IJBPAS, February, 2020, 9(2): 80-93

ISSN: 2277–4998

International Journal of Biology, Pharmacy and Allied Sciences (IJBPAS)

'A Bridge Between Laboratory and Reader'

www.ijbpas.com

PHARMACOGNOSTICAL STUDY AND PHYTOCHEMICAL EVALUATION OF THE LEAVES OF *ARTABOTRYS ODORATISSIMUS (ROXB)R. BR (Roxb) R. Br*

SARITHA K[1*] AND MURALI R[2], SRINIVASAN N[3]

1: Asst. Prof., Department of Pharmacognosy, Research scholar, Annamalai University, Tamil Nadu, India,

2: Asst. Prof., Department of Pharmacy, Annamalai University, Tamil Nadu, India,

3: Asst. Prof., Department of Pharmacy, Annamalai University, Tamil Nadu, India,

***Corresponding Author: E Mail: saritha.kodithala@gmail.com**

Received 17th Aug. 2019; Revised 6th Sept. 2019; Accepted 2nd Sept. 2019; Available online 1st Feb. 2020

https://doi.org/10.31032/IJBPAS/2020/9.2.4940

ABSTRACT

Plants are the important sources for several drugs. In recent years many drug formulations are based on plant products. In the present work, pharmacognostic studies of *Artabotrys odoratissimus (Roxb) R.Br (Roxb) R.Br* was attempted which included physicochemical, phytochemical, macroscopic, microscopic studies and organoleptic evaluation. The plant powder characteristics were also elucidated. The physicochemical examinations were finished by utilizing WHO prescribed parameters, for example loss on drying, ash values (total ash, water soluble ash, acid insoluble ash etc.), and extractive qualities. The subjective phytochemical investigation uncovered the presence of different secondary metabolites. The morphological studies exhibited the organoleptic and surface characteristics of the plant and its parts. The microscopic study showed the presence of various characteristics of whole plant. The pharmacognostic characters enrolled in this investigation will help in identification of the crude drug; the standardization parameters laid down will ensure the efficacy of drug and also distinguish the drug from its adulterants. The distinctive characters will likewise be useful for the preparation of monograph of this plant.

Keywords: WHO, extraction, organoleptic, microscopy, physico-chemical

INTRODUCTION

Medicinal plants are playing most active role in traditional medicines for the treatment of various diseases[1-7]. However a key obstacle, which has hindered the promotion in use of alternative medicines in the developed countries, is no evidence of documentation and absence of stringent quality control measures. There is a need for the record of all the research work carried out on traditional medicines in the form of documentation. With this drawback, it becomes extremely important to make surety about the standardization of the plant and parts of plant to be used as a medicine. For the process of standardization, we can use different techniques and methodology to achieve our goal in the stepwise manner *e.g.* pharmacognostic and phytochemical studies. These steps and processes are helpful in identification and standardization of the plant material. Correct characterization and quality assurance of starting material is an essential step to ensure reproducible quality of herbal medicine which will help us to justify its safety and efficacy[8-11]. For this purpose we have done pharmacognostic studies of *Artabotrys odoratissimus (Roxb) R.Br (Roxb) R.Br* (Annonaceae).Artabotrys odoratissimus (Roxb) R.Br R.Br. is a shrub native to eastern Asia and belongs to Annonaceae family[12].

This plant is commonly known as Manoranjini (Malayalam), Manoranjitham (kannada), Harchampa (Hindi), Kalchampa (Beng), Lilochampa (Guj), and Harachampaka (San). It is a straggling shrub, leaves oblong - lanceolate, 6-10 x 2-4 cm long, acuminate, cuneate at base, shining above. Flowers fragrant, solitary or few flowered, peduncle. Sepals 3, recurved. Petals 6 in two whorls of 3 each, clawed at base, green at first turning yellow. Berries yellow 6-10 in a cluster. Flowering and fruiting occur most part of the year. It is indigenous to Indian Peninsula and Srilanka[13]

Medicinal properties and uses

Artabotrys odoratissimus (Roxb) R.Br is an ornamental plant[14]. Fruits of this plant are recorded as containing fixed and volatile oil glycosides and resins, extracts are reported to exhibit hypotensive and spasmogenic as well as cardiac stimulating effects on some animals and cardiac depressant on others[15]. Decoctions of the leaves are used as a remedy for cholera and have been found to exhibit antifertility effects in rats[16-20]**.** The essential oil of *A. odoratissimus* has shown excellent to good antihelmintic property against tape worms, earth worms and round worms. Its flowers are used in the treatment of vomiting, biliousness, blood and heart diseases, itching, sweating, foul breath, thirst and headache[21]. The plant leaf of

Artabotrys odoratissimus (Roxb)R.Br have a major contribution in the treatment of Cardiac stimulant, uterine stimulant, muscle relaxant[22].

MATERIALS AND METHODS

Collection of plant material

The plant *Artabotrys odoratissimus* belonging to the family Annonaceae were collected from Chittoor district of Andhra Pradesh and was identified and authentified by Dr. Madhavachetty, plant taxonomist, Asst. Prof, Dept. of Botany. The plant voucher No. is 0823 dated 17-12- 2018.

Scientific Classification

SCIENTIFIC CLASSIFICATION	
Botanical Name	***Artabotrys odoratissimus (Roxb)R.Br***
Kingdom	**Plantae**
clade	**Angiosperms**
Order	**Magnoliales**
Family	**Annonaceae**
Genus	***Artabotrys***
Species	***Artabotrys odoratissimus (Roxb)R.Br***

Instruments and Chemicals

Compound microscope, stage micrometer, camera lucida, drawing sheets, glass slides, cover slips, watch glass and other common glassware were the basic apparatus and instruments used for the study. Microphotographs were taken using a Leica (Dissecting Microscope Lighting System-DMLS) microscope attached with Leitz (Magnification Power System- MPS) 32 camera. Solvents viz., 95% Methanol (MeOH), Petroleum ether (PE), chloroform and reagents such as phloroglucinol, glycerin, hydrochloric acid, chloral hydrate, and sodium hydroxide were procured from Merck Specialities Pvt. Ltd., Balanagar, Hyderabad, Telangana.

Macroscopic and Organoleptic Characterization

The macroscopic and organoleptic measurement were performed for important characters of fresh leaves like phyllotaxy, size, shape, colour, venation, presence or absence of petiole, apex, margin, base, lamina, texture, surface, odour and taste[23-24].

Microscopic Characterization of the leaf

Free hand transverse sections of leaf lamina and midrib were prepared, stained with safranin, mounted on glass slides using glycerine and observed under light microscope by reported methods[16, 17]. Photomicrographs of the microscopical sections were taken with the help of Digital Microscope (MOTIC) provided with MOTIC IMAGE PLUS 2.0 software[25-26].

Determination of Stomatal Number and Stomatal Index

A small piece of leaf was cleaned by boiling with sodium hypochlorite solution. The upper and lower epidermis was peeled separately. The peeled epidermis was

placed on slide and mounted with glycerine water. Average number of stomata per mm2 of the epidermis of the leaf (Stomatal number) is calculated from the microphotographs taken using camera attached microscope[18,19].Values for upper and lower epidermis were determined separately using the equation:

Stomatal index (SI) = S × 100 /E+S.

Where, S= the number of stomata per unit area and E = the number of epidermal cells in the same unit area of leaf[27-28].

Determination of Vein-Islet Number and Vein-Let Termination Number

Vein islet is the minute area of photosynthetic tissue encircled by the ultimate division of the conducting strands. Vein termination number is the number of veinlet terminations per mm^2 of leaf surface. A piece of the leaf was cleared by boiling in chloral hydrate solution and camera lucida and drawings board were arranged and 1 mm. line was drawn with help of stage mm. A square was constructed on this line in the centre of the field. The slide was placed on the stage. The veins included within the square were traced off, completing the outline of those islets which overlap two adjustment side of the square. The average number of vein islet from the four adjoining square, to get the value for one square mm was calculated[20]. The number of veinlet termination present within the square was counted and the average number of veinlet termination number from the four adjoining square to get the value for 1 square mm was found known as vein termination number[29].

Determination of Palisade Ratio

A piece of the leaf was boiled in chloral hydrate and was placed under microscope. Camera lucida and drawing board were arranged and the outline of four cells of the epidermis was traced using 4 mm objective. Then, palisade layer was focused down and sufficient cells for covering the tracing of the epidermal cells were traced off. The outline of those palisade cells which were intersected by the epidermal walls was completed. The palisade cells under the four epidermal cells (including cells which are more than half and excluding cells which are less than half within the area of epidermal cells) were counted. The determination for five groups of four epidermal cells from different part of the leaf was repeated. The average number of cells beneath epidermal cells was calculated known as palisade ratio[30].

Extraction

The authenticated plant materials were subjected for soxhlet extraction in which the solvent vapour generated by gently heating the reservoir condenses and is allowed to drip back onto the porous sample cup. The liquid condensate that

drips onto the sample performs the extraction which then passes through the container and back into the reservoir. The cycle is repeated continuously and can be sustained as long as needed.The dried materials were coarsely powdered; the powder was packed in filter paper and loaded into the thimble of soxhlet extractor. The solvent was used for extraction was poured into flask. The soxhlet extraction was carried out for 72 hours. Later the extracted solvent was evaporated under reduced pressure to get waxy extract.

Determination of Extractive Values

5 g of air dried coarse powder of extracts macerated with100 ml. of solvent (petroleum ether, chloroform, alcohol and water) in a glass-stoppered conical flask with frequent shaking for 6 hours and then allowed to stand for 18hours. There after it was filtered rapidly taking care against loss of solvent. About 25ml. of the filtrate was evaporated in a flat-bottomed dish to dryness using water bath and then dried at 105^0C for 6 hours, cooled in a desiccator for 30 minutes and weighed immediately[31].

Physicochemical Analysis

The fresh leaves were separated from the collected plants, thoroughly washed with fresh water, shade dried and powdered. The physicochemical parameters like ash value, acid-insoluble ash value, water soluble ash value, moisture content, foaming index and swelling index of *Artabotrys odoratissimus (Roxb)R.Br* leaves were determined according to the quality control methods for medicinal plant materials[32-33].

Determination of Ash

The ash remaining following ignition of medicinal plant materials is determined by three different methods which measure total ash, acid-insoluble ash and water-soluble ash. The total ash method is designed to measure the total amount of material remaining after ignition. This includes both "physiological ash", which is derived from the plant tissue itself, and "non-physiological" ash, which is the residue of the extraneous matter (e.g. sand and soil) adhering to the plant surface. Acid-insoluble ash is the residue obtained after boiling the total ash with dilute hydrochloric acid, and igniting the remaining insoluble matter. This measures the amount of silica present, especially as sand and siliceous earth. Water-soluble ash is the difference in weight between the total ash and the residue after treatment of the total ash with water[34].

Total Ash value

The method for determination of total ash is designed to measure the total amount of material remaining after ignition. This includes both “physiological ash”, which is derived from the plant tissue itself, and “non-physiological ash”, which is residue of extraneous matter (e.g. sand and soil) adhering to the plant surface. When

vegetable drugs are incinerated, they leave an inorganic ash in some plants called the total ash.

Four grams of the ground air-dried sample was weighed into previously ignited, dried and tarred silica crucible. The material was spread evenly as a thin layer. Kept on a gas burner under a low flame and ignited slowly to obtain a carbonized residue. It was then placed in the muffle furnace and the temperature of the muffle was adjusted to 450-500 0C and heated for 3 hours, cooled in a desiccator and weighed[35].

Determination of Acid Insoluble Ash

Acid insoluble ash is the residue obtained after boiling the total ash with dilute hydrochloric acid, and igniting the remaining insoluble matter. This measures the amount of silica present, especially as sand and siliceous earth. Total ash treated with dilute hydrochloric acid reacts with minerals to form soluble salts and the residue which consists mainly of silica is the acid insoluble ash.

To the Silica crucible containing the total ash obtained 25 ml of hydrochloric acid was added, covered with a watch glass and boiled gently for 5 minutes on a hot plate. The watch glass was rinsed with 5 ml of hot water and these washings added to the crucible and filtered. The insoluble matter was collected on an ashless filter paper by filtration. The filter paper was rinsed repeatedly with hot water until the filtrate was neutral /free from acid. The filter paper containing the insoluble matter was transferred to the original crucible, dried on a hot plate and ignited to a constant weight in the muffle furnace at 450-500 ºC. The silica crucible was removed from the muffle furnace and allowed to cool in a desiccator for 30 minutes, and then weighed without delay[36].

Determination of Water-Soluble Ash

To the crucible containing the total ash, add 25 ml of water and boil for 5 minutes. Collect the insoluble matter in a sintered-glass crucible or on an ashless filter-paper. Wash with hot water and ignite in a crucible for 15 minutes at a temperature not exceeding 450°C. Subtract the weight of this residue in mg from the weight of total ash. Calculate the content of water-soluble ash in mg per g of air-dried material[37].

Determination of Foaming Index

Many medicinal plant materials contain saponins that can cause persistent foam when an aqueous decoction is shaken. The foaming ability of an aqueous decoction of plant materials and their extracts is measured in terms of a foaming index. Finely divided (sieve No. 1250) plant powder 1g was kept into a 500 ml flask containing 100 mL of boiling water for 30 min. Then cooled and filtered into a 100ml volumetric flask and sufficient water was added to make up the volume. The prepared

decoction was transferred into 10 stoppered test tubes each 1 mL, 2 ml up to 10 ml. The volume of the liquid in each tube was adjusted to 10 mL with water. The tubes were duly stoppered and shaken in a lengthwise motion for 15 sec (two shakes per second) and allowed to stand for 15 min. The foam height in each tube was measured[38].

Determination of Swelling Index

Many medicinal plant materials are of specific therapeutic or pharmaceutical utility because of their swelling properties, especially gums and those containing an appreciable amount of mucilage, pectin or hemicellulose. Leaf powder (1g) was taken in a measuring cylinder (25 mL) and suspended in 25 mL distilled water for 1h by thorough mixing every 10 min. After 3 h, volume in mL occupied by the plant material including any sticky mucilage was measured. The experiment was repeated three times for accuracy and the swelling index was calculated[39].

Preliminary Phytochemical Investigations

The qualitative chemical tests carried out for the identification of the different nature phytoconstituents present in the powered crude drugs by standard procedures. They are usually tested for the presence of alkaloids, flavonoids, phenols, tannins, cardiac glycosides, triterpenes, steroids and saponins[40-41].

Florescence Analysis

A small quantity of dry leaf powder is placed on oil free clean microscopic slide and 1-2 drops of freshly prepared reagent solution is added, mixed by gentle tilting the slide and wait for few minutes. Then the slide is kept inside the UV chamber and observe the colour in visible light, short (254 nm) and long (365 nm) ultra violet radiations. The color observed by application of different reagents in different radiations is recorded[42-43].

Some constituents exhibit fluorescence in the visible range in daylight. The ultra violet light produces fluorescence in many natural products which do not visibly fluoresce in daylight. If substance themselves are not fluorescent, they may usually convert into fluorescent derivatives or decomposition products by applying different reagents. Hence crude drugs are often assessed qualitatively in this way and it is an important factor for pharmacognostic evaluation of crude drugs[44].

RESULTS AND DISCUSSION

Macroscopic and Organoleptic Characterization

Macroscopic and organoleptic characters of the fresh leaves of *Artabotrys odoratissimus (Roxb) R.Br* were noted and the results were presented in the **Table 1**.

Microscopic Characterization

Transverse section of the leaf shows two distinguished regions, midrib region and laminar region. T.S. of midrib of leaf showed chained, numerous and small epidermal cells. T.S. of lamina showed cuticle and thick-walled cells in lower and upper epidermis. Epidermal cells were large in size, elongated and compact **(Figure 1)**.

Quantitative Leaf Microscopy

A quantitative leaf characteristic which includes Stomatal no., Stomatal index, vein-islet number, vein-let termination number and Palisade Ratio were observed and the results were shown in the **Table 2**.

Physicochemical Parameters

The results of the various physicochemical constants of raw material lie within the limit which is given in Table 3; this signifies that the purity and quality of raw material was good enough. Insufficient drying may lead to spoilage by moulds and bacteria and makes possible the enzymatic destruction of active principles. Not only the ultimate dryness of the drug is important, equally important is the rate at which the moisture is removed and the condition under which it is removed thus the determination of moisture content also provide the method of preparation of drug and it is observed that the moisture content of the drug was found to be 4.2 ± 2.15% w/w which signify that the drug is properly dried and properly stored.

The total ash is particularly important in the evaluation of purity of drugs, i.e. the presence or absence of foreign matter such as metallic salts or silica. An analytical result for total ash was found to be 6.3% w/w. The amount of acid-insoluble matter present was 4.63 % w/w. The water soluble ash was found to be 2.42% w/w, this parameter is used to detect the presence of material exhausted by water. As the ash values of the crude drugs lies within the fair limit which signify its quality and purity and gives idea about the total inorganic content. The water soluble extractive values indicated the presence of sugar, acids and inorganic compounds.

The water-soluble extractive value found to be 10.72 ± 0.25% w/w and the alcohol soluble extractive was found to be 5.03 ± 0.23% w/w which signify that the large amount of constituents of leaves was soluble in water and alcohol.

Phytochemical Screening

The qualitative chemical tests were carried out for the identification of the different nature of phytoconstituents present in the crude drugs of *Artabotrys odoratissimus (Roxb)R.Br*by standard procedures. They are usually tested for the presence of alkaloids, flavonoids, phenols, tannins, cardiac glycosides, triterpenes, steroids and saponins. The results were shown in **Table 4**.

Fluorescence Analysis

The fluorescence analysis of the powder drug was done and results are given in Table No. 5. The powder was treated with various reagents and the mixture was observed under UV light (254nm and 366nm). Fluorescence study is an essential parameter for first line standardization of crude drug. In fluorescence the fluorescent light is always of greater wavelength than the exciting light. Light rich in short wavelengths is very active in producing fluorescence and for this reason ultraviolet light produces fluorescence in many substances which do not visibly fluoresce in daylight.

Table 1: Macroscopic and organoleptic characters of *Artabotrys odoratissimus (Roxb)R.Br* leaf

S.no.	Macroscopic parameters	Observation
1	Colour	Green
2	Odour	Pleasant
3	Taste	Bitter
4	Phyllotaxy	Alternate
5	Shape	ovate
6	Venation	Pinnate
7	Base	Acuminate
8	Apex	Acute
9	Margin	Entire

Figure 1: Leaves of *Artabotrys odoratissimus (Roxb)R.Br*

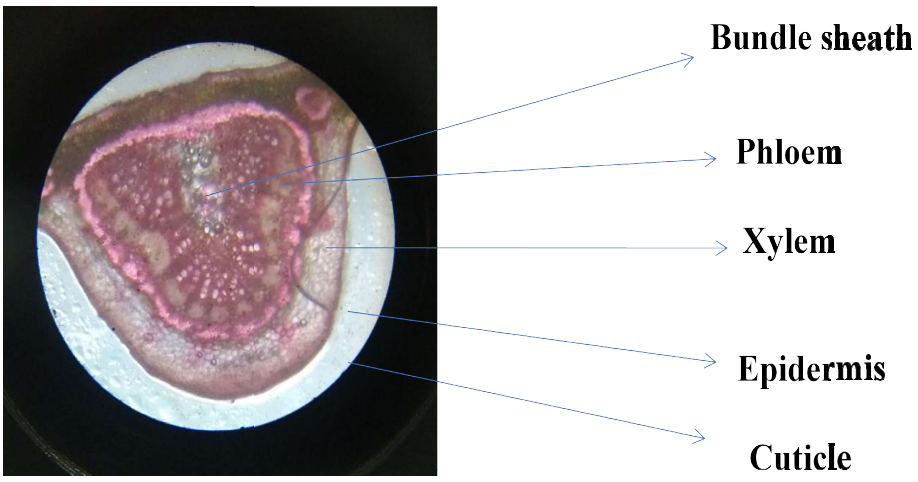


Figure 2: Transverse section of *Artabotrys odoratissimus (Roxb) R.Br* leaf

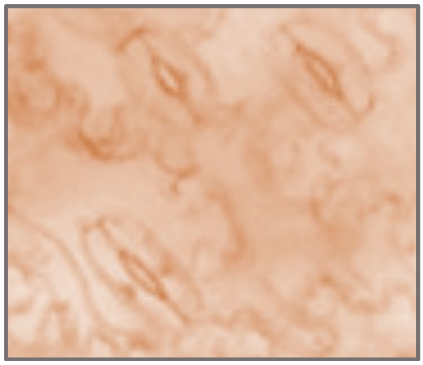


Figure 3: Leaf showing Stomatal no and Stomatal index

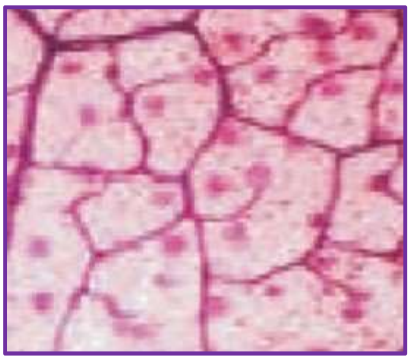
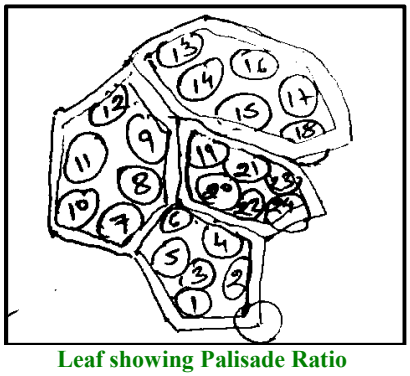


Fig no- 4 Leaf structure showing vein-let termination and vein-islet number



Leaf showing Palisade Ratio

$$\text{Palisade Ratio} = \frac{24}{4} = 6$$

Table 2: Quantitative Leaf Microscopy of *Artabotrys odoratissimus (Roxb)R.Br*

S.no.	Parameter	Range
1	Stomatal number	7
2	Stomatal index	11.26
3	Vein islet number	11
4	Vein let termination number	12
5	Palisade Ratio	06

Table 3: Physicochemical parameters of *Artabotrys odoratissimus (Roxb)R.Br*leaves

S.no.	Parameters	Values
1	Total Ash value	6.3% w/w
2	Acid insoluble ash	4.63 % w/w
3	Water soluble ash	2.42% w/w
4	Moisture content (loss on drying)	4.2 ± 2.15% w/w
5	Foaming index (10ml.conc.)	1ml.
6	Swelling index	5.11 ml.
7	Alcohol-soluble extractive	5.03 ± 0.23% w/w
8	Water-soluble extractive	10.72 ± 0.25% w/w

Values are expressed as mean ± SD of six values.

Table 4.Phytochemical screening of different extracts of *Artabotrys odoratissimus (Roxb)R.Br*leaves.

Tested Group	Ether Extract	Chloroform Extract	Methanol Extract	Aqueous Extract
Alkaloids	---	---	+++	+++
Glycosides	---	---	+++	---
Phenolic compound	---	---	+++	+++
Steroids & Sterols	---	+++	+++	+++
Saponins	---	---	---	+++
Flavones & Flavonoids	---	---	---	+++
Carbohydrates	---	---	+++	+++
Tannins	---	+++	+++	---

Note: (+++) Present (---) Absent

Table 5: Observations of *Artabotrys odoratissimus (Roxb)R.Br*leaves powder under visible light and UV (254nm and 366nm)) light.

S. No.	Treatment	Visible light	Observation (Colour developed)	
			UV-254nm	UV-366nm
1	Sample as such	Light Green	Green	Green
2	Powder + 1N aq. NaoH	Brown	Green	Black
3	Powder + 1N alc. NaOH	Brown green	Black	Blackish green
4	Powder + 1 N HCl.	Green	Black	Purplish green
5	Powder + 50% HNO_3	Brown	Black	Green
6	Powder + 50% H_2SO_4	Black	Black	Greenish Black
7	Powder + Methanol	Reddish brown	Green	Black
8	Powder + NH_3	Brown	Light Green	Greenish Black
9	Powder + I_2	Greenish Brown	Dark black	Green
10	Powder + $FeCl_3$	Brownish	Black	Light green

CONCLUSION

In the present study as per establishment of pharmacognostical standard and quantitative parameter, a great bulk of information on identity, purity and quality of plant material is gained while evaluating the macroscopy, microscopy, powder and physicochemical characters. Thus, all studied standardization parameter like pharmacognostical study; phytochemical screening and physicochemical parameter provide the knowledge in the identification and authentication of leaf of *Artabotrys odoratissimus (Roxb)R.Br.*

REFERENCES

[1] Chandra R, Mahato M, Mondal SC, Kumar K, Kumar J. Ethnomedicinal formulations used by traditional herbal practitioners of Ranchi, Jharkhand. Indian. J Tradit Knowle 2007; 6(4):599-601.

[2] Qureshi Md. Shamim, Giri IC, Panday VK, Choudhary R, Patel J. Anthelmintic activity of Smilax zeylanica leaf. Research J. of Pharmacognosy and Phytochemistry 2009; 1(1): 78-79

[3] Siddiqui MB, Husain W. Medicinal plants of wide use in India with special reference to Sitapur district (Uttar Pradesh). Fitoterapia 1994; 65:3.

[4] Jitendra B Jain, Sheetal C, Kumane S, Bhattacharya. Medicinal flora of Madhya Pradesh and Chattisgarh – A Review. Indian Journal of

Traditional Knowledge 2006; 5(2):237-242.

[5] Saxena SK, Tripathi JP. Ethnobotany of Bundelkhand I Medicinal uses of wild trees by tribals inhabitants of Bundelkhand region. J Econ Taxon Bot 1989; 13(2):381- 389.

[6] Qureshi R, Bhatti GR. Floristic and ethnobotanical study of Desert-Nara Region, Sindh, Shah Abdul Latif University. Pakistan Research Repository. 2004; 12(1):454.

[7] Md. Shamim Qureshi, Venkateshwar Reddy A, Jitendra Patel, Safwan Ali Khan, Mohd Khaliquzzama. Evaluation of the Antipyretics activity of Trapa bispinosa root. Journal of Pharmacy and Chemistry 2011; 5(1):3-4.

[8] Ahmad I, Aqil F, Owais M. Modern phytomedicine: turning medicinal plants into drugs. New York: John Wiley & Sons; 2006.

[9] Willow JH. Traditional herbal medicone research methods: identification, analysis, bioassay and pharmaceutical and clinical studies. New York: John Wiley & Sons; 2011.

[10] Benzie IF, Wachtel-Galor S. Herbal medicine: biomolecular and clinical aspects, oxidative stress and disease. 2nd ed. Florida: CRC Press; 2011. p. 499.

[11] Odugvemi T. A textbook of medicinal plants from Nigeria. Nigeria: Tolu Odugbemi; 2008.

[12] Choudhury, M.H., Shaheen, S., Ilias, M., Gray, A.I. & Waterman, P.G., J. Nat. Prod., 50, 1987, 762. 2.

[13] The Wealth of India, A dictionary of Indian Material and Industrial Products Vol.1:A, Raw material publication and Information Directorate Council of Scientific and Industrial Research. New Delhi: 2005, p. 433.

[14] Sharma, M., Srilakshami, Desiraju, S., Chaurey, D. & Mehta, B.K., Grasas Y Aceite, 53(2), 2002, 187-189.

[15] Connolly, J.D., Haque, M.E., Hasan, C.M. & Haider S.S., Fitoterapia. 65, 1994, 92.

[16] Chakabati, B., Chaudhuri, A., & Chowdhury, P.R., Journals of Indian Medical Association, 51(5), 1968, 227-229.

[17] Thakur DS, Kumar P, Kujur A, Kumar P, Kumar R. Contribution of Male Contraception in World Population. Journal of Pharmaceutical Sciences and Research. 2010; 1:384-393.

[18] Parveena Devi, Pradeep Kumar, Nidhi, Isha Dhamija. Antifertility activity of medicinal plants on male and female reproduction. International Journal of Pharmaceutical Sciences and Research. 2013; 4:122-128.

[19] Suresh Joshia, Aksha Sharmaa, Mridula Chaturvedib. Antifertility potential of some medicinal plants in males: an overview. Int. J Pharm. Sci. 2011; 3(5):204-217.

[20] A.P. Singh and M. Sahai, Planta Medica., 62, 192 (1996).

[21] Sidhiqui, N., & Garg, S.C., J. Sci. Ind. Res., 33, 1990, 526-537.

[22] C.P. Khare. Indian Medicinal Plants. Springer. 2007. pp. 65.

[23] Trease GE, Evans WC. Trease and Evans Pharmacognosy. Edn 15, Harcourt Brace & Co: Asia, Pvt. Ltd. and W.B. Saunders Company Ltd, Toranto, 2002, 537.

[24] Wallis TE. Text book of Pharmacognosy. Edn 5, CBS Publishers and Distributors; New Delhi 2005.

[25] SG Nayak, MA Iyengar. Anatomy of Crude Drugs.Edn 11, Manipal Press Limited, Manipal, 2008, 01-08.

[26] MA Iyengar. Pharmacognosy of Powdered Crude Drugs. Edn 5, Manipal Press Limited, Manipal, 1997, 21-31.

[27] Kokate CK. Practical Pharmacognosy. Vallabh prakashan, 1996, 115-120.

[28] World Health Organization. Quality control methods for herbal materials. Geneva: WHO Press, Malta, 2011, 20.

[29] Srinivasa B, Kumar A, Prabhakarn V, Lakshman K, Nandeesh R, Subramanyam P. Pharmacognostical studies of *Portulaca oleracea* Linn Braz. J Pharmacogn. 2008; 18(4): 527–531

[30] Kokate CK. Practical Pharmacognosy. Vallabh prakashan, 1996, 115-120.

[31] Kumar S, Kumar V, Prakash O. Pharmacognostic study and anti-inflammatory activity of *Callistemon lanceolatus* leaf. Asian Pac J Trop Biomed. 2011; 1(3):177-81.

[32] World Health Organization. Quality control methods for herbal materials. Geneva: WHO Press, Malta, 2011, 28.

[33] Indian Pharmacopoeia. Edn 4, Vol.2, Government of India, Ministry of health and welfare, Controller of publications, New Delhi, 1996, A53-4.

[34] World Health Organization. Quality control methods for herbal materials. Geneva: WHO Press, Malta, 2011, 29-30.

[35] World Health Organization. Quality control methods for herbal materials. Geneva: WHO Press, Malta, 2011, 31-33.

[36] World Health Organization. Quality control methods for herbal materials. Geneva: WHO Press, Malta, 2011, 34-36.

[37] World Health Organization. Quality control methods for herbal materials. Geneva: WHO Press, Malta, 2011, 37-40.

[38] World Health Organization. Quality control methods for herbal materials. Geneva: WHO Press, Malta, 2011, 41-44.

[39] World Health Organization. Quality control methods for herbal materials. Geneva: WHO Press, Malta, 2011, 45-47.

[40] Kokate C. K. Pharmacognosy. Edn 50, Vol.1-2, Nirali Prakashan, Pune 2010, Appendix A1-A6.

[41] Harborne JB. Phytochemical methods. Edn 2, Chapman and Hall, London, 1973.

[42] Kokoski J, Kokoski R, Salma FJ. Fluorescence of powdered vegetable drugs under ultraviolet radiation. J. Am Pharm Ass 1958; 47:715-717.

[43] Mohammad Shamim Qureshi, A. Venkateshwar Reddy, G.S. Kumar, Lubna Nousheen. Pharmacognostic, Physicochemical Standardization and Phytochemical Analysis of leaves of *Hydrolea zeylanica* Vahl. (Hydrophyllaceae). Research Journal of Pharmacognosy and Phytochemistry. 2017; 9(1): 1-7.

[44] Zhao Z, Liang Z, Guo P. Macroscopic identification of Chinese medicinal materials: Traditional experiences and modern understanding. J Ethnopharmacol 2011; 131:556-561.