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**FORMULATION AND EVALUATION OF NANOPARTICLES USING CHITOSAN AS
POLYMERS AND *OCIMUM TENUIFLORUM* LEAF AND *ZINGIBER OFFICINALE*
STEM EXTRACTS**

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ABSTRACT

Aim and objective: The aim of the present study was to develop the nanoparticles consisting etoricoxib with ethanolic extract of ginger (*Zingiber officinale*, *Zingiberaceae*) root and tulsi (*Ocimum sanctum L*, *Lamiaceae*) leaves prepared nanoparticles using Chitosan as polymers treatment of diabetes complications.

Material and method: Nanoparticles were prepared by the antisolvent method by using a different concentration of polymer (Chitosan) and extract while concentration of etoricoxib was remain constant in all the formulations. The prepared nanoparticle was evaluated for different parameters.

Result & discussion: The formulation of nanoparticles was evaluated for various parameters such as viscosity, washability, consistency, pH, spreadability and in-vitro drug release study. The constancy of the formulations size of pores was found ranging from 924 nm was observed. The pH of the nanoparticles was found to be 6.1 ± 0.009 (F2) to 6.1 ± 0.098 (F7) whereas the viscosity was found in the range of 0.901 ± 0.010 (F7) to 1.200 ± 0.055 micm³ (F9). The Spreadability was found in the range of 61.9 ± 0.025 % (F5) to 71.01 ± 0.016 % (F4). The drug content of etoricoxib in the formulations was found in the range of 63.9 ± 0.11 (F8) to 77.5 ± 0.06 (F9). GE 58.9 ± 0.01

(F7) to 62.9 ± 0.05 (F1) and BE 35.9 ± 0.01 (F3) to 64.9 ± 0.05 (F9). The drug release was taken up to 24 h and found to be 99 % for all the formulations.

Conclusion: The results showed that the nanoparticles were prepared successfully.

Keywords: Nanoparticles, Chitosan Polymers, *Ocimum Tenuiflorum*, *Zingiber Officinale*

INTRODUCTION

Nanoparticles contain particular characteristics such the same as the smaller size of particles, a specific shape, and particle size distribution. They have a larger surface area than macro particles because of their small size (1-100 nm) the term is sometime used for larger particles up to 500 nm. [1]. According the equation of Noyes-Whitney the interfacial surface area is increased by reducing the particle size in the nano range therefore the solubility and rate of dissolution is poorly water-soluble drugs can be improved [2]. They contain potential to improve the therapeutic effects of drugs from side to side increasing the efficacy of the drug, lesser toxic effects of the drug, and attain the steady-state therapeutic level and also humanizing the solubility and drug stability [3]. The increase in the dissolution rate of the poorly water-soluble drug is essential to optimize the bioavailability [4].

Etoricoxib is a potent compound of COX-2 inhibitors. It inhibits the isoform-2 of the enzyme cox-2 and generates the prostaglandins from arachidonic acid. It is a BCS class II drug and used as an anti-

inflammatory agent in the treatment of many diseases [1]. When Etoricoxib administered orally, it shows severe side effects and low dissolution rate [5]. To avoid these problems, it requires solubility enhancement and site-specific drug delivery. So the site-specific topical carriers improve the efficiency of the drug through sustained release [1].

The antisolvent precipitation method is used to prepare the nano and micro particles. It is a bottom-up method which provides suitable procedures at a maintained temperature [6]. Under the sonication process, it is a rapid and low-cost method. The antisolvent solution is prepared by the polymer, surfactant and the mixture of both as a stabilizing agent then the solution of drug in an organic solvent is prepared and added to the antisolvent. This is important to maintain the conditions such as the rapid formation of particles and no growth of particle size for the preparation of nanoparticles [7]. The precipitation of nanoparticles is achieved by various steps such as mixing of solution and antisolvent, supersaturation, nucleation, and growth through condensation. Rapid and

supersaturation precipitation occurs through the main driving force. During the crystallization process, supersaturation generated and properties of crystal such as size, shape, purity depend on the uniformity and rate of supersaturation [6]. Nile *et al* studied the various effects of ginger (*Zingiber officinale*, Zingiberaceae) extract such as anti-inflammatory, antioxidant, and xanthine oxidase inhibitory activities. They determined the anti-inflammatory activity of ginger extract through diene conjugate, 13-glucuronidase inhibition, hyaluronidase inhibition and lipoxidase inhibition assay in vitro. They found that the 6-gingerol, 8-gingerol, 10-gingerol, 6-shadow, and 6-paradol are the active constituents of ginger which have various properties such as antioxidant, anti-inflammatory, anticancer activities [8]. Zlotek *et al* found that the basil leaves contains various properties such as anti-inflammatory, antioxidant, and anticancer activity [9]. Chitra *et al* developed the nanoparticles of the ginger extract by using poloxamer 188 as stabilizing agent through green synthesis and found that the zinger extract with poloxamer 188 incorporated gold nanoparticles is more effective than pure zinger extract [10]. In the present investigation, etoricoxib nanoparticles were prepared by an

antisolvent method by using Chitosan as polymers polysaccharide and phyto-constituents such as ethanolic extract of ginger root and tulsi leaves. Then the prepared nanoparticles were incorporated into the gel for enhancing the solubility of drug. The prepared nanoparticles incorporated gel was applied locally for targeting the skin to reduce the pain and inflammation associated with human skin melanoma cancer. The extract used in formulation contains antioxidant properties and potentiate the action of drug etoricoxib.

MATERIALS AND METHOD

Materials:

Ginger (Zinger *Zingiber officinale*, Zingiberaceae) root, tulsi (*Ocimum sanctum* L, Lamiaceae) leaves were collected from a local area of omicron 1A Greater Noida U.P.

Method:

Preparation of ginger rhizome and tulsi leaves extract:

Fresh plant leaves of *Ocimum sanctum* L. were collected from a local area of omicron 1A Greater Noida U.P. (28° 35' N, 77° 12' E). The leaves were washed thoroughly under tap water followed by sterile distilled water. Then leaves were dried under shaded condition at room temperature. Sampling of *Ocimum sanctum* L. was planted in the month of March. Middle aged fresh leaves of

Ocimum sanctum L. were plucked during the month of September- October in the morning between 9-10 a.m. (IST) when dew was less and temperature was also not so high and fresh ginger stem (*Zingiber officinale*) were procured from local market. The stem were washed with distilled water and dried in oven at 40 °C, about 5 to 10 min.

Extraction of plant material:

Dry ginger was crushed to a coarse powder and extracted with 95 % ethanol by simple

maceration process. Solvent was evaporated at room temperature. The residue obtained was dried. As well as the dried leaves material (in 20 gm) of *Ocimum sanctum* L. was extracted with 200 ml volumes of solvents, Ethanol, chloroform and n-butanol, separately at room temperature, in succession about 24 hours. The organic solvent was separated [11].

Preparation of nanoparticles (Table 1):

Table 1: Formulation of nanoparticles

| S. No. | Formulation | Ingredients | | |
|--------|-------------|-----------------|----------------|---------------|
| | | Polymer (µg/ml) | Ginger (mg/ml) | Tulsi (mg/ml) |
| 1. | F1 | 25 | 4.861 | - |
| 2. | F2 | 25 | - | 4.323 |
| 3. | F3 | 25 | 2.430 | 2.161 |
| 4. | F4 | 35 | 4.861 | - |
| 5. | F5 | 35 | - | 4.323 |
| 6. | F6 | 35 | 2.430 | 2.161 |
| 7. | F7 | 45 | 4.861 | - |
| 8. | F8 | 45 | - | 4.323 |
| 9. | F9 | 45 | 2.430 | 2.161 |

Preparation of Nanoparticles:

Preparation of polymer solution: 10 mg Chitosan as polymers was dissolved in solvent and make up the volume up to 10 ml. Then concentration of 25, 35, 45 µg/ml were prepared by mixing 1.25, 1.75 and 2.25 ml solution of polysaccharide in distilled water and make up the volume up to 50 ml. Drug (Etoricoxib) solution was prepared 2000 µg/ml by dissolving 20 mg of drug in acetone and make up the volume up to 10 ml. Solution of ginger and tulsi extract with concentration of 4.681 mg/ml and 4.333 mg/ml respectively,

was prepared in acetone. Then the mixture of extract was prepared by dissolving 2.340 mg/ml of ginger and 2.116 mg/ml of tulsi in acetone. For the formulation of F1, F4 & F7, 10 ml of polymer solution with concentration of polymer 25, 35 & 45 µg/ml respectively, were taken separately & mixed with the help of magnetic stirrer maintaining temperature at 37 °C with a constant stirring. Then 1ml ginger extract solution of concentration 4.681 mg/ml was mixed to the polymer solution drop wise. For the formulation of F2, F5 & F8, 10 ml polymer solution with concentration

of polymer 25, 35 & 45 µg/ml was taken and drop wise 1 ml tulsi extract solution of concentration 4.332 mg/ml was mixed to the polymer solution. For the formulation of F3, F6 & F9 the 10 ml polymer solution with concentration of polymer 25, 35 & 45 µg/ml was taken and drop wise 1 ml mixture of ginger and tulsi extract solution of concentration 2.340 and 2.116 mg/ml was mixed to the polymer solution. The temperature was brought down to the room temp. Then, 5 ml solution of etoricoxib concentration of 2000 µg/ml was added to the each solution with continuous stirring and wait till the acetone was completely evaporated.

Characterization of nanoparticles:

- 1. Physical appearance:** All the prepared batches of nanoparticles were checked for their clarity.
- 2. Particle size analysis:** The particle sizes of the nanoparticles were determined by using zeta analyzer.
- 3. Scanning electron microscope (SEM) analysis:** Surface properties of nanoparticles were studied by using SEM images. SEM study was carried out using zeis EVO analyzer at Amity University, Noida.
- 4. Drug Content:** Drug content was used to determine the amount of drug

present in nanoparticles. Formulation (50 mg) was taken and mixed with 10ml of phosphate buffer of pH 6.8 which resulted in the concentration of 100µg/ml. Now keep the solution for 24 h & after that again shake it. Then filter it to Whatman filter paper. The absorbance is taken in UV spectrophotometer at, max 230, 278 and 672 for etoricoxib, GE and TE, respectively. The drug content was calculated by using formula given in equation 1.

$$\text{Drug content} = \text{Absorbance} \times \text{Dilution factor} \dots \dots \dots \text{Equation 1}$$

RESULT AND DISCUSSION

The nanoparticles were developed by the antisolvent method. The solution of etoricoxib, GE and TE were prepared in organic solvent which is used as solvent in the formulation of nanoparticles. The solution of Chitosan as polymers was prepared in organic solvents which are used as antisolvent in the preparation of formulations. Hence, the nanoparticles were developed by the mixing of solvent, antisolvent, supersaturation and nucleation. The supersaturation for one-component crystals in liquids can be expressed using equation 3:

$$S = \frac{C_0}{C^*} \dots \dots \dots \text{Equation 2}$$

Where the C is the definite concentration of the active pharmaceutical ingredient in the solution (mol/L) and C^* is the equilibrium solubility (mol/L) of an active pharmaceutical ingredient in a mixture of organic solvent and antisolvent [12, 13]. It has been established that when the degree of supersaturation is high it leads to the higher nucleation rates and results in lower Gibbs free energy [12, 14, 15].

$$B^\circ \propto \exp\left(\frac{-\Delta G_{Cr}}{kT}\right) \dots \dots \dots \text{Equation 3}$$

Where, B° is the rate of nucleation, k is Boltzmann's constant, T is the absolute temperature and G , is the critical free energy. According to the theory of classical nucleation, in the absence of foreign particles and surfaces surroundings, the new solid phase is generated and termed as homogenous nucleation. If the nucleation promoted by the existing foreign particles, it is termed as heterogeneous nucleation. Both the nucleation is known as primary nucleation. The secondary nucleation is started through mechanical abrasion or thermodynamically effects of native crystals.

The free energy for the nucleation of homogeneous is expressed by the help of following equation as [16]:

$$\Delta G_{Cr} = \frac{16\pi\gamma^3 s l^{\nu 2}}{3(kT)^2 (ln(1+S))^2} \dots \dots \dots \text{Equation 4}$$

Hence, after combining the equation 4 and 5, the homogeneous nucleation rate in the solution can be expressed by using equation 6:

$$B^\circ = A_{hom} \exp\left(-\frac{16\pi\gamma^3 s l^{\nu 2}}{3(kT)^2 (ln(1+S))^2}\right) \dots \dots \dots \text{Equation 5}$$

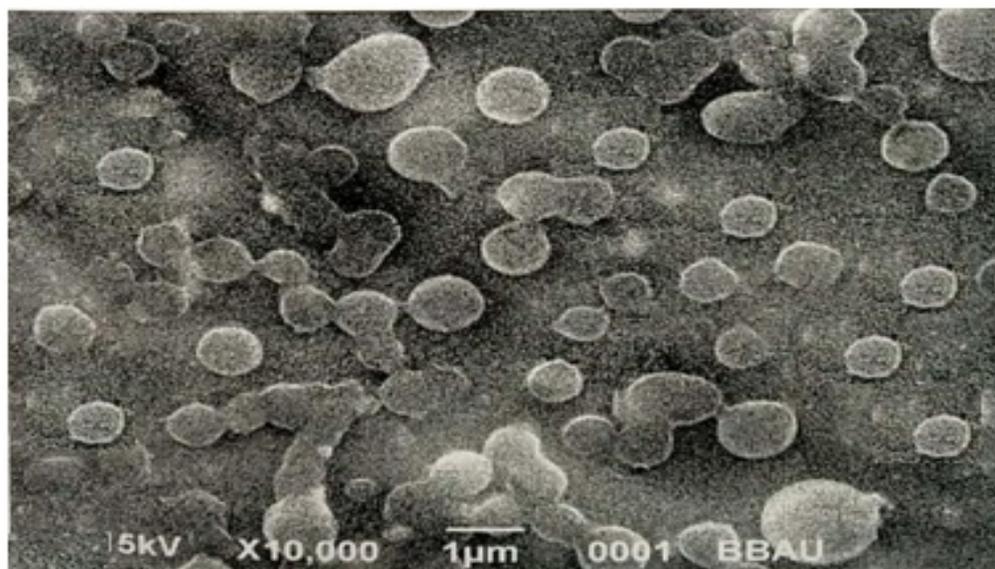
Where B° is the rate of nucleation, A_{hom} is the pre-exponential factor. At the solid-liquid interface, γ_{sl} is interfacial tension, $.l$) is molar volume, and T is temperature. A_{hom} is depending on the attached solute mechanism on growing particle surface, according to the Lindenberg and Mazzotti [17]. The magnitude order of A_{hom} usually varies from 10^{32} to 10^{36} . The rates of nucleation mainly depend on the interfacial energy (γ) and supersaturation seen from above equation [16].

In another research, Vasaya *et al* prepared etoricoxib nanosuspension with various stabilizers such as poloxamer and PVP for the enhancement of solubility and dissolution rate through high-pressure homogenization for the treatment of acute gouty arthritis. The results found that the solubility and dissolution rate were increased. Hence the bioavailability is also increased [5]. Kesharwani *at al* developed gel of etoricoxib for the treatment of arthritis. The solid lipid nanoparticles were prepared at low temperature through emulsification and solidification process by using stearic acid and tween 80. They compared the carbopol

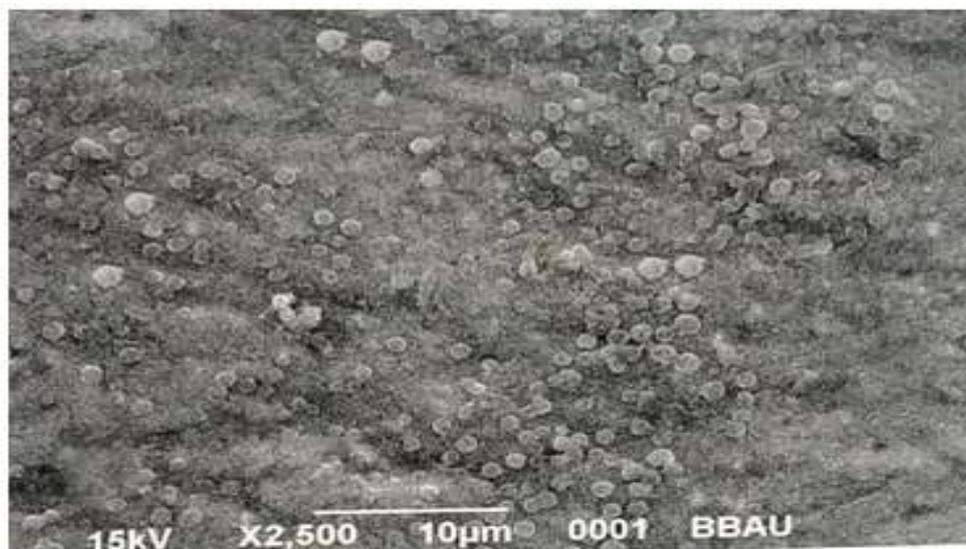
gel and hydroxyl propyl methyl cellulose (HPMC) gel and found that the carbopol gel was suitable for the treatment and showed good anti-inflammatory activity [20]. Bilthariya *et al* prepared nanoparticles of Etoricoxib by using bovine serum albumin with folic acid for the activation of macrophages on the folate receptor in the treatment of arthritis. The nanoparticles were prepared by desolvation method and the result showed that these nanoparticles have the potential to increase the therapeutic effect of drug [21].

In the present study, all the prepared batches of nanoparticles were checked for their clarity. All the formulations were found to be

clear and transparent. The particle sizes of the nanoparticles were determined by using zeta analyzer. 1% solution of the formulation was employed for the zeta analyzer. SEM images of nanoparticles are shown in **Figure 1**. The size of pores was found ranging from 924 nm (F1) to 1084 nm (F9). The drug content for the nanoparticles of etoricoxib varied from 66.8 ± 0.05 % (F4) to 85.1 ± 0.04 % (F9). The content of GE and BE were found in range from 60.3 ± 0.04 % (F4) to 72.1 ± 0.05 % (F9) and 59.5 ± 0.04 % (F8) to 80.5 ± 0.03 % (F9), respectively. The drug content of the nanoparticles is shown in **Table 2, Figure 2**.



(a)

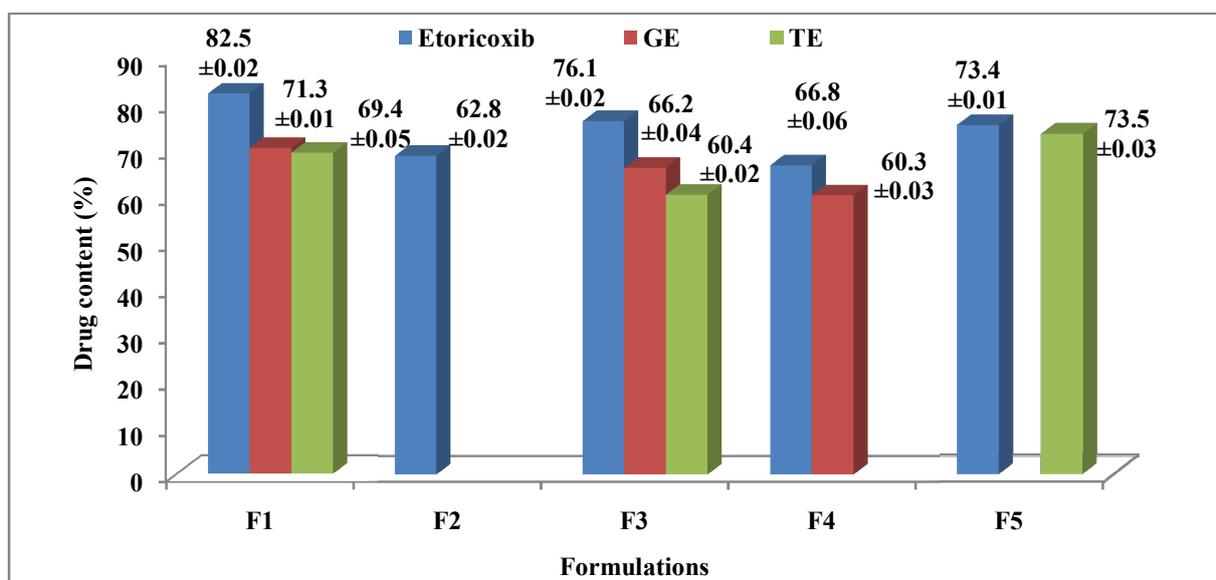


(b)

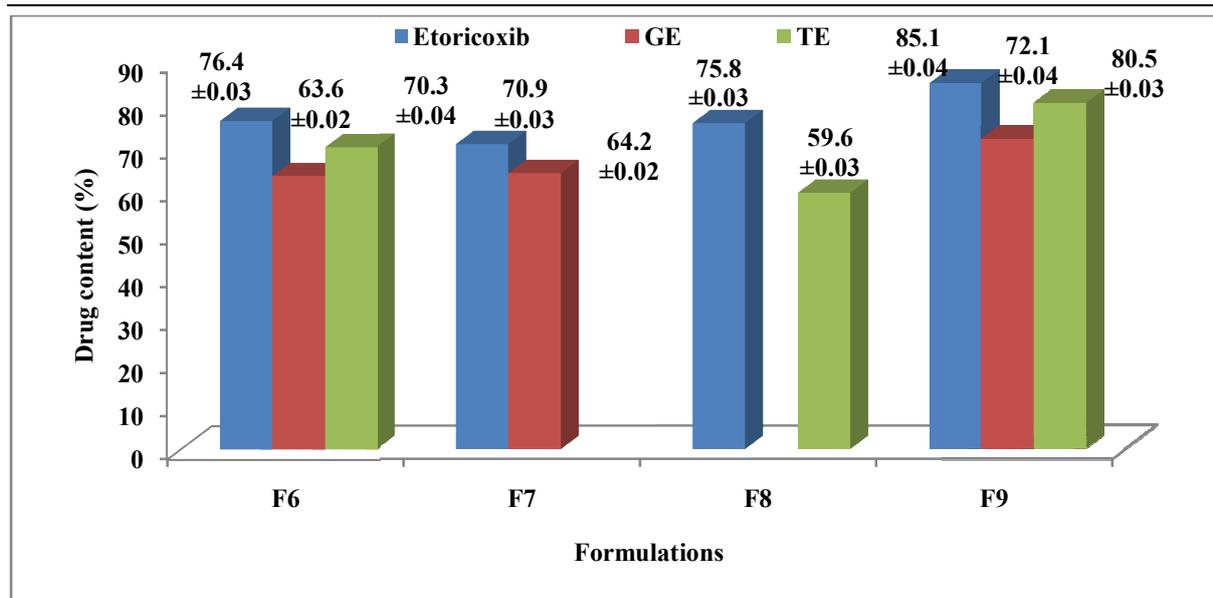
Figure 1: SEM analysis of F1 (a) X10,000 (b) X2,500

Table 2: The drug content of the nanoparticles

| Batches | Drug Content (%) | | | Particle size (nm) |
|---------|------------------|-----------|-----------|--------------------|
| | Etoricoxib | GE | TE | |
| F1 | 82.5±0.02 | 71.3±0.01 | 69.4±0.05 | 924 |
| F2 | 68.5±0.05 | - | - | 979 |
| F3 | 76.5±0.01 | 66.1±0.05 | 61.2±0.05 | 844 |
| F4 | 66.5±0.01 | 61.2±0.04 | - | 023 |
| F5 | 74.5±0.05 | - | 73.1±0.05 | 973 |
| F6 | 75.9±0.01 | 62.9±0.05 | 71.1±0.02 | 1031 |
| F7 | 74.9±0.01 | 62.2±0.05 | - | 1054 |
| F8 | 82.5±0.02 | 71.3±0.01 | 69.4±0.05 | 985 |
| F9 | 85.6±0.01 | 72.9±0.03 | 80.9±0.09 | 1084 |



(a)



(b)

Figure 2: The drug content of the nanoparticles

CONCLUSION

In this present work, Etoricoxib nanoparticles were prepared by using Chitosan as polymers and phytoconstituents such as ethanolic extract of Ginger (*Zingiber officinale*, *Zingiberaceae*) root and tulsi (*Ocimum sanctum*, *Lamiaceae*) leaves and for enhancing the solubility of drug and bioavailability as well. The prepared nanoparticles were applied for treatment of diabetes complications, reduce the pain and inflammation. The extract used in formulation contains antioxidant properties and used against the inflammation of the skin. The results concluded that the nanoparticles were more effective. Formulation reduces the pain without causing adverse effects because it avoids the first-pass metabolism of the drug and the

formulation is nongreasy in nature. It can be concluded that etoricoxib with phytoconstituents (GE and TE) improved its potential to control the growth of human melanoma cell line (SKMEL-2) in in-vitro conditions.

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