA:**

IL-33 is an important member of IL-1 superfamily & having important role in the pathogenesis of various autoimmune & inflammatory disorders like RA. It is produced as 30-Kd precursor protein which is cleaved by caspase-1 to release active IL-

33 with molecular weight 18-Kd [92]. IL-33 having its specific receptor known as ST2L which is mainly expressed in Th2 cells, NKT cells, eosinophils, basophils & in mast cells [93-100]. For signaling pathway of IL-33 mediated NF $\kappa$ B activation & cytokine production by Th2 cells, tumor necrosis factor receptor associated factor 6 (TRAF6) plays an important role as an adapter protein [101]. Expression of IL-33 in inflamed synovial membrane largely increased [102]. In collagen induced arthritis (CIA) model, ST2 expression is increased in the mast cells through which IL-33 produces its response. IL-33 causes activation of mast cells which leads to release of proinflammatory cytokines like IL-6 & IL-1 $\beta$ . IL-33 also stimulates CD4+T cells to release cytokines like IL-13 & IL-5. These cytokines cause B lymphocyte activation and antibody production which ultimately responsible for inflammation of articular joints and degranulation of mast cells [103]. Activated T cells and macrophages produces various proinflammatory cytokines that stimulates the release of IL-33 from macrophages which is a powerful chemoattractant to neutrophils [104].

#### **A.6. Role of IL-36 in RA:**

IL-36, a member of the IL-1 superfamily, is an inflammatory cytokine. IL-36 consist of

three agonists: IL-36 $\alpha$ , IL-36 $\beta$  and IL-36 $\gamma$ , and IL-36 receptor antagonist (IL-36Ra). The encoding genes for the protein family IL 36 are located on human chromosome 2 [105]. In order to obtain their complete agonist or antagonist function, IL-36  $\alpha$ ,  $\beta$ ,  $\gamma$  and IL-36Ra cytokines need to be processed. IL-36 Ra does not have antagonistic potential in its native form; similarly, IL-36  $\alpha$ ,  $\beta$  and  $\gamma$  are 100-1000 times less active than their refined counterparts [108]. Neutrophils releases various enzymes like elastase, cathepsin G (Also known as Cat G) & proteinase-3 which are responsible for IL-36 $\alpha$ , IL-36 $\beta$  and IL-36 $\gamma$  activation and processing. Elastase or Cat G cleaves at alanine 4 & lysine 3 position of inactive IL-36 $\alpha$  respectively and causes formation of activated IL-36 $\alpha$  [109]. Cat G cleaves at arginine 5 (Arg5) to produce activated IL-36 $\beta$ . Elastase or proteinase-3 cleaves at the position of Valine 15 to produce activated IL-36 $\gamma$  from IL-36 $\gamma$  precursors [109]. IL-36Ra precursors converted to activated IL-36Ra neutrophil elastase [110].

IL-36 cytokines are expressed in the synovial tissue of RA patients by different cells: plasma cells, macrophages and to a lesser degree, fibroblasts, endothelial cells and dendritic cells [106, 107]. IL-36  $\alpha$ , IL-36Ra & IL-36 receptors (IL-36R) are expressed in

the synovium of RA patients and also found that expression of IL-36  $\alpha$  is higher in RA than osteoarthritis [111]. During inflammatory response in CIA model in mice, gene expression of IL-36  $\alpha$ , IL-36 $\beta$ , IL-36Ra & IL-36 receptors (IL-36R) is increased [112]. IL-36R expressed on FLS cells of synovial membrane which on binding with IL-36 activates FLS cells to produce variety of inflammatory cytokines like IL-6 & IL-8 & MMPs [113]. IL-36  $\gamma$  helps in differentiation of T lymphocytes and release of IL-9 from lymphocytes (Th9) [114], which helps in increased survival of neutrophils and enhanced differentiation of Th17 cells in the synovium [115]. Synovium and peripheral blood of inflammatory arthritic patients having increased levels of Th9 cells [116] which correlates with the disease progression in RA [115]. IL-36 and IL-36R can stimulate dendritic cells & helps in the maturation of dendritic cells. Dendritic precursor cells on exposure to IL-16 releases IL-12 and helps in T cell to Th1 cell differentiation. IL-36 $\beta$  having a stimulatory effect on Th1 effects [117]. However, it has been seen that magnitude of the stimulatory responses of IL-36 in chondrocytes and fibroblast like synoviocytes (FLS) are marked lower than that of IL-1 which indicates that IL-36

probably the main inflammatory cytokine in human RA [118].

IL-36Ra plays an important antagonistic role in IL-36 mediated cytokine release and shows an antagonistic effect on various inflammatory disorders like RA and crohn's disease [22]. IL-36Ra blocks IL-36R by inhibiting IL-1RAcP recruitment. At the same time single immunoglobulin IL-1R-related molecule (SIGIRR) recruited by IL-36Ra which inhibits TIR domain to produce antagonistic effects [22, 23].

#### A.7. Role of IL-37 in RA:

Interleukin-37 (IL-37) was first found and recognized by Kumar *et al.* in 2020 through computational sequence analysis [124]. Nold *et al.* identified that IL-37 could inhibit innate immune response [125]. The human IL-37 gene has a length of 3,617kb and located in chromosome 2 [126]. Five IL-37 isoforms have been identified: IL-37a, IL-37b, IL-37c, IL-37d & IL-37e which are encoded by six exons [126]. All IL-37 isoforms produced as precursor molecules which upon cleavage by caspase-1 converted to active forms. Cleavage site of caspase-1 is located between amino acid residues (D20 & E21) expressed by exons [127]. Caspase-1 mutation inhibits the translocation of IL-37 into nucleus to form complex with Smad3 gene (Mothers against decapentaplegic

homolog 3) which responsible for downregulation of transcription of some key genes [127]. Binding of IL-37 to IL-18  $\alpha$  facilitates IL-1R8 recruitment instead of IL-18R  $\beta$  [128]. Tyr 536 & Ser447 residues are absent in toll/IL-1 receptor (TIR) domain of IL-1R8. Binding of TIR domain of IL-1R8 to MyD88 results in inadequate signal transduction and formation of multiple signaling that inhibits the inflammatory reactions [128].

IL-37 can be found in many tissues, but level of expression in healthy human tissue is poor [125]. Tissue specificity is the characteristics of IL-37 expression. Thymus glandular cells, lungs, liver, colon, uterus, natural killer cells, activated B lymphocytes mainly express IL-37a, IL-37b & IL-37c whereas testis & bone marrow mainly express IL-37d and IL-37e. Brain only express IL-37a, Kidney only express IL-37b and heart only express IL-37c [126].

Elevated levels of IL-37 can be detected in plasma and synovial tissue of RA patients but it is nearly undetectable in healthy individuals [125]. Decreased levels of IL-37 can be detected in the patients with remissive state of RA compared to active cases of RA. Recombinant IL-37 administered by intra-articular injection in streptococcal induced arthritic model [129] in mice or in collagen

induced arthritic model [130] in mice causes downregulation of Th17 cytokine & IL-17 production and improves symptoms of RA. IL-37 also decreases the expression of IL-1 $\beta$ , IL-6. IL-37 also have an inhibitory effect on Th17 proliferation but don't have any significant role in differentiation of Th17 cells [130]. Apart from these, IL-37 inhibits osteoclast formation (osteoclastogenesis) mediated by RANKL and also inhibits bone resorptive action of osteoclast cells [131]. It has been seen that IL-37 binds to IL-18R because IL-37 having some homology in amino acid sequence with IL-18 (Lys-124 & glucoses-35). Binding of IL-37 to IL-18R $\alpha$  recruits IL-IR8 to produce IL-37/ IL-18R $\alpha$ / IL-IR8 complex which blocks the activity of IL-18 [133].

#### A.8. Role of IL-38 in RA:

The newest IL-1 family member is IL-38. Four exons of IL-38 gene are located on chromosome 2q13-14.1 in cluster of IL-1 family gene [14]. IL-38 gene located in close proximity of IL-36Ra gene (IL-36RN) and 49,479 base pair upstream from IL-1Ra gene (IL-1RN) [123]. IL-38 precursor protein having molecular mass of 16.9kD and having 152 amino acid sequence [15]. It has been seen that caspase-1 cleavage site and signal peptide is absent in IL-38 [16, 124]. IL-38 shares 41% amino acid sequence homology

with IL-1Ra gene and 43% with IL-36Ra gene [17, 124]. However, IL-38 having 14-30% amino acid sequence homology with IL-1 $\beta$  and other cytokines of IL-1 superfamily.

IL-38 having important role in inhibition of various proinflammatory cytokine signaling pathways by binding with various receptors like IL-1R1 (interleukin-1 receptor 1), IL-36R (interleukin-36R) and IL-1RAPL1 (interleukin-1 receptor accessory protein-like 1) thereby regulates the functions of various mononuclear cells, macrophages and T lymphocytes. IL-1RI is an IL-1R family member which comprises of IL-1R1 to IL-1R10. Binding of IL-1 to IL-1R1 results in activation of the receptor and leads to formation of interleukin-1 receptor 1 (IL-1R1)-interleukin-1 receptor accessory protein (IL-1RAcP) complex. IL-1R1-IL-1RAcP complex recruits myeloid differentiation primary response 88 (MyD88) through TIR domain which further activates NF- $\kappa$ B pathway and mitogen activated protein kinase (MAPK) pathways. MAPK pathway act through p<sup>38</sup>, extracellular regulated protein kinases (ERK) 1/2 and c-Jun N-terminal kinase (JNK) which ultimately responsible for activator protein-1 (AP-1) activation. AP-1 and NF- $\kappa$ B stimulates proinflammatory cytokine production by binding with specific genes

[20, 21]. IL-1Ra blocks recruitment of IL-1RAcP and recruits inhibitory receptor single immunoglobulin IL-1R-related molecule (SIGIRR) that inhibit the cytokine release pathways. As IL-38 shares 41% amino acid sequence homology with IL-1Ra, it indicates that IL-38 might have similar anti-inflammatory property of IL-1Ra [17]. However, it has been seen that IL-38 having affinity to IL-1R1 but the affinity is much lower compared to IL-1 and IL-1Ra [17].

Apart from IL-1R1, IL-38 also act as an antagonist to IL-36R. IL-36 binds to IL-36R and activate it. Activation of IL-36R causes recruitment of interleukin-1 receptor accessory protein (IL-1RAcP) which forms cytosolic toll-interleukin 1 receptor (TIR) domain [18]. IL-1R1-IL-1RAcP complex recruit myeloid differentiation primary response 88 (MyD88) through TIR domain which ultimately responsible for release of various proinflammatory cytokines like TNF- $\alpha$ , IL-17, IFN- $\gamma$  [19]. IL-38 inhibits the binding of IL-36 to IL-36R as well as recruitment of interleukin-1 receptor accessory protein (IL-1RAcP) therefore prevent the release of various proinflammatory cytokines.

IL-38 also act on IL-1RAPL1 (interleukin-1 receptor accessory protein-like 1) receptor. It is an orphan receptor characterized by

presence of intracellular toll/IL-1 receptor (TIR) domain and three extracellular immunoglobulin domains [20]. It has been seen that JNK/API pathway is activated by IL-1RAPL1 [24]. Full length IL-38 on binding with IL-1RAPL1 increases the production of IL-6 by activating JNK/API pathway whereas truncated IL-38 on binding with IL-1RAPL1 inhibit activation of Th17 cells which responsible for decreasing the release of IL-8 and IL-6 that ultimately decreases activation of macrophages [25].

IL-38 have important anti-inflammatory effects on autoimmune disorders like RA. Apart from Th1 and Th2 cells, a novel subtype of T lymphocyte has been identified which was named Th17 cells because of its capacity to express IL-17. The most important difference between Th17 cells and other T cells is that Th17 cells can express IL-22 and IL-17 [135, 136]. In various studies it has been seen that Th17 cells having direct involvement in the pathogenesis of various autoimmune disorders like RA, psoriasis, inflammatory bowel disease etc. [137]. mRNA expression of IL-38 significantly increased in CIA model [113]. Adeno-associated virus -IL-38 (AAV-IL-38) injected to mice of CIA model showed significant reduction of expression of various proinflammatory cytokines released

from Th17 cells (IL-22, IL-17, IL-23), chemokines etc. [138]. Clinical score and severity of RA decreases significantly in AAV-IL-38 treated mice of CIA model. After AAV-IL-38 treatment, density of macrophages in the inflamed synovial area significantly decreases [138]. IL-38 also downregulate the expression of TNF $\alpha$ , IL-10 levels. when IL-38 knock out mice induced K/BxN model of RA, expression of IL-6, IL-1 $\beta$  in the joint significantly increased compared to control group of mice which indicates the role of IL-38 in the reduction of IL-6, IL-1 $\beta$  expression [139].

#### **B. Role of TNF- $\alpha$ in RA:**

An important signaling protein in adaptive and innate and immune response is tumor necrosis factor-alpha (TNF- $\alpha$ ). It is having important role in the regulation of development of embryo, formation of germinal center, sleep-wake cycle etc. Apart from these, TNF- $\alpha$  responsible for release of various proinflammatory cytokines and enhancement of adhesion and permeability of lymphocyte, monocyte, neutrophil at the site of inflammation [140, 141]. Macrophages are mainly responsible for release of TNF- $\alpha$ . However, apart from macrophages, B lymphocytes, some types of T lymphocytes, natural killer cells, fibroblasts also release TNF- $\alpha$  at low level [142, 143]. Human TNF-

$\alpha$  gene located on chromosome 6. 1.7kb messenger RNA codes for TNF- $\alpha$ . Initially 26-kDa membrane bound TNF- $\alpha$  (mTNF- $\alpha$ ) with 233 amino acid sequence produced. Soluble TNF- $\alpha$  (sTNF- $\alpha$ ) with molecular weight of 17kDa is produced from mTNF- $\alpha$  by TNF- $\alpha$  converting enzyme [144, 145]. TNF- $\alpha$  converting enzyme cleaves mTNF- $\alpha$  at C-terminal domain of extracellular region to produce sTNF- $\alpha$  with 157 amino acids.

Two different types of TNF- $\alpha$  receptors have been identified which are responsible for TNF- $\alpha$  mediated signal transduction. Both tumor necrosis factor- receptor 1 (TNF-R1) and tumor necrosis factor- receptor 2 (TNF-R2) are transmembrane receptors with equal size of intracellular and extracellular domain and with a single transmembrane chain. Gene encoding for TNF-R1 located on chromosome 12p13. TNF-R1 having approximate molecular weight of 55-60kDa and composed of 455 amino acid sequence whereas, gene encoding for TNF-R2 located on chromosome 1p36. TNF-R2 having approximate molecular weight of 70-80 kDa and composed of 461 amino acid sequence. In almost all cells in human body TNF-R1 is expressed and can be activated by both sTNF $\alpha$  and mTNF $\alpha$  whereas, TNF-R2 mainly expressed in T cells, microglia, oligodendrocytes and endothelial cells [146].

TNF-R2 is fully activated by mTNF $\alpha$  only. On binding with TNF-R2, mTNF $\alpha$  produces stable complex to dissociate [147]. sTNF $\alpha$  shows low affinity towards TNF-R2 and results in weak signal transduction [148].

There are numerous proinflammatory responses are present which are exerted by TNF- $\alpha$ . In the patient with RA TNF- $\alpha$  can be detected in both synovial tissue and synovial fluid [149]. Apart from these, In the cartilage-pannus junction TNF- $\alpha$  can be detected which indicates its association in degradation of cartilage [150]. Though the factors that increases TNF $\alpha$  release from inflamed synovium is not properly known, recent studies indicate that IL-15 can be a potential candidate for stimulating the release of TNF $\alpha$  from inflamed synovium [151]. There is significant proof that TNF- $\alpha$  has a pathogenic role in synovitis [152]. TNF- $\alpha$  increases the expression of various adhesion molecules like intracellular adhesion molecule 1(ICAM 1), endothelial leukocyte adhesion molecule (ELAM-1), vascular cell adhesion molecule 1(VCAM 1) which act as chemoattractant to inflammatory cells like monocytes and neutrophils, resulting in aggregation of inflammatory cells in the synovium [153]. TNF- $\alpha$  stimulates proliferation of synovial cell lining which leads to formation of synovial hyperplasia

[153]. Stimulation of synovial cell lining by TNF- $\alpha$  results in production of various proinflammatory cytokines like IL-1, GM-CSF and various types of MMPs like collagenase, stromelysin which ultimately leads to destruction of cartilage in RA [153]. Stimulation of synovial cell lining by TNF- $\alpha$  also responsible for formation of PGE<sub>2</sub> which involved in generation of pain [154]. Administration of anti-TNF- $\alpha$  monoclonal antibody to TNF- $\alpha$  over expressed mice with RA shows significant prevention of disease indicating potential role of TNF- $\alpha$  in RA [8]. Inhibition of synovitis can be detected in CIA model by the administration of anti-TNF- $\alpha$  monoclonal antibody also indicates potential role of TNF- $\alpha$  in RA [155].

## CONCLUSION

RA is a progressive degenerative disorder of joints which is characterized by severe synovial inflammation and degradation of cartilage. Apart from the catabolic effects of various pro inflammatory cytokines various anabolic anti-inflammatory cytokines also expressed. Dysregulation of balance between proinflammatory and anti-inflammatory cytokines responsible for articular cartilage destruction in RA. Various types macrophage derived proinflammatory cytokines like IL-1 $\alpha$ , IL-1 $\beta$ , IL-18, IL-33, IL-36 ( $\alpha$ ,  $\beta$  &  $\gamma$ ) & TNF-  $\alpha$  shows numerous inflammatory

response that ultimately responsible for cartilage destruction in RA. In this article impact of various macrophage derived pro and anti-inflammatory cytokines on RA have been discussed. Authors of this article tried to summarize the important information regarding the role of macrophage derived pro & anti-inflammatory cytokines in the pathogenesis of RA.

#### REFERENCES

- [1] Guo Q, Wang Y, Xu D, Nossent J, Pavlos NJ, Xu J. Rheumatoid arthritis: pathological mechanisms and modern pharmacologic therapies. *Bone research*. 2018 Apr 27;6(1):1-4.
- [2] Kahlenberg JM, Fox DA. Advances in the medical treatment of rheumatoid arthritis. *Hand clinics*. 2011 Feb 1;27(1):11-20.
- [3] Devouassoux G, Cottin V, Lioté H, Marchand E, Frachon I, Schuller A, Béjui-Thivolet F, Cordier JF. Characterisation of severe obliterative bronchiolitis in rheumatoid arthritis. *European Respiratory Journal*. 2009 May 1;33(5):1053-61.
- [4] Kelly C, Saravanan V. Treatment strategies for a rheumatoid arthritis patient with interstitial lung disease. *Expert opinion on pharmacotherapy*. 2008 Dec 1;9(18):3221-30.
- [5] Bergholm R, Leirisalo-Repo M, Vehkavaara S, Mäkimattila S, Taskinen MR, Yki-Järvinen H. Impaired responsiveness to NO in newly diagnosed patients with rheumatoid arthritis. *Arteriosclerosis, thrombosis, and vascular biology*. 2002 Oct 1;22(10):1637-41.
- [6] Artifoni M, Rothschild PR, Brézin A, Guillevin L, Puéchal X. Ocular inflammatory diseases associated with rheumatoid arthritis. *Nature Reviews Rheumatology*. 2014 Feb;10(2):108.
- [7] Arend WP. The pathophysiology and treatment of rheumatoid arthritis. *Arthritis & Rheumatism: Official Journal of the American College of Rheumatology*. 1997 Apr;40(4):595-7.
- [8] Keffer J, Probert L, Cazlaris H, Georgopoulos S, Kaslaris E, Kioussis D, Kollias G. Transgenic mice expressing human tumour necrosis factor: a predictive genetic model of arthritis. *The EMBO journal*. 1991 Dec;10(13):4025-31.
- [9] Bertolini DR, Nedwin GE, Bringman TS, Smith DD, Mundy GR. Stimulation of bone resorption and inhibition of bone formation in vitro by human tumour necrosis factors. *Nature*. 1986 Feb 6;319(6053):516-8.

- [10] Konttinen YT, Ainola M, Valleala H, Ma J, Ida H, Mandelin J, Kinne RW, Santavirta S, Sorsa T, López-Otín C, Takagi M. Analysis of 16 different matrix metalloproteinases (MMP-1 to MMP-20) in the synovial membrane: different profiles in trauma and rheumatoid arthritis. *Annals of the rheumatic diseases*. 1999 Nov 1;58(11):691-7.
- [11] Jacobs RA, Perrett D, Axon JM, Herbert KE, Scott DL. Rheumatoid synovial cell proliferation, transformation and fibronectin secretion in culture. *Clinical and experimental rheumatology*. 1995;13(6):717-23.
- [12] Szopińska et al. The pathogenesis of rheumatoid arthritis in radiological science. Part I: Formation of inflammatory infiltrates within the synovial membrane. *Journal of ultrasonography*. 2012 Jun; 12(49): 202–213.
- [13] Dinarello CA. Introduction to the interleukin-1 family of cytokines and receptors: Drivers of innate inflammation and acquired immunity. *Immunological reviews*. 2018 Jan;281(1):5.
- [14] Nicklin MJ, Barton JL, Nguyen M, FitzGerald MG, Duff GW, Kornman K. A sequence-based map of the nine genes of the human interleukin-1 cluster. *Genomics*. 2002 May 1;79(5):718-25.
- [15] Yuan X, Peng X, Li Y, Li M. Role of IL-38 and its related cytokines in inflammation. *Mediators of inflammation*. 2015 Jan 1;2015.
- [16] Lin H, Ho AS, Haley-Vicente D, Zhang J, Bernal-Fussell J, Pace AM, Hansen D, Schweighofer K, Mize NK, Ford JE. Cloning and characterization of IL-1HY2, a novel interleukin-1 family member. *Journal of Biological Chemistry*. 2001 Jun 8;276(23):20597-602.
- [17] Bensen JT, Dawson PA, Mychaleckyj JC, Bowden DW. Identification of a novel human cytokine gene in the interleukin gene cluster on chromosome 2q12-14. *Journal of Interferon & Cytokine Research*. 2001 Nov 1;21(11):899-904.
- [18] Veerdonk FV, Netea M. New insights in the immunobiology of IL-1 family members. *Frontiers in immunology*. 2013 Jul 8;4:167.
- [19] Nishida A, Hidaka K, Kanda T, Imaeda H, Shioya M, Inatomi O, Bamba S,

- Kitoh K, Sugimoto M, Andoh A. Increased expression of interleukin-36, a member of the interleukin-1 cytokine family, in inflammatory bowel disease. *Inflammatory bowel diseases*. 2016 Feb 1;22(2):303-14.
- [20] Garlanda C, Dinarello CA, Mantovani A. The interleukin-1 family: back to the future. *Immunity*. 2013 Dec 12;39(6):1003-18.
- [21] Boraschi D, Tagliabue A. The interleukin-1 receptor family. *In Seminars in immunology* 2013 Dec 15; 25(6): 394-407.
- [22] Boutet MA, Bart G, Penhoat M, Amiaud J, Brulin B, Charrier C, Morel F, Lecron JC, Rolli-Derkinderen M, Bourreille A, Vigne S. Distinct expression of interleukin (IL)-36 $\alpha$ ,  $\beta$  and  $\gamma$ , their antagonist IL-36Ra and IL-38 in psoriasis, rheumatoid arthritis and Crohn's disease. *Clinical & Experimental Immunology*. 2016 May;184(2):159-73.
- [23] Riva F, Bonavita E, Barbati E, Muzio M, Mantovani A, Garlanda C. TIR8/SIGIRR is an interleukin-1 receptor/toll like receptor family member with regulatory functions in inflammation and immunity. *Frontiers in immunology*. 2012 Oct 29;3:322.
- [24] Pavlowsky A, Zanchi A, Pallotto M, Giustetto M, Chelly J, Sala C, Billuart P. Neuronal JNK pathway activation by IL-1 is mediated through IL1RAPL1, a protein required for development of cognitive functions. *Communicative & integrative biology*. 2010 May 1;3(3):245-7.
- [25] Mora J, Schlemmer A, Wittig I, Richter F, Putyrski M, Frank AC, Han Y, Jung M, Ernst A, Weigert A, Brüne B. Interleukin-38 is released from apoptotic cells to limit inflammatory macrophage responses. *Journal of molecular cell biology*. 2016 Oct 1;8(5):426-38.
- [26] Pelletier JP, Mineau F, Fernandes JC, Duval N, Martel-Pelletier J. Diacerhein and rhein reduce the interleukin 1beta stimulated inducible nitric oxide synthesis level and activity while stimulating cyclooxygenase-2 synthesis in human osteoarthritic chondrocytes. *The Journal of rheumatology*. 1998 Dec 1;25(12):2417-24.
- [27] Cross M, Smith E, Hoy D, Carmona L, Wolfe F, Vos T, Williams B, Gabriel S, Lassere M, Johns N, Buchbinder R. The global burden of rheumatoid arthritis: estimates from the global burden of disease 2010 study. *Annals*

- of the rheumatic diseases. 2014 Jul 1;73(7):1316-22.
- [28] Myasoedova E, Crowson CS, Kremers HM, Therneau TM, Gabriel SE. Is the incidence of rheumatoid arthritis rising?: results from Olmsted County, Minnesota, 1955–2007. *Arthritis & Rheumatism*. 2010 Jun;62(6):1576-82.
- [29] Hunter TM, Boytsov NN, Zhang X, Schroeder K, Michaud K, Araujo AB. Prevalence of rheumatoid arthritis in the United States adult population in healthcare claims databases, 2004–2014. *Rheumatology international*. 2017 Sep 1;37(9):1551-7.
- [30] Eriksson JK, Neovius M, Ernestam S, Lindblad S, Simard JF, Askling J. Incidence of rheumatoid arthritis in Sweden: a nationwide population-based assessment of incidence, its determinants, and treatment penetration. *Arthritis care & research*. 2013 Jun;65(6):870-8.
- [31] Lesuis N, Befrits R, Nyberg F, van Vollenhoven RF. Gender and the treatment of immune-mediated chronic inflammatory diseases: rheumatoid arthritis, inflammatory bowel disease and psoriasis: an observational study. *BMC medicine*. 2012 Dec 1;10(1):82.
- [32] Isomäki P, Punnonen J. Pro-and anti-inflammatory cytokines in rheumatoid arthritis. *Annals of medicine*. 1997 Jan 1;29(6):499-507.
- [33] Dayer JM. The pivotal role of interleukin-1 in the clinical manifestations of rheumatoid arthritis. *Rheumatology*. 2003 May 1;42(suppl\_2): ii3-10.
- [34] Wilbrink B, Nietfeld JJ, Den Otter W, Van Roy JL, Bijlsma JW, Huber-Bruning O. Role of TNF $\alpha$ , in relation to IL-1 and IL-6 in the proteoglycan turnover of human articular cartilage. *Rheumatology*. 1991 Aug 1;30(4):265-71.
- [35] Sims JE, Gayle MA, Slack JL, Alderson MR, Bird TA, Giri JG, Colotta F, Re F, Mantovani A, Shanebeck K. Interleukin 1 signaling occurs exclusively via the type I receptor. *Proceedings of the National Academy of Sciences*. 1993 Jul 1;90(13):6155-9.
- [36] Dinarello CA. Overview of the interleukin-1 family of ligands and receptors. In *Seminars in immunology* 2013 Dec 15 (Vol. 25, No. 6, pp. 389-393). Academic Press.
- [37] Auron PE, Webb AC, Rosenwasser LJ, Mucci SF, Rich A, Wolff SM,

- Dinarello CA. Nucleotide sequence of human monocyte interleukin 1 precursor cDNA. Proceedings of the National Academy of Sciences. 1984 Dec 1;81(24):7907-11.
- [38] Piccioli P, Rubartelli A. The secretion of IL-1 $\beta$  and options for release. In Seminars in immunology 2013 Dec 15 (Vol. 25, No. 6, pp. 425-429). Academic Press.
- [39] De Lange-Brokaar BJ, Ioan-Facsinay A, Van Osch GJ, Zuurmond AM, Schoones J, Toes RE, Huizinga TW, Kloppenburg M. Synovial inflammation, immune cells and their cytokines in osteoarthritis: a review. Osteoarthritis and cartilage. 2012 Dec 1;20(12):1484-99.
- [40] Melchiorri C, Meliconi R, Frizziero L, Silvestri T, Pulsatelli L, Mazzetti I, Borzi RM, Ugucioni M, Facchini A. Enhanced and coordinated in vivo expression of inflammatory cytokines and nitric oxide synthase by chondrocytes from patients with osteoarthritis. Arthritis & Rheumatism. 1998 Dec;41(12):2165-74.
- [41] Martin MU, Wesche H. Summary and comparison of the signaling mechanisms of the Toll/interleukin-1 receptor family. Biochimica et Biophysica Acta (BBA)-Molecular Cell Research. 2002 Nov 11;1592(3):265-80.
- [42] Kawai T, Akira S. TLR signaling. In Seminars in immunology 2007 Feb 1 (Vol. 19, No. 1, pp. 24-32). Academic Press.
- [43] Roman-Blas JA, Jimenez SA. NF- $\kappa$ B as a potential therapeutic target in osteoarthritis and rheumatoid arthritis. Osteoarthritis and cartilage. 2006 Sep 1;14(9):839-48.
- [44] B Marcu K, Otero M, Olivotto E, Maria Borzi R, B Goldring M. NF- $\kappa$ B signaling: multiple angles to target OA. Current drug targets. 2010 May 1;11(5):599-613.
- [45] Shakibaei M, Schulze-Tanzil G, John T, Mobasher A. Curcumin protects human chondrocytes from IL-1 $\beta$ -induced inhibition of collagen type II and  $\beta$ 1-integrin expression and activation of caspase-3: an immunomorphological study. Annals of Anatomy-Anatomischer Anzeiger. 2005 Nov 2;187(5-6):487-97.
- [46] Mengshol JA, Vincenti MP, Coon CI, Barchowsky A, Brinckerhoff CE. Interleukin-1 induction of collagenase 3 (matrix metalloproteinase 13) gene expression in chondrocytes requires

- p38, c-Jun N-terminal kinase, and nuclear factor  $\kappa$ B: differential regulation of collagenase 1 and collagenase 3. *Arthritis & Rheumatism: Official Journal of the American College of Rheumatology*. 2000 Apr;43(4):801-11.
- [47] Vincenti MP, Brinckerhoff CE. Transcriptional regulation of collagenase (MMP-1, MMP-13) genes in arthritis: integration of complex signaling pathways for the recruitment of gene-specific transcription factors. *Arthritis research & therapy*. 2002 Apr 1;4(3):157.
- [48] Verma P, Dalal K. ADAMTS-4 and ADAMTS-5: key enzymes in osteoarthritis. *Journal of cellular biochemistry*. 2011 Dec;112(12):3507-14.
- [49] Dayer JM, Beutler B, Cerami A. (1985). Cachectin/tumor necrosis factor stimulates collagenase and prostaglandin E2 production by human synovial cells and dermal fibroblasts. *J. Exp Med.*;162:2163-8.
- [50] Singh JA, Arayssi T, Duray P, Schumacher HR. Immunohistochemistry of normal human knee synovium: a quantitative study. *Annals of the rheumatic diseases*. 2004 Jul 1;63(7):785-90.
- [51] Kontny E, Maśliński W. Patogeneza reumatoidalnego zapalenia stawów. W: Wiland P (red.): *Reumatologia*. 2010:15-31.
- [52] Rose BJ, Kooyman DL. A tale of two joints: the role of matrix metalloproteases in cartilage biology. *Disease markers*. 2016 Oct;2016.
- [53] Hattori N, Mochizuki S, Kishi K, Nakajima T, Takaishi H, D'Armiento J, Okada Y. MMP-13 plays a role in keratinocyte migration, angiogenesis, and contraction in mouse skin wound healing. *The American journal of pathology*. 2009 Aug 1;175(2):533-46.
- [54] Shi J, Son MY, Yamada S, Szabova L, Kahan S, Chrysovergis K, Wolf L, Surmak A, Holmbeck K. Membrane-type MMPs enable extracellular matrix permissiveness and mesenchymal cell proliferation during embryogenesis. *Developmental biology*. 2008 Jan 1;313(1):196-209.
- [55] Chubinskaya S, Kuettner KE, Cole AA. Expression of matrix metalloproteinases in normal and damaged articular cartilage from human knee and ankle joints. *Laboratory investigation; a journal of*

- technical methods and pathology. 1999 Dec;79(12):1669-77.
- [56] Wu H, Du J, Zheng Q. Expression of MMP-1 in cartilage and synovium of experimentally induced rabbit ACLT traumatic osteoarthritis: immunohistochemical study. *Rheumatology International*. 2008 Nov 1;29(1):31.
- [57] Elliott S, Hays E, Mayor M, Sporn M, Vincenti M. The triterpenoid CDDO inhibits expression of matrix metalloproteinase-1, matrix metalloproteinase-13 and Bcl-3 in primary human chondrocytes. *Arthritis Res Ther*. 2003 Oct;5(5):1-7.
- [58] Salo T, Mäkelä M, Kylmäniemi M, Autio-Harminen H, Larjava H. Expression of matrix metalloproteinase-2 and-9 during early human wound healing. *Laboratory investigation; a journal of technical methods and pathology*. 1994 Feb;70(2):176.
- [59] Unemori EN, Bair MJ, Bauer EA, Amento EP. Stromelysin expression regulates collagenase activation in human fibroblasts. Dissociable control of two metalloproteinases by interferon-gamma. *Journal of Biological Chemistry*. 1991 Dec 5;266(34):23477-82.
- [60] Burrage PS, Mix KS, Brinckerhoff CE. Matrix metalloproteinases: role in arthritis. *Front Biosci*. 2006 Jan 1;11(1):529-43.
- [61] Shiomi T, Lemaître V, D'Armiento J, Okada Y. Matrix metalloproteinases, a disintegrin and metalloproteinases, and a disintegrin and metalloproteinases with thrombospondin motifs in non-neoplastic diseases. *Pathology international*. 2010 Jul;60(7):477-96.
- [62] Burrage PS, Mix KS, Brinckerhoff CE. Matrix metalloproteinases: role in arthritis. *Front Biosci*. 2006 Jan 1;11(1):529-43.
- [63] Xue M, March L, Sambrook PN, Jackson CJ. Differential regulation of matrix metalloproteinase 2 and matrix metalloproteinase 9 by activated protein C: relevance to inflammation in rheumatoid arthritis. *Arthritis & Rheumatism: Official Journal of the American College of Rheumatology*. 2007 Sep;56(9):2864-74.
- [64] Gearing AJ, Beckett P, Christodoulou M, Churchill M, Clements JD, Davidson AH, Drummond AH, Galloway WA, Gilbert R, Gordon JL, Leber TM. Processing of tumour

- necrosis factor- $\alpha$  precursor by metalloproteinases. *Nature*. 1994 Aug 18;370(6490):555-7.
- [65] Schönbeck U, Mach F, Libby P. Generation of biologically active IL-1 $\beta$  by matrix metalloproteinases: a novel caspase-1-independent pathway of IL-1 $\beta$  processing. *The Journal of Immunology*. 1998 Oct 1;161(7):3340-6.
- [66] Hot A, Miossec P. Effects of interleukin (IL)-17A and IL-17F in human rheumatoid arthritis synoviocytes. *Annals of the rheumatic diseases*. 2011 May 1;70(5):727-32.
- [67] Claveau D, Sirinyan M, Guay J, Gordon R, Chan CC, Bureau Y, Riendeau D, Mancini JA. Microsomal prostaglandin E synthase-1 is a major terminal synthase that is selectively up-regulated during cyclooxygenase-2-dependent prostaglandin E2 production in the rat adjuvant-induced arthritis model. *The Journal of Immunology*. 2003 May 1;170(9):4738-44.
- [68] Fattahi MJ, Mirshafiey A. Prostaglandins and rheumatoid arthritis. *Arthritis*. 2012;2012.
- [69] H. Sano, "The role of lipid mediators in the pathogenesis of rheumatoid arthritis," *Inflammation and Regeneration*, vol. 31, no. 2, pp. 151–156, 2011.
- [70] Abramovitz M, Adam M, Boie Y, Carrière MC, Denis D, Godbout C, Lamontagne S, Rochette C, Sawyer N, Tremblay NM, Belley M. The utilization of recombinant prostanoid receptors to determine the affinities and selectivities of prostaglandins and related analogs. *Biochimica et Biophysica Acta (BBA)-Molecular and Cell Biology of Lipids*. 2000 Jan 17;1483(2):285-93
- [71] Sugimoto Y, Narumiya S. Prostaglandin E receptors. *Journal of Biological Chemistry*. 2007 Apr 20;282(16):11613-7.
- [72] Miwa M, Saura R, Hirata S, Hayashi Y, Mizuno K, Itoh H. Induction of apoptosis in bovine articular chondrocyte by prostaglandin E2 through cAMP-dependent pathway. *Osteoarthritis and Cartilage*. 2000 Jan 1;8(1):17-24.
- [73] Attur M, Al-Mussawir HE, Patel J, Kitay A, Dave M, Palmer G, Pillinger MH, Abramson SB. Prostaglandin E2 exerts catabolic effects in osteoarthritis cartilage: evidence for signaling via the EP4 receptor. *The Journal of*

- Immunology. 2008 Oct 1;181(7):5082-8.
- [74] Otsuka S, Aoyama T, Furu M, Ito K, Jin Y, Nasu A, Fukiage K, Kohno Y, Maruyama T, Kanaji T, Nishiura A. PGE2 signal via EP2 receptors evoked by a selective agonist enhances regeneration of injured articular cartilage. *Osteoarthritis and cartilage*. 2009 Apr 1;17(4):529-38.
- [75] Aigner T, McKenna L, Zien A, Fan Z, Gebhard PM, Zimmer R. Gene expression profiling of serum-and interleukin-1 $\beta$ -stimulated primary human adult articular chondrocytes—A molecular analysis based on chondrocytes isolated from one donor. *Cytokine*. 2005 Aug 7;31(3):227-40.
- [76] Afonso V, Champy R, Mitrovic D, Collin P, Lomri A. Reactive oxygen species and superoxide dismutases: role in joint diseases. *Joint bone spine*. 2007 Jul 1;74(4):324-9.
- [77] Haskill S, Martin G, Van Le L, Morris J, Peace A, Bigler CF, Jaffe GJ, Hammerberg C, Sporn SA, Fong S. cDNA cloning of an intracellular form of the human interleukin 1 receptor antagonist associated with epithelium. *Proceedings of the National Academy of Sciences*. 1991 May 1;88(9):3681-5.
- [78] Gabay C, Porter B, Fantuzzi G, Arend WP. Mouse IL-1 receptor antagonist isoforms: complementary DNA cloning and protein expression of intracellular isoform and tissue distribution of secreted and intracellular IL-1 receptor antagonist in vivo. *The Journal of Immunology*. 1997 Dec 15;159(12):5905-13.
- [79] Dripps DJ, Brandhuber BJ, Thompson RC, Eisenberg SP. Interleukin-1 (IL-1) receptor antagonist binds to the 80-kDa IL-1 receptor but does not initiate IL-1 signal transduction. *Journal of Biological Chemistry*. 1991 Jun 5;266(16):10331-6.
- [80] Arend WP, Welgus HG, Thompson R, Eisenberg SP. Biological properties of recombinant human monocyte-derived interleukin 1 receptor antagonist. *The Journal of clinical investigation*. 1990 May 1;85(5):1694-7.
- [81] Horai R, Saijo S, Tanioka H, Nakae S, Sudo K, Okahara A, Ikuse T, Asano M, Iwakura Y. Development of chronic inflammatory arthropathy resembling rheumatoid arthritis in interleukin 1 receptor antagonist-deficient mice. *The Journal of experimental medicine*. 2000 Jan 17;191(2):313-20.

- [82] Fantuzzi G, Dinarello CA. Interleukin-18 and interleukin-1 $\beta$ : two cytokine substrates for ICE (caspase-1). *Journal of clinical immunology*. 1999 Jan 1;19(1):1-1.
- [83] Okamura H, Tsutsui H, Komatsu T, Yutsudo M, Hakura A, Tanimoto T, Torigoe K, Okura T, Nukada Y, Hattori K, Akita K. Cloning of a new cytokine that induces IFN- $\gamma$  production by T cells. *Nature*. 1995 Nov;378(6552):88-91.
- [84] Naik SM, Cannon G, Burbach GJ, Singh SR, Swerlick RA, Ansel JC, Caughman SW, Wilcox JN. Human keratinocytes constitutively express interleukin-18 and secrete biologically active interleukin-18 after treatment with pro-inflammatory mediators and dinitrochlorobenzene. *Journal of investigative dermatology*. 1999 Nov 1;113(5):766-72.
- [85] Gracie JA, Forsey RJ, Chan WL, Gilmour A, Leung BP, Greer MR, Kennedy K, Carter R, Wei XQ, Xu D, Field M. A proinflammatory role for IL-18 in rheumatoid arthritis. *The Journal of clinical investigation*. 1999 Nov 15;104(10):1393-401.
- [86] Udagawa N, Horwood NJ, Elliott J, Mackay A, Owens J, Okamura H, Kurimoto M, Chambers TJ, Martin TJ, Gillespie MT. Interleukin-18 (interferon- $\gamma$ -inducing factor) is produced by osteoblasts and acts via granulocyte/macrophage colony-stimulating factor and not via interferon- $\gamma$  to inhibit osteoclast formation. *The Journal of experimental medicine*. 1997 Mar 17;185(6):1005-12.
- [87] Olee T, Hashimoto S, Quach J, Lotz M. IL-18 is produced by articular chondrocytes and induces proinflammatory and catabolic responses. *The Journal of Immunology*. 1999 Jan 15;162(2):1096-100.
- [88] Okamura H, Tsutsui H, Komatsu T, Yutsudo M, Hakura A, Tanimoto T, Torigoe K, Okura T, Nukada Y, Hattori K, Akita K. Cloning of a new cytokine that induces IFN- $\gamma$  production by T cells. *Nature*. 1995 Nov;378(6552):88-91.
- [89] Takeda K, Tsutsui H, Yoshimoto T, Adachi O, Yoshida N, Kishimoto T, Okamura H, Nakanishi K, Akira S. Defective NK cell activity and Th1 response in IL-18-deficient mice. *Immunity*. 1998 Mar 1;8(3):383-90.
- [90] Dai SM, Shan ZZ, Nishioka K, Yudoh K. Implication of interleukin 18 in

- production of matrix metalloproteinases in articular chondrocytes in arthritis: direct effect on chondrocytes may not be pivotal. *Annals of the rheumatic diseases*. 2005 May 1;64(5):735-42.
- [91] Futani H, Okayama A, Matsui K, Kashiwamura S, Sasaki T, Hada T, Nakanishi K, Tateishi H, Maruo S, Okamura H. Relation between interleukin-18 and PGE2 in synovial fluid of osteoarthritis: a potential therapeutic target of cartilage degradation. *Journal of Immunotherapy*. 2002 Mar 1;25: S61-4.
- [92] Cho ML, Jung YO, Moon YM, Min SY, Yoon CH, Lee SH, Park SH, Cho CS, Jue DM, Kim HY. Interleukin-18 induces the production of vascular endothelial growth factor (VEGF) in rheumatoid arthritis synovial fibroblasts via AP-1-dependent pathways. *Immunology letters*. 2006 Mar 15;103(2):159-66.
- [93] Dinarello CA. An IL-1 family member requires caspase-1 processing and signals through the ST2 receptor. *Immunity*. 2005 Nov 1;23(5):461-2.
- [94] Chen J, Duan L, Xiong A, Zhang H, Zheng F, Tan Z, Gong F, Fang M. Blockade of IL-33 ameliorates Con A-induced hepatic injury by reducing NKT cell activation and IFN- $\gamma$  production in mice. *Journal of Molecular Medicine*. 2012 Dec 1;90(12):1505-15.
- [95] Bourgeois E, Van LP, Samson M, Diem S, Barra A, Roga S, Gombert JM, Schneider E, Dy M, Gourdy P, Girard JP. The pro-Th2 cytokine IL-33 directly interacts with invariant NKT and NK cells to induce IFN- $\gamma$  production. *European journal of immunology*. 2009 Apr;39(4):1046-55.
- [96] Na HJ, Hudson SA, Bochner BS. IL-33 enhances Siglec-8 mediated apoptosis of human eosinophils. *Cytokine*. 2012 Jan 1;57(1):169-74.
- [97] Cherry WB, Yoon J, Bartemes KR, Iijima K, Kita H. A novel IL-1 family cytokine, IL-33, potently activates human eosinophils. *Journal of allergy and clinical immunology*. 2008 Jun 1;121(6):1484-90.
- [98] Blom L, Poulsen BC, Jensen BM, Hansen A, Poulsen LK. IL-33 induces IL-9 production in human CD4<sup>+</sup> T cells and basophils. *PloS one*. 2011 Jul 6;6(7):e21695.
- [99] Schneider E, Petit-Bertron AF, Bricard R, Levasseur M, Ramadan A, Girard

- JP, Herbelin A, Dy M. IL-33 activates unprimed murine basophils directly in vitro and induces their in vivo expansion indirectly by promoting hematopoietic growth factor production. *The Journal of Immunology*. 2009 Sep 15;183(6):3591-7.
- [100] Ho LH, Ohno T, Oboki K, Kajiwara N, Suto H, Iikura M, Okayama Y, Akira S, Saito H, Galli SJ, Nakae S. IL-33 induces IL-13 production by mouse mast cells independently of IgE-FcεRI signals. *Journal of leukocyte biology*. 2007 Dec;82(6):1481-90.
- [101] Allakhverdi Z, Smith DE, Comeau MR, Delespesse G. Cutting edge: The ST2 ligand IL-33 potently activates and drives maturation of human mast cells. *The Journal of Immunology*. 2007 Aug 15;179(4):2051-4.
- [102] Funakoshi-Tago M, Tago K, Hayakawa M, Tominaga SI, Ohshio T, Sonoda Y, Kasahara T. TRAF6 is a critical signal transducer in IL-33 signaling pathway. *Cellular signalling*. 2008 Sep 1;20(9):1679-86.
- [103] Murphy GE, Xu D, Liew FY, McInnes IB. Role of interleukin 33 in human immunopathology. *Annals of the rheumatic diseases*. 2010 Jan 1;69(Suppl 1):i43-7.
- [104] Xu D, Jiang HR, Kewin P, Li Y, Mu R, Fraser AR, Pitman N, Kurowska-Stolarska M, McKenzie AN, McInnes IB, Liew FY. IL-33 exacerbates antigen-induced arthritis by activating mast cells. *Proceedings of the National Academy of Sciences*. 2008 Aug 5;105(31):10913-8.
- [105] Verri WA, Souto FO, Vieira SM, Almeida SC, Fukada SY, Xu D, Alves-Filho JC, Cunha TM, Guerrero AT, Mattos-Guimaraes RB, Oliveira FR. IL-33 induces neutrophil migration in rheumatoid arthritis and is a target of anti-TNF therapy. *Annals of the rheumatic diseases*. 2010 Sep 1;69(9):1697-703.
- [106] Gabay C, Towne JE. Regulation and function of interleukin-36 cytokines in homeostasis and pathological conditions. *Journal of leukocyte biology*. 2015 Apr;97(4):645-52.
- [107] Frey S, Derer A, Messbacher ME, Baeten DL, Bugatti S, Montecucco C, Schett G, Hueber AJ. The novel cytokine interleukin-36α is expressed in psoriatic and rheumatoid arthritis synovium. *Annals of the rheumatic diseases*. 2013 Sep 1;72(9):1569-74.

- [108] Boutet MA, Bart G, Penhoat M, Amiaud J, Brulin B, Charrier C, Morel F, Lecron JC, Rolli-Derkinderen M, Bourreille A, Vigne S. Distinct expression of interleukin (IL)-36 $\alpha$ ,  $\beta$  and  $\gamma$ , their antagonist IL-36Ra and IL-38 in psoriasis, rheumatoid arthritis and Crohn's disease. *Clinical & Experimental Immunology*. 2016 May;184(2):159-73.
- [109] Towne JE, Renshaw BR, Douangpanya J, Lipsky BP, Shen M, Gabel CA, Sims JE. Interleukin-36 (IL-36) ligands require processing for full agonist (IL-36 $\alpha$ , IL-36 $\beta$ , and IL-36 $\gamma$ ) or antagonist (IL-36Ra) activity. *Journal of Biological Chemistry*. 2011 Dec 9;286(49):42594-602.
- [110] Henry CM, Sullivan GP, Clancy DM, Afonina IS, Kulms D, Martin SJ. Neutrophil-derived proteases escalate inflammation through activation of IL-36 family cytokines. *Cell reports*. 2016 Feb 2;14(4):708-22.
- [111] Macleod T, Doble R, McGonagle D, Wasson CW, Alase A, Stacey M, Wittmann M. Neutrophil elastase-mediated proteolysis activates the anti-inflammatory cytokine IL-36 receptor antagonist. *Scientific reports*. 2016 Apr 22;6:24880.
- [112] Frey S, Derer A, Messbacher ME, Baeten DL, Bugatti S, Montecucco C, Schett G, Hueber AJ. The novel cytokine interleukin-36 $\alpha$  is expressed in psoriatic and rheumatoid arthritis synovium. *Annals of the rheumatic diseases*. 2013 Sep 1;72(9):1569-74.
- [113] Boutet MA, Bart G, Penhoat M, Amiaud J, Brulin B, Charrier C, Morel F, Lecron JC, Rolli-Derkinderen M, Bourreille A, Vigne S. Distinct expression of interleukin (IL)-36 $\alpha$ ,  $\beta$  and  $\gamma$ , their antagonist IL-36Ra and IL-38 in psoriasis, rheumatoid arthritis and Crohn's disease. *Clinical & Experimental Immunology*. 2016 May;184(2):159-73.
- [114] Magne D, Palmer G, Barton JL, Mézin F, Talabot-Ayer D, Bas S, Duffy T, Noger M, Guerne PA, Nicklin MJ, Gabay C. The new IL-1 family member IL-1F8 stimulates production of inflammatory mediators by synovial fibroblasts and articular chondrocytes. *Arthritis research & therapy*. 2006 Jun 1;8(3):R80.
- [115] Harusato A, Abo H, Ngo VL, Yi SW, Mitsutake K, Osuka S, Kohlmeier JE, Li JD, Gewirtz AT, Nusrat A, Denning TL. IL-36 $\gamma$  signaling controls the induced regulatory T cell–Th9 cell

- balance via NF $\kappa$ B activation and STAT transcription factors. *Mucosal immunology*. 2017 Nov;10(6):1455-67.
- [116] Chowdhury K, Kumar U, Das S, Chaudhuri J, Kumar P, Kanjilal M, Ghosh P, Sircar G, Basyal RK, Kanga U, Bandyopadhyaya S. Synovial IL-9 facilitates neutrophil survival, function and differentiation of Th17 cells in rheumatoid arthritis. *Arthritis research & therapy*. 2018 Dec;20(1):1-2.
- [117] Ciccica F, Guggino G, Rizzo A, Manzo A, Vitolo B, La Manna MP, Giardina G, Sireci G, Dieli F, Montecucco CM, Alessandro R. Potential involvement of IL-9 and Th9 cells in the pathogenesis of rheumatoid arthritis. *Rheumatology*. 2015 Dec 1;54(12):2264-72.
- [118] Vigne S, Palmer G, Lamacchia C, Martin P, Talabot-Ayer D, Rodriguez E, Ronchi F, Sallusto F, Dinh H, Sims JE, Gabay C. IL-36R ligands are potent regulators of dendritic and T cells. *Blood, The Journal of the American Society of Hematology*. 2011 Nov 24;118(22):5813-23.
- [119] Magne D, Palmer G, Barton JL, Mézin F, Talabot-Ayer D, Bas S, Duffy T, Noger M, Guerne PA, Nicklin MJ, Gabay C. The new IL-1 family member IL-1F8 stimulates production of inflammatory mediators by synovial fibroblasts and articular chondrocytes. *Arthritis research & therapy*. 2006 Jun 1;8(3):R80.
- [120] Woolley DE, Crossley MJ, Evanson JM. Collagenase at sites of cartilage erosion in the rheumatoid joint. *Arthritis & Rheumatism: Official Journal of the American College of Rheumatology*. 1977 Jul;20(6):1231-9.
- [121] Tetlow LC, Woolley DE. Mast cells, cytokines, and metalloproteinases at the rheumatoid lesion: dual immunolocalisation studies. *Annals of the Rheumatic Diseases*. 1995 Nov 1;54(11):896-903.
- [122] Trabandt A, Gay RE, Fassbender HG, Gay S. Cathepsin B in synovial cells at the site of joint destruction in rheumatoid arthritis. *Arthritis & Rheumatism*. 1991 Nov;34(11):1444-51.
- [123] Radovits BJ, Fransen J, Al Shamma S, Eijsbouts AM, Van Riel PL, Laan RF. Excess mortality emerges after 10 years in an inception cohort of early rheumatoid arthritis. *Arthritis care & research*. 2010 Mar;62(3):362-70.
- [124] Walsh NC, Reinwald S, Manning CA, Condon KW, Iwata K, Burr DB, Gravallesse EM. Osteoblast function is

- compromised at sites of focal bone erosion in inflammatory arthritis. *Journal of Bone and Mineral Research*. 2009 Sep;24(9):1572-85.
- [125] Kumar S, McDonnell PC, Lehr R, Tierney L, Tzimas MN, Griswold DE, Capper EA, Tal-Singer R, Wells GI, Doyle ML, Young PR. Identification and initial characterization of four novel members of the interleukin-1 family. *Journal of Biological Chemistry*. 2000 Apr 7;275(14):10308-14.
- [126] Nold MF, Nold-Petry CA, Zepp JA, Palmer BE, Bufler P, Dinarello CA. IL-37 is a fundamental inhibitor of innate immunity. *Nature immunology*. 2010 Nov;11(11):1014-22.
- [127] Boraschi D, Lucchesi D, Hainzl S, Leitner M, Maier E, Mangelberger D, Oostingh GJ, Pfaller T, Pixner C, Posselt G, Italiani P. IL-37: a new anti-inflammatory cytokine of the IL-1 family. *European cytokine network*. 2011 Nov 1;22(3):127-47.
- [128] Bulau AM, Nold MF, Li S, Nold-Petry CA, Fink M, Mansell A, Schwerd T, Hong J, Rubartelli A, Dinarello CA, Bufler P. Role of caspase-1 in nuclear translocation of IL-37, release of the cytokine, and IL-37 inhibition of innate immune responses. *Proceedings of the National Academy of Sciences*. 2014 Feb 18;111(7):2650-5.
- [129] Nold-Petry CA, Lo CY, Rudloff I, Elgass KD, Li S, Gantier MP, Lotz-Havla AS, Gersting SW, Cho SX, Lao JC, Ellisdon AM. IL-37 requires the receptors IL-18R $\alpha$  and IL-1R8 (SIGIRR) to carry out its multifaceted anti-inflammatory program upon innate signal transduction. *Nature immunology*. 2015 Apr;16(4):354.
- [130] Cavalli G, Koenders M, Kalabokis V, Kim J, Tan AC, Garlanda C, Mantovani A, Dagna L, Joosten LA, Dinarello CA. Treating experimental arthritis with the innate immune inhibitor interleukin-37 reduces joint and systemic inflammation. *Rheumatology*. 2016 Dec 1;55(12):2220-9.
- [131] Ye L, Jiang B, Deng J, Du J, Xiong W, Guan Y, Wen Z, Huang K, Huang Z. IL-37 alleviates rheumatoid arthritis by suppressing IL-17 and IL-17-triggering cytokine production and limiting th17 cell proliferation. *The Journal of Immunology*. 2015 Jun 1;194(11):5110-9.
- [132] Tang R, Yi J, Yang J, Chen Y, Luo W, Dong S, Fei J. Interleukin-37 inhibits

- osteoclastogenesis and alleviates inflammatory bone destruction. *Journal of cellular physiology*. 2019 May;234(5):7645-58.
- [133] Dinarello CA. Interleukin-18. *Methods*. 1999 Sep 1;19(1):121-32.
- [134] Jia H, Liu J, Han B. Reviews of interleukin-37: functions, receptors, and roles in diseases. *BioMed research international*. 2018 Jan 1;2018.
- [135] Burgler S, Ouaked N, Bassin C, Basinski TM, Mantel PY, Siegmund K, Meyer N, Akdis CA, Schmidt-Weber CB. Differentiation and functional analysis of human TH17 cells. *Journal of Allergy and Clinical Immunology*. 2009 Mar 1;123(3):588-95.
- [136] Harrington LE, Hatton RD, Mangan PR, Turner H, Murphy TL, Murphy KM, Weaver CT. Interleukin 17-producing CD4<sup>+</sup> effector T cells develop via a lineage distinct from the T helper type 1 and 2 lineages. *Nature immunology*. 2005 Nov;6(11):1123-32.
- [137] Sims JE, Smith DE. The IL-1 family: regulators of immunity. *Nature Reviews Immunology*. 2010 Feb;10(2):89-102.
- [138] Boutet MA, Najm A, Bart G, Brion R, Touchais S, Trichet V, Layrolle P, Gabay C, Palmer G, Blanchard F, Le Goff B. IL-38 overexpression induces anti-inflammatory effects in mice arthritis models and in human macrophages in vitro. *Annals of the rheumatic diseases*. 2017 Jul 1;76(7):1304-12.
- [139] Takenaka SI, Kaieda S, Kawayama T, Matsuoka M, Kaku Y, Kinoshita T, Sakazaki Y, Okamoto M, Tominaga M, Kanesaki K, Chiba A. IL-38: A new factor in rheumatoid arthritis. *Biochemistry and biophysics reports*. 2015 Dec 1;4:386-91.
- [140] Pasparakis M, Alexopoulou L, Episkopou V, Kollias G. Immune and inflammatory responses in TNF alpha-deficient mice: a critical requirement for TNF alpha in the formation of primary B cell follicles, follicular dendritic cell networks and germinal centers, and in the maturation of the humoral immune response. *The Journal of experimental medicine*. 1996 Oct 1;184(4):1397-411.
- [141] Mackay F, Loetscher H, Stueber D, Gehr G, Lesslauer W: Tumour necrosis factor alpha (TNF-alpha)-induced cell adhesion to human endothelial cells is under dominant control of one TNF receptor type, TNF-R55. *J Exp Med*. 1993;177:1277-86.

- [142] Bradley JR. TNF-mediated inflammatory disease. *The Journal of Pathology: A Journal of the Pathological Society of Great Britain and Ireland*. 2008 Jan;214(2):149-60.
- [143] Tracey K, Vlassara M, Cerami A. Cachectin/tumour necrosis factor. *Lancet (British edition)*. 1989(8647):1122-6.
- [144] Luettig B, Decker T, Lohmann-Matthes ML. Evidence for the existence of two forms of membrane tumor necrosis factor: an integral protein and a molecule attached to its receptor. *The Journal of Immunology*. 1989 Dec 15;143(12):4034-8.
- [145] Moss ML, Jin SL, Milla ME, Burkhart W, Carter HL, Chen WJ, Clay WC, Didsbury JR, Hassler D, Hoffman CR, Kost TA. Cloning of a disintegrin metalloproteinase that processes precursor tumour-necrosis factor- $\alpha$ . *Nature*. 1997 Feb;385(6618):733-6.
- [146] Dopp JM, Sarafian TA, Spinella FM, Kahn MA, Shau H, De Vellis J. Expression of the p75 TNF receptor is linked to TNF-induced NF $\kappa$ B translocation and oxyradical neutralization in glial cells. *Neurochemical research*. 2002 Nov 1;27(11):1535-42.
- [147] Grell M, Douni E, Wajant H, Löhdén M, Clauss M, Maxeiner B, Georgopoulos S, Lesslauer W, Kollias G, Pfizenmaier K, Scheurich P. The transmembrane form of tumor necrosis factor is the prime activating ligand of the 80 kDa tumor necrosis factor receptor. *Cell*. 1995 Dec 1;83(5):793-802.
- [148] Krippner-Heidenreich A, Tübing F, Bryde S, Willi S, Zimmermann G, Scheurich P. Control of receptor-induced signaling complex formation by the kinetics of ligand/receptor interaction. *Journal of Biological Chemistry*. 2002 Nov 15;277(46):44155-63.
- [149] Saxne T, Palladino Jr MA, Heinegard D, Talal N, Wollheim FA. Detection of tumor necrosis factor  $\alpha$  but not tumor necrosis factor  $\beta$  in rheumatoid arthritis synovial fluid and serum. *Arthritis & Rheumatism: Official Journal of the American College of Rheumatology*. 1988 Aug;31(8):1041-5.
- [150] Chu CQ, Field M, Allard S, Abney E, Feldmann M, Maini RN. Detection of cytokines at the cartilage/pannus junction in patients with rheumatoid arthritis: implications for the role of cytokines in cartilage destruction and

- repair. *Rheumatology*. 1992 Oct 1;31(10):653-61.
- [151] McInnes IB, Leung BP, Sturrock RD, Field M, Liew FY. Interleukin-15 mediates T cell-dependent regulation of tumor necrosis factor- $\alpha$  production in rheumatoid arthritis. *Nature medicine*. 1997 Feb 1;3(2):189-95.
- [152] Arend WP, Dayer JM. Inhibition of the production and effects of interleukin-1 and tumor necrosis factor alpha in rheumatoid arthritis. *Arthritis and rheumatism*. 1995 Feb;38(2):151.
- [153] Vasanthi P, Nalini G, Rajasekhar G. Role of tumor necrosis factor-alpha in rheumatoid arthritis: a review. *APLAR Journal of Rheumatology*. 2007 Dec;10(4):270-4.
- [154] Hess A, Axmann R, Rech J, Finzel S, Heindl C, Kreitz S, Sergeeva M, Saake M, Garcia M, Kollias G, Straub RH. Blockade of TNF- $\alpha$  rapidly inhibits pain responses in the central nervous system. *Proceedings of the National Academy of Sciences*. 2011 Mar 1;108(9):3731-6.
- [155] Williams RO, Feldmann M, Maini RN. Anti-tumor necrosis factor ameliorates joint disease in murine collagen-induced arthritis. *Proceedings of the National Academy of Sciences*. 1992 Oct 15; 89(20): 9784-8.
- [156] Lewis EJ, Bishop J, Bottomley KM, Bradshaw D, Brewster M, Broadhurst MJ, Brown PA, Budd JM, Elliott L, Greenham AK, Johnson WH. Ro 32-3555, an orally active collagenase inhibitor, prevents cartilage breakdown in vitro and in vivo. *British journal of pharmacology*. 1997 May; 121(3): 540-6.
- [157] Yoshihara Y, Nakamura H, Obata KI, Yamada H, Hayakawa T, Fujikawa K, Okada Y. Matrix metalloproteinases and tissue inhibitors of metalloproteinases in synovial fluids from patients with rheumatoid arthritis or osteoarthritis. *Annals of the rheumatic diseases*. 2000 Jun 1; 59(6): 455-61.