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**INVESTIGATION OF NOVEL FORMAZANS FOR ATTENUATION OF
SUBARACHNOID HEMORRHAGE INDUCED CEREBRAL VASOSPASM IN RATS**

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ABSTRACT

Cerebral vasospasm is delayed-onset cerebral arterial narrowing in response to blood clots left in the subarachnoid space after spontaneous aneurysmal subarachnoid hemorrhage (SAH). In the present study, we aimed to synthesize and investigate the cerebroprotective effects of formazans (quinazolinoneformazans – QF₁ and QF₂) for attenuation of subarachnoid hemorrhage induced cerebral vasospasm in rats. Subarachnoid hemorrhage (SAH) was performed by using a blood injection method. The neurological status of all animals was evaluated prior to surgery and 24hr post-surgery before euthanization and brain tissue harvesting. The brains were subjected for histopathological analysis. The cerebral vasospasm was noted measuring the diameter of major carotid artery (MCA) segment and the values from SAH and Sham animals were compared. All animals recovered from surgery and returned to their baseline activity with no motor weakness on spontaneous gait. There was no indication that the procedure caused any significant damage to the brain stem. Histological examination showed normal arterial wall features in control animals. Animals with SAH demonstrated significantly corrugated and thickened wall structure with decreased diameter on cross-section in SAH + vehicle group were significantly different from the sham group. The diameter was attenuated whereas the thickness was increased in the SAH + vehicle group as compared with the sham group. Simvastatin had shown great changes in these parameters while formazans exhibited mild changes in the parameters compared to SAH + Vehicle. From

the results obtained the current work demonstrated that the synthesized formazans (QF₁ and QF₂) does not exhibit motor weakness on spontaneous gait. From the histopathological study the formazans have limited significance in attenuation of SAH induced cerebral vasospasm.

Keywords: Subarachnoid Hemorrhage (SAH), Cerebral vasospasm, Simvastatin, Formazans, Major carotid artery

INTRODUCTION

Cerebral vasospasm is delayed-onset cerebral arterial narrowing in response to blood clots left in the subarachnoid space after spontaneous aneurysmal subarachnoid haemorrhage (SAH). High mortality rate due to subarachnoid haemorrhage in humans is mainly due to its complications like cerebral vasospasm, hyponatremia etc. Subarachnoid hemorrhage (SAH) accounts for 5% of strokes [1], it is associated with high mortality and severe morbidity. Although medical technology and treatment modality continues to advance, the prognosis of SAH patients is still poor. Traditionally, delayed cerebral vasospasm, which defined as a pathological demonstration occurred in the late phase of SAH (3-7 days). To date, tremendous research efforts have been made to reduce SAH-induced cerebral vasospasm [2-4]. Although positive results were obtained from animal experiments [5-7], however, anti-vasospasm treatments were failed to improve the SAH patient's outcome in most clinical trials [8]. These results from clinical trials make researchers raise doubt on the role of vasospasm, and attempt

finding new targets in treating patients suffering from SAH.

Among the wide variety of nitrogen heterocycles that has been explored for developing pharmaceutically important molecules the compounds bearing quinazoline nucleus has wide range of importance in medicinal chemistry, due to wide spectrum of biological activities exhibited by them. Quinazoline formazans have exhibited a variety of biological activities. Literature survey revealed that various substituted quinazolinones are known to possess antimicrobial [9-11], analgesic [12, 13], anti-inflammatory [14, 15], anthelmintic [16], anticonvulsant [17], anti-tubercular [18], anticancer [19], anti-HIV [20] properties. All these valid observations led us to investigate some quinazolinone formazan for attenuation of subarachnoid hemorrhage induced cerebral vasospasm in rats.

MATERIALS AND METHODS

Synthesis of formazans:

Two formazans were synthesized viz. 1-(4-Nitrophenyl)-3-(4-chlorophenyl)-4-[benzamido-(2-methyl-3-quinazolin)-4-one]formazan [QF₁] and 1-(3-Fluoro-4-

chlorophenyl)-3-(4-chlorophenyl)-4-[benzamido-(2-methyl-3-quinazolin)-4-one] formazan [QF₂] for the present study by the following steps (Scheme-I) [21-23]:

Step-1: Synthesis of 2-methyl-benzoxazin-4-one:

A mixture of equimolar quantities (0.1 mole) 2-amino benzoic acid (anthranilic acid) and acetic anhydride (25 ml) were refluxed for 2hrs and the reaction was monitored by TLC for the completion of reaction. The reaction mixture was poured in ice-cold water. The resulting mass was filtered, dried and recrystallized from DMSO.

Step- 2: Synthesis of 3-(4-carboxyphenyl)-2-methyl-3-quinazolin-4-one:

To a solution of 2-methyl-benzoxazin-4-one (0.01 mole) in alcohol para amino benzoic acid (PABA) was added, refluxed for 4 hrs and the reaction was monitored by TLC for the completion of reaction.

The resulting product is added to ice-cold water, filtered, dried and recrystallized from DMSO.

Step- 3: Synthesis of 3-(4-benzoylchloro)-2-methyl-3-quinazolin-4-one:

To the solution of 3-(4-carboxyphenyl)-2-methyl-3-quinazolin-4-one in alcohol (0.01 mole) double the molar concentration thionyl chloride (0.02 mole) is added. The resulting mixture was concentrated and solid residue formed at the bottom is

collected dried and recrystallized from DMSO.

Step- 4: Synthesis of 3-(4-hydrazinobenzoyl)-2-methyl-3-quinazolin-4-one:

To the solution of 3-(4-benzoylchloro)-2-methyl-3-quinazolin-4-one (0.01 mole) in alcohol double the molar concentration (0.02 mole) of hydrazine hydride is added, refluxed for 4 hrs. The resulting product is concentrated and the residue is collected, dried and recrystallized from DMSO.

Step- 5: Synthesis of 3-(4-arylazobenzamido)-2-methyl-3-quinazolin-4-one derivatives:

To a solution of 3-(4-hydrazinobenzoyl)-2-methyl-3-quinazolin-4-one (0.01 mole) in alcohol, aldehyde or substituted aldehyde (0.01mole) is added, stirred well to get the respective quinazolinone derivatives. The resulting product is filtered, dried and recrystallized.

Step- 6: Synthesis of 1-substituted phenyl-3-substituted phenyl-4-[benzamido (2-methyl-3-quinazolin)-4-one] formazans:

The diazonium salts derived from the respective amines (0.01mole) were added to aryl 3-(4-arylazobenzamido)-2-methyl-3-quinazolin-4-one derivatives in pyridine at 0-5°C with stirring for 30 minutes and the resultant mixture was added to ice-cold water. The resulting product is filtered, dried and recrystallized from DMSO.

Experimental animals and groups

All experimental protocols were approved by the Institutional animal ethics committee (Reg. No. 1048/PO/Re/S/07/CPCSEA: 18/IAEC/CLPT/2016-17) and were carried out as per the CPCSEA guidelines.

36 Adult male Sprague-Dawley rats (275-300 g) were taken and were randomly divided into four groups: Sham group (sham, n=6); Subarachnoid haemorrhage (SAH) treated with physiological saline as vehicle (SAH +vehicle n=6) ; SAH + QF₁ (n=6); SAH + QF₂ (n=6) and SAH treated with simvastatin which has been previously shown to be neuroprotective and in ameliorating cerebral vasospasm (SAH+ simvastatin, n=6). The neurological status of all animals was evaluated prior to surgery and 24hr post-surgery as earlier reported before euthanization and brain tissue harvesting and the brains were subjected for histopathological analysis.

Induction of SAH [24, 25]:

SAH was performed by using a blood injection method as described previously, with small modification. Animals were anesthetized with halothane (3% induction & 1-2% maintenance). The right femoral artery is exposed. The posterior scalp is incised at the midline and the skull exposed at the junction of the occipital bone. A 30-gauge needle is used to puncture the atlanto-occipital membrane 45° caudally, from the midline toward the left side of the

animal, into the cisterna magna. Blood (50–60µl) is withdrawn from the femoral artery and injected through the 30-gaugeneedle into the cisterna magna. The incision closed tightly with sutures to prevent cerebrospinal fluid (CSF) or blood leakage. The leg incision is closed with suture. The peak of the persistent phase of vascular narrowing is between 1 and 3 days after SAH; this is an appropriate time frame for measuring the effects of vascular constriction. All animals were kept in an air-conditioned room with temperature maintained at 27°C, and the rats were allowed access to food and water ad libitum before and after surgery.

Drug administration:

Thirty minutes after the procedure, SAH+simvastatin group received an intraperitoneal injection of simvastatin (10 mg/kg). Sham + vehicle group was administered with physiological saline. SAH+QF₁ and SAH + QF₂ received an intraperitoneal injection of synthetic quinazoline formazans respectively (100 mg/kg).

Neurological evaluation [26]:

Neurological symptoms were evaluated by direct observation of spontaneous activity after the recovery from anesthesia. The evaluation was recorded immediately upon recovery and at 24h postoperatively as either normal activity, weakness on gait with or without generalized weakness.

Assessment of cerebral vasospasm

After 24 hrs, animals were re-anesthetized, were then decapitated and the brains were carefully removed preserving the vasculature. The brains were subjected to histopathological study. The cerebral vasospasm was noted by measuring the diameter of the major carotid artery segment and the values from SAH and sham animals were compared.

Statistical analysis:

All statistical analysis was performed using Graphpad Prism 5. The data are expressed as mean \pm S.E.M. Statistical differences between the various groups were assessed with a one-way ANOVA. Differences in the means of arterial diameters were compared by using Dunnett's multiple comparisons test. Because diameters greater than the mean are not physiologically relevant, only the lower tail was considered and were used for vessel diameter comparisons. A value of $p < 0.001$ and $p < 0.0001$ vs SAH + vehicle was considered statistically significant.

RESULTS

All animals recovered from surgery and returned to their baseline activity with

nomotor weakness on spontaneous gait. Gross pathological examination of the meninges and external surface of the brain revealed diffuse blood prominently over the brain stem and the area of the Circle of Willis. There was no indication that the procedure caused any significant damage to the brain stem.

Histological examination showed normal arterial wall features in control animals. Animals with SAH demonstrated significantly corrugated and thickened wall structure with decreased diameter on cross-section (**Figure 1**) in SAH + vehicle group were significantly different from the sham group. The diameter was attenuated whereas the thickness was increased in the SAH + vehicle group ($214.8 \pm 7.44\mu\text{m}$) as compared with the sham group ($259.7 \pm 6.46\mu\text{m}$). Simvastatin had shown great changes in these parameters (262.8 ± 5.87 ; $p < 0.0001$ vs. SAH+Vehicle) while SAH + QF₁ and SAH + QF₂ groups (treated with the formazans) moderately differ in the parameter compared with the SAH group ($232.2 \pm 4.47 \mu\text{m}$; $229.8 \pm 5.05 \mu\text{m}$, $p < 0.001$ respectively) (**Table 1, Figure 2**).



Figure 1: Quantification of Vasospasm in Thickness and diameter (μm)

Table 1: Quantification of vasospasm (MCA diameter)

Treatment	MAC Diameter (μm)
Sham	259.7 ± 6.46
SAH + Vehicle	214.8 ± 7.44
SAH + Simvastatin	$262.8 \pm 5.87^{****}$
SAH + QF ₁	$232.2 \pm 4.47^{***}$
SAH + QF ₂	$229.8 \pm 5.05^{***}$

Note: Each value represents the mean \pm SEM (n = 6). *** $p < 0.001$ and **** $p < 0.0001$ compared with SAH + Vehicle group

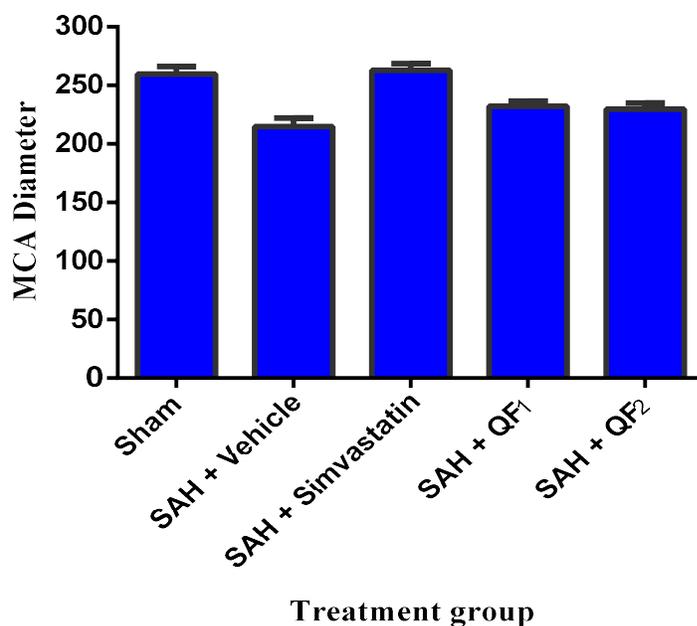


Figure 2: Quantification of vasospasm (MCA diameter)

DISCUSSION

Statins, are widely used clinically for their lipid-lowering properties [27], recent evidence shows that statins are also effective in ameliorating cerebral vasospasm, which occurs as a sequel of SAH [28]. Two randomized clinical trials investigating the use of statins for vasospasm after aneurysmal SAH showed that acute treatment with statins after SAH is safe and ameliorates vasospasm [29]. In the present study simvastatin significantly reversed the decrease in arterial diameter as well as the vessel wall thickening observed in the MCA affected by cerebral vasospasm at 24 hrs after SAH. Both Formazans (QF₁ and QF₂) moderately influence the cerebral vasospasm as indicated by morphological data (compared with sham animals).

CONCLUSION

From the results obtained the current work demonstrated that the synthesized formazans (QF₁ and QF₂) does not exhibit motor weakness on spontaneous gait. From the histopathological study the formazans have limited significance in attenuation of SAH induced cerebral vasospasm.

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CONFLICTS OF INTEREST

There are no Conflict of Interest.

REFERENCES

- [1] Chen S, Feng H, Sherchan P, Klebe D, Zhao G, Zhang J. “Controversies and evolving new mechanisms in subarachnoid hemorrhage” *Progress in Neurobiology*, 2014; 115: 64-91.
- [2] Velat GJ, Kimball MM, Mocco JD, Hoh BL. “Vasospasm after aneurismal subarachnoid hemorrhage: Review of randomized controlled trials and meta-analyses in the literature”, *World Neurosurgery*, 2011; 76(5): 446-454.
- [3] Golan E, Vasquez DN, Ferguson ND, Adhikari NKJ, Scales DC. “Prophylactic magnesium for improving neurologic outcome after aneurysmal subarachnoid hemorrhage: systematic review and meta-analysis”, *Journal of Critical Care*, 2013; 28(2): 173-181.
- [4] Junpeng Ma, Siqing Huang, Lu Ma, Yi Liu, Hao Li, Chao You “Endothelin-receptor antagonists for aneurismal subarachnoid hemorrhage: an updated meta-analysis of randomized controlled trials”, *Critical Care*, 2012;16: R198.
- [5] Gao Cheng, Liu Wei, Sun Zhi-dan, Zhao Shi-guang, Liu Xiang-zhen. “Atorvastatin ameliorates cerebral vasospasm and early brain injury after subarachnoid hemorrhage and inhibits caspase-dependent apoptosis pathway”, *BMC Neuroscience*, 2009; 10: 7.
- [6] Hong Y, Guo S, Chen S, Sun C, Zhang J, Sun X. “Beneficial effect of hydrogen-rich saline on cerebral vasospasm after experimental subarachnoid hemorrhage in rats”, *Journal of Neuroscience Research*, 2012; 90 (8): 1670-1680.
- [7] Sugawara T, Ayer R, Jadhav V, Chen W, Tsubokawa T, Zhang JH. “Simvastatin attenuation of cerebral vasospasm after subarachnoid hemorrhage in rats via increased phosphorylation of Akt and endothelial nitric oxide synthase”, *Journal of Neuroscience Research*, 2008; 86 (16): 3635-3643.
- [8] Wang X, Li YM, Li WQ, Huang CG, Lu YC, Hou LJ. “Effect of clazosentan in patients with aneurismal subarachnoid hemorrhage: a meta-analysis of randomized controlled trials”, *PLOS One*, 2012, 7(10): e47778.
- [9] Narendra Babu A, Rama Rao N “Synthesis and biological evaluation of some novel formazans”, *Journal of Pharmacy Research*, 2011; 4(1): 3-5.
- [10] Narendra Babu A, Rama Rao N “Synthesis and antimicrobial activity of 1-substituted phenyl-3-substituted phenyl-4-[(3,4,5-trimethoxy)-5-benzyl]-4-amino pyrimidine formazans”, *Asian Journal of Chemistry*, 2011; 23(1): 278-280.

- [11] Abdel Hamide SG “Synthesis and antimicrobial screening of some novel quinazolinones”, Journal of Indian Chemical Society, 1997; 74: 619-623.
- [12] Narendra Babu A, Rama Rao N, “Synthesis, Characterization and biological activity of some novel formazan derivatives”, Asian Journal of Chemistry, 2011; 23(3): 1349-1352.
- [13] Priyadarshini R, Rathinavel G “Synthesis and Pharmacological evaluation of thiazolyl and benzimido-quinazolines”, International Journal of Chemical Sciences, 2009; 7: 1099.
- [14] Narendra Babu A, Rama Rao N, et al “Synthesis of some new quinazolinone formazans as anti-Inflammatory and anthelmintic agents”, Journal of Pharmacy Research, 2011; 4(4):983-985.
- [15] Rajput CS, Ashok Kumar, Sudheer Kumar B, Jai Singh “Synthesis and anti-inflammatory activity of 2-[5'-(4-pyridinyl)-1',2',3'-oxadiazol-2-ylthiomethyl]-3-substituted-aryl-6-substituted-quinazolin-4-ones”, Asian Journal of Chemistry, 2008; 20(8): 6246-6252.
- [16] Rama Rao N, Mukkanti K, Sudhakar Rao G, Narendra Babu A, “Microwave Synthesis of some new quinazoline formazans for their antimicrobial and anthelmintic activities”, Current Trends in Biotechnology and Pharmacy, 2010; 4(1): 545-550.
- [17] Desai AR, Desai KR “Niementowski reaction: Microwave induced and conventional synthesis of quinazolinone and 3-methyl-1H-5-pyrazolone and their antimicrobial activity”, Arkivoc, 2005; XIII: 98.
- [18] Shirodkar PY, Meghana MV, et al “Synthesis and screening of some novel mannich bases of 6-nitro-N-arylamino methyl-1,2,3,4-tetrahydro-4-oxo-2-thio quinazolines for their anti-tubercular activity”, Indian Journal of Heterocyclic Chemistry, 2000; 9: 239-240.
- [19] Murugan, V, Thomas CC, Rama Sarma GVS, Kumar EP, Suresh B. “Synthesis of 2-Substituted Quinazolin-4 (3H)-Ones as a new class of anticancer agents”, Indian Journal of Pharmaceutical Sciences, 2003; 65(4): 386-389.
- [20] Pathak AV, Venkatesh Perumal US, Meena RS, Thirumurugan K. “Anti HIV and antibacterial activities of 2-substituted thiadiazoloquinazolines”, Indian Journal of Pharmaceutical Sciences, 2003; 65(3): 293-295.
- [21] Pandey VK, Pathak LP, Mishra SK. “Synthesis and characterisation of isoquinolinyl quinazolines and a study of their antiviral and antifungal

- activities”, Indian Journal of Chemistry, 2005; 44 B (9): 1940-1943.
- [22] VarshaJatav, Jain SK, Kashaw SK, Mishra P. “Synthesis and antimicrobial activity of novel 2-Methyl-3-(1'3'4'-Thiadiazoyl)-4-(3H) Quinazolinones”, Indian Journal of Pharmaceutical Sciences, 2006; 68 (3): 360-363.
- [23] SalehMd A, Abdel-Megeed Md F, Abdel Md A, Shokr ABM. “Synthesis of novel 3H-Quinazolin-4-ones containing Pyrazolinone, Pyrazole and Pyrimidinonemoieties”, Molecules, 2003; 8 (4): 363-373.
- [24] Lin CL, Calisaneller T, Ukita N, Yoshimura R, Wang Y, Lee KS. “Animal Models for acute neurological Injuries”, Humana Press (Springer), USA 2003;pp. 287-292.
- [25] Shao A, Guo S, Tu S, Ammar A, Tang J, Hong Y. Wu, H. Zhang, J. “Astragaloside IV Alleviates Early Brain Injury Following Experimental Subarachnoid Hemorrhage in Rats”, International Journal of Medical Sciences, 2014; 11(10): 1073-1081
- [26] Lin CL, Calisaneller T, Ukita N, Dumont AS, Kassell NF, Lee KS. “A murine model of subarachnoid hemorrhage-induced cerebral vasospasm”, Journal of Neuroscience Methods, 2003; 123(1):89–97.
- [27] Prager R. “Treatment of hyperlipidemia with HMG-CoA reductase inhibitors”, Wiener Medizinische Wochenschrift, Suppl, 1989; 105: 17–20.
- [28] McGirt MJ, Robert Blessing, Alexander MJ, Nimjee SM, Woodworth GF, Friedman AH, Graffagnino C, Laskowitz DT, Lynch JR “Risk of cerebral vasospasm after subarachnoid hemorrhage reduced by statin therapy: a multivariate analysis of an institutional experience”, Journal of Neurosurgery, 2006, 105 (5) :671–674.
- [29] Lynch JR, Wang H, McGirt MJ, James Floyd, Friedman AH, Coon AL, Robert Blessing, Alexander MJ, Graffagnino C, Warner DS, Laskowitz DT. “Simvastatin reduces vasospasm after aneurysmal subarachnoid hemorrhage: results of a pilot randomized clinical trial”, Stroke, 2005; 36 (9): 2024–2026.