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THE THERAPEUTIC VALUE OF LYSINE AGAINST CANCER: A COMPREHENSIVE REVIEW

SOUVIK DEBNATH¹, AVINABA MUKHERJEE^{2#}, SAUMEN KARAN¹, TAPAN
KUMAR CHATTERJEE^{1*}

1: Division of Pharmacology Research Laboratory, Department of Pharmaceutical
Technology, Jadavpur University, Kolkata-700032, India

#2: Department of Zoology, Charuchandra College, University of Calcutta, Kolkata-700029,
India

1: Division of Pharmacology Research Laboratory, Department of Pharmaceutical
Technology, Jadavpur University, Kolkata-700032, India

*1: **Corresponding Author: Prof. Tapan Kumar Chatterjee, Dean, Department of
Pharmaceutical Science and Technology, JIS University, Kolkata; Former Professor,
Division of Pharmacology, Department of Pharmaceutical Technology, Director,
Jadavpur University, Jadavpur, Kolkata-700032, India: E Mail:**

dr.tkchatterjee@jisuniversity.ac.in

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ABSTRACT

Current treatment methods of cancer include surgical intervention and the usage of chemosynthetic drugs or natural products. However, the synthetic drugs show broad-spectrum cytotoxicity and a higher dose for cellular inhibition. Therefore, using an amino acid like Lysine for the cancer treatment raises a new hope, as this therapeutic molecule bears no cytotoxicity towards the normal cells and works by altering the cellular metabolism from the very first stage of cancer development. Lysine was found to be significant when applied as a single moiety or in a derived form and also in a combinational form with the other drugs. This review elaborates on the current state of development, recent advancement of this amino acids highlighting the application of Lysine against several cancer cells. Overall, the effectiveness of Lysine and the molecules that it targets for the inhibition of cancer proliferation emerge a new area in the anticancer research, which proves Lysine as a potential aspirant drug in cancer therapy.

Keywords: L-lysine; cancer; cytotoxicity; apoptosis; therapy

INTRODUCTION

Cancer is the second leading cause of human death after cardiovascular diseases [1]. Almost 90% of cancer treated deaths are due to the metastasis of cancer cells. Cancer cells through their metastatic ability cause 90% of all cancer-related death [2]. Although struggling efforts against cancer have grown tremendously in the past few years, still it is the leading cause of death in economically developed countries. An increase in the worldwide death rate was found in cancer patients which are 7.1 million and 8.2 million in 2007 and 2012, respectively [3]. By 2030 the global burden is expected to grow to 21.7 million new cancer cases [4]. Several approaches, including surgery, chemotherapy, and radiation therapy are available for the treatment of cancer, but they often lack target specificity that compromises therapeutic benefits [5]. Development of tumor resistance against the available synthetic drugs together with the patient noncompliance invokes the requirement of safe and newer drugs and for the same, extensive research on several synthetic drug molecules is being undertaken for cancer drug discovery. Thus the imperatives for novel drugs for cancer therapies with improved specificity and safety are highly desirable.

One of the cancer treatments that seem to custom Controlled Amino Acid Treatment

(CAAT). CAAT works by restricting certain amino acids from the diet to combat the growth and reproduction of the cancerous cells. However, the treatment model of this appears to be target-specific, noncytotoxic towards normal cells or tissues and also easily sustainable in normal physiological circumstances. Amongst the amino acids, lysine was found to be more appropriate over the others. It is having a simple chemical structure and good biochemical properties for the treatment of cancer cells. Having simple in structure and for some other biochemical properties lysine seems to be the most suitable amino acid for the treatment of cancer cells. This review opens up the role of lysine as an appropriate anticancer agent for the improvement of modern science in anticancer chemotherapy.

Benefits and Usage of CAAT as Cancer Therapeutic Agent

Amino acids used in CAAT studies fabricating good results as they are inoffensive to the normal cells. Although there are so many other explanations for their usage. Using amino acids as CAAT is demonstrating effective increase of studies as it is target-specific, non-cytotoxic concerning the normal cells and tissues. However, this is not the only cause of why amino acids are used for the treatment of cancer (Table 1). Amino acids have also

different effects on cells. Some help them to grow and imitate while others keep them healthy and true to the DNA blueprint entrenched in the cell at its construction.

There deceptions the prominence of using CAAT against cancer cells, as they can moderate the metabolism and associated pathway of cancer cells.

Table 1: Alteration of amino acid management pathways in cancer

Amino acid metabolism			
Gene	Function	Cancer alteration/relevance	References
GLS	Rate limiting step in glutaminolysis	Increased translational efficiency downstream of MYC, miR-23a/b	[98]
PHGDH	Rate limiting step in serine biosynthesis	Genomic amplification and over-expression	[99][100]
SHMT2	Diversion of serine into mitochondrial one-carbon metabolism	Over-expression drives hypoxia resistance	[99][101][102]
GLDC	A key component of the glycine cleavage complex	Prevents toxic glycine accumulation, drives broad metabolic changes	[102][103]
ASNS	Asparagine biosynthesis	Increased expression in glioblastoma	[104]
Amino acid sensing			
mTOR	Protein kinase controls translation in response to nutrient sufficiency signals	Activating point mutations, amino acid starvation fails to inactivate mTORC1	[105] [106]
FLCN	the mTORC1 positive regulator, GTPase activating protein for RAG C/D	Loss of function mutations in Birt–Hogg–Dubé syndrome	[107][108][109]
DEPDC5	GATOR1 component, a negative regulator of mTORC1, GTPase activating protein for RAGA/B	Deletion of 22q12.2, amino acid starvation fails to inactivate mTORC1	[110]
NPRL2	GATOR1 component, a negative regulator of mTORC1, GTPase activating protein for RAGA/B	Deletion of 3p21.3, amino acid starvation fails to inactivate mTORC1	[110]

A brief overview of L-Lysine and its advantage over the other amino acid

L-lysine is an essential amino acid and we have to be contingent on animals for its procurement in the body. It is having an amended anticancer effect than other amino acids. L-lysine is an indispensable amino acid, not produced in an animal's body, but it originates in nature still, an ingenious plotline as it has the most advantageous over the other amino acids when being used as anticancer agents. Upon intake, L-lysine encourages healthy tissue function,

growth, and healing that progresses the immune system. L-lysine is most effective in oral supplements. It is very convenient as an anticancer agent inside the body and it is under unceasing research. As a natural disease-fighting agent, L-lysine benefits the human body in a diversity of ways, many of which are only recently experiencing research.

L-Lysine Help in the Treatment of Cancer

In the contemporary drug distribution research targeted drug transfer can be

accomplished by using soft neutraceuticals like L-lysine as they do not disturb the normal cellular function. In 2007, researchers at Florida State University studied the effects of “lysine conjugates” on damaged strands of DNA, like the ones found in cancer. Essentially, this ingredient can localizethe damaged strand by recognizing “cleavage” in the situation a damaged spot and cause the respite of the strand to cleave tear a part additionally. The cell is typically incapable to repair this damage, leading to apoptosis; the suicidal death of cells.

The anti-neoplastic activity of Poly-L-lysine

Numerous studies have specified that Poly-L-lysine bears some activity against murine tumors [6]. Poly-L-lysine has also been found to exhibit a huge number of unique membrane belongings which comprise the aptitude to improve the cellular uptake [7] specifically in the agglutination of lymphocytes of the cancer patients [8, 9].

These possessions are possibly accompanying with the polycationic character of Poly-L-lysine and conceivably due to precise interactions on the cell membrane. Poly-L-lysine exhibited noticeable concentration - dependent cytotoxicity to HeLa cells in culture as well as anintense antineoplastic effect against Ehrlich ascites carcinoma, Dalton's lymphoma ascites cells and Sarcoma-180

tumorcells in White Swiss mice [10]. The results indicate that poly-lysine is cytotoxic against cancer cells, but it has some selective possessions because at intermediate concentrations it can produce remission from Ehrlich ascites without producing much toxicity. This implies that, at the time of poly-lysine addition, protein, DNA, and RNA synthesis can be completely inhibited. The important point, however, is that treatment with poly (L-lysine) at 20 µg/ml, a minimum 75% of the cells remained viable even though their DNA, RNA, and protein synthesis had been stopped for the longer period. Moreover, it was found that poly-lysine had several other properties on HeLa cells in culture. Histologically, at low poly-lysine concentrations that did not significantly affect subsequent cell viability for the shorter time period, but its effects for the longer time period and cells viability significantly decreased, the cells became shrinkage and apoptotic bodies forms. At higher concentrations of poly-lysine, the cells were lysed and their components covered the surface of the plate as diffuse crenated discs. Under these circumstances, the cells were not viable and apoptotic bodies formed. Poly-L-lysine has profound cytotoxic effects. In addition, it is found to be most effective against Ehrlich ascites carcinoma cancer cells induced poly-lysine is very effective in preventing tumor

growth in mice. Our studies indicate that initially, the L-isomer is much more effective at preventing cell growth than the D isomer. Probably the growth of cancer cells is inhibited by poly-L-lysine due to its ability to increase the efflux of small molecules that are required for RNA and protein synthesis. For example, furthermore to the increased efflux of potassium, it was found that poly-lysine endorses the leakage of inorganic phosphate, carbohydrates, free amino acids, small peptides, and adenosine 5'-monophosphate into the supernatant [11]. Previous researchers [12] disclose that poly-L-lysine, as well as other poly-cations such as poly-L-ornithine and poly-L-arginine, can trigger membrane phospholipase A2 in 3T3-4a Swiss mouse fibroblasts. These findings deliver a fascinating mechanism by which poly-lysine may disturb membrane permeability to small molecules by endorsing the hydrolysis of membrane phospholipid.

Based on the surveillance, the scientists propose some model for the communication of poly-lysine with cancer cells. Those are; Poly-lysine, either poly (L-lysine) or poly (D-lysine) dilemmas forcefully to the cell membrane and induces morphological changes. Secondly, accompanying such binding is a rapid leakage of small molecules across the cell membrane, perhaps promoted by phospholipase A2

activation. Then concomitant with the poly-lysine binding is the breakage of nuclear DNA, the rapid loss of RNA producing inhibition of protein synthesis [13]. When poly-L-lysine is used in growing cancer cells in the culture condition, it reasons a superior degree of inhibition for short periods than that of the D isomer. After approximately 10 hr, however, the L isomer is deceptively besmirched and the abrasion modifies itself because the cells return to control growth. In contrast, with increasing time, poly-D-lysine demonstrates time-dependent cellular growth inhibition on other-hand caused the toxicity in EAC, HeLa and Lewis lung carcinoma-induced tumor growth in the mice model. PLL has potent activity in an *in-vivo* model especially against, induced Ehrlich ascites tumors in mice but little effect upon L1210 induced tumors.

The systemic anti-angiogenic activity of Poly-L-lysine in the form of a dendrimer

Several studies propose that the Poly-L-lysine may be used for therapy of solid tumors, and also in a combination with their capability to carry other therapeutic or diagnostic mediators may offer competences for the proposal of theranostic systems. The antiangiogenic activity of the Poly-L-lysine dendrimer can produce biological activity and therapeutic effectiveness in the absence of any other therapeutic agent. Methotrexate (MTX)

conjugated to PEGylated PLL dendrimers have also shown to accumulate in solid Walker 256 and HT-1080 tumors induced rats and mice model [14]. The study by Fox et al., 2009 therapeutic efficacy of PEGylated PLL dendrimer-camptothecin conjugates in C26 and HT-29 tumor models [15]. Further work shows that G6 Poly-L-lysine dendrimer (MW 8149 Da) can accrue and continue in solid tumor sites after systemic administration and demonstration of antiangiogenic activity in the absence of cytotoxicity in normal tissues. The occurrence of Poly-L-lysine dendrimer at lower concentrations was originated to inhibit endothelial cell migration and three-dimensional tubule formation. G6 Poly-L-lysine dendrimer was appraised herein at 25 mg/kg and 50 mg/kg after intravenous administration via tail vein injection in mice. There is previously reported that blood half-lives of <10 min, suggesting rapid accretion of the dendrimers in the vascular endothelium, a process driven by electrostatic communications [16, 17]. Poly-L-lysine-dendrimer accumulation at the tumor site takes place via strong electrostatic interactions between the cationic Poly-L-lysine-dendrimer surface and the negatively charged, heparin-rich ramparts of the tumor neovasculature. Antiangiogenic activity *in-vivo* succeeding systemic administration of PLL dendrimers was primarily acquired using the Matrigel

plug assay. The competence of systemically administered Poly-L-lysine dendrimer to hinder microbial development *in-vivo* was additionally evaluated in the dorsal skinfold window chamber model. The gradation of vascularization and significant amendment in the emerging microvascular architecture were observed by intravital microscopy [18, 19]. The Poly-L-lysine-dendrimer-treated group exhibited condensed numbers of microvessels subsequently treatment with Poly-L-lysine dendrimer compared to the control group. An additional indication of the Poly-L-lysine-dendrimer antiangiogenic motion was provided by the important reduction of the number of CD31⁺ cells in the tumor sections of the treated groups compared to untreated control groups. The intensification in TUNEL staining of solid tumors in C57/BL6 mice further designated that the Poly-L-lysine-dendrimer encouraged apoptosis in a large tumor cell population. To regulate whether a therapeutic consequence can be initiated from the Poly-L-lysine-dendrimer antiangiogenic activity, the B16F10 murine melanoma tumor model was used. It has been recognized by others that a decrease of microvessel density in B16F10 tumors can amend the tumor cell proliferation and lead to tumor cell demise by apoptosis or necrosis [20]. In this study, only two intravenous administrations of the Poly-L-lysine dendrimer were able to prime

to the postponement of tumor growth [21]. Poly-L-lysine-dendrimer treatment led to an interruption in tumor volume expansion the time of 4.7 ± 2.5 days compared to 3.7 ± 1.6 days when preserved with vehicle control. Therefore, the Poly-L-lysine dendrimer seems comparable in deferring B16F10 tumor growth to Avastin. Many molecules have been previously described to exhibit antiangiogenic activities *in-vitro*; nevertheless, most have not been effective *in-vivo*, either due to the absence of an optimum pharmacokinetic outline or due to systemic toxicity at vigorous biological doses.

Researchers also described that, the complexation of the chemotherapeutic drug doxorubicin (DOX) with the sixth-generation cationic poly-L-lysine dendrimer (DM). DOX-DM complexation was established by fluorescence polarization measurement, proton nuclear magnetic resonance spectroscopy, and molecular modeling. Heightened dispersion of DOX-DM (at 1:10 molar), paralleled to the free DOX, into prostate 3D multicellular tumor spheroids (MTS) was established by confocal laser scanning microscopy. Additionally, DOX-DM developments accomplished significantly advanced cytotoxicity in the DU145 MTS system compared to the free drug, as revealed by progression delay curves. Incubation of MTS with low DOX

concentration complexed with DM led to a significant interruption in MTS evolution compared to untreated MTS or MTS treated with free DOX. DOX-DM multifaceted maintenance was also accomplished in a Call-6 lung cancer xenograft model in tumor-bearing mice. Therapeutic experimentations in B16F10 tumor-bearing mice originate to deliver the heightened therapeutic effectiveness of DOX when complexed to DM [22]. This study proposes that the cationic poly-L-lysine DM molecules deliberate here could, in accumulation to their systemic antiangiogenic possessions, multifaceted chemotherapeutic drugs such as DOX and advance their accretion and cytotoxicity into MTS and solid tumors *in-vivo* [23]. Such a tactic proposes new competencies for the strategy of conjoining antiangiogenic/ anticancer therapeutics. Intravenous administration of only two doses at 50 mg/kg Poly-L-lysine dendrimer occasioned in determine daccretion in solid B16F10 solid tumor model, lessening in vascularization, extensive apoptosis/ necrosis within the tumor tissue, and a statistically significant reduction in tumor volume, in the absence of any outstanding histological or physiological irregularity in nontumor tissues such as liver and kidneys. The dendrimer exhibited therapeutic effectiveness compared to the commercially obtainable anti-VEGF

antibody bevacizumab (Avastin) in adjuvanting the growth of a subcutaneous B16F10 melanoma tumor model.

Antitumor Effect of L-Lysine as Combinational Therapy

On bladder cancer cell line

The synergistic anti-cancer effect of ascorbic acid, proline, lysine, and epigallocatechin gallate (EGCG) on several cancer cell lines in tissue culture studies was greater than that of the individual nutrients. The synergistic anti-cancer effects of ascorbic acid, proline, and lysine on several cancer cell lines in tissue culture studies were greater than that of the individual nutrients [24]. The modulation of matrix metalloproteinases (MMP) on the degradation of the extracellular matrix (ECM) plays a serious role in the development of tumors and metastasis and has been originating to correlate with the aggressiveness of tumor growth and invasiveness of cancer [25, 26]. Afterward, researchers have recognized the perception designated by Rath and Pauling [27, 28] and ensued in recognizing a novel formulation of lysine, ascorbic acid and proline which has shown significant anticancer activity against a large number of cancer cell lines, blocking cancer growth, tissue invasion and MMP expression both *in-vitro* [29, 30] and *in-vivo* [31, 32]. The detailed mixture of lysine, proline and ascorbic acid is an

outstanding application for preemptive and therapeutic use in the treatment of bladder cancer [33, 34]. However, other studies on animal models and clinical trials are essential for more exploration of the role of nutrient supplementation in the treatment of bladder cancer.

On Pancreatic Cancer Cell Line

The dose-dependent inhibitory effect of the nutrient combination of lysine, proline, arginine, ascorbic acid, and green tea extract on the MMP-9 expression of the pancreatic cancer cells was dependable with its dose-dependent inhibition of matrix invasion [35]. Also, this mixture of nutrients possibly improved the constancy and forte of the connective tissue, as optimization of amalgamation and construction of collagen fibrils depends on hydroxylation of proline and lysine residues in collagen fibers. Researchers already reported that the synergistic anticancer effect of proline and lysine on several cancer cell lines in tissue culture studies was superior to that of the individual nutrients [28]. Furthermore, in contrast to chemotherapy which causes undiscerning cellular and ECM damage, morphological studies of pancreatic cancer cells treated with NM presented that level at the highest concentrations of the nutrient mixture (NM), the pancreatic cells were not adversely exaggerated, demonstrating that this formulation is safe for normal [35].

A derivative of L-Lysine as a Natural Source

Epsilon-poly-l-lysine (ε-PL) from Streptomyces or Kitasatospora

Epsilon-poly-l-lysine (ε-PL) produced by *Streptomyces* or *Kitasatospora* is a homopolymer of L-lysine [36], and it is also decomposable, water-soluble and thermally stable. It is established to non-toxic in humans [37]. ε-PL is an auspicious natural antimicrobial that is extensively used to preserve packaged food in convinced countries for its broad antimicrobial activity against Gram-negative and Gram-positive bacteria, yeasts, and molds [38]. However, the accomplishment of this approach is also reproduced in anticancer research as ε-PL is initiated to reduce the tumor size. The resection lowers the degeneration rate and is also suitable [39, 40]. The compounds also exhibited better antioxidant and antitumor activity [36]. These findings indicate that the incorporation of ε-PL could be used in the pharmaceutical industry.

L-lysine α-oxidase from Trichoderma faureoviride

L-Lysine-α-oxidase (LO), catalyzes the adaptation of L-lysine into α-keto-ε-aminocaproic acid, H₂O₂, and ammonia with the ingesting of oxygen. The enzyme was isolated for the first time in the 1980s from *Trichoderma viride* Y244-2 [41, 42].

The anticancer activity of LO is closely related to L-lysine plasma concentration exhaustion [41]. A single intravenous injection of LO (30U/kg) into BDF₁ mice in 1 h reduced the L-lysine concentration to an indiscernibly low level. This diminished L-lysine level was continued for 12 h and then improved gradually. In addition to the enzymatic dispossession of L-lysine, the cytotoxicity of LO could be attributed to H₂O₂ production. DNA breakage by H₂O₂ may be restrained as the primary mechanism of the LO cytotoxic effect on tumor cells *in-vitro* [43]. The amino acid oxidizing enzymes, LO from *T. viride* or *T. Harzianum* Rifai has a comparatively narrow range of substrates [41]. Only L-amino acids allow a positive charge at the end of the side radical, such as L-ornithine and L-arginine, are rehabilitated by LO, but at a much leisurely rate than L-lysine. LO was revealed to be less toxic human carcinoma cell lines, with IC₅₀ values ranging from 3.0 x 10⁻⁶ to 7.8 x 10⁻² U/ml (3.2 x 10⁻⁸ to 8.2 x 10⁻⁴ mg/ml). Compared with other anticancer enzymes, LO is much more active *in-vitro* than L-asparaginases or novel anticancer enzymes such as arginine deiminase and onconase [44-46]. The most sensitive cell line was K562 (IC₅₀=3x 10⁻⁶ U/ml). LO may have been a result of the different compassions to L-lysine deficiency and oxidative stress. Although the basic level of cytotoxicity is

determined by H₂O₂ production [43], possibly the sensitivity to depletion of L-lysine concentration could be the main standard for the prediction of LO effectiveness in various types of cancer. The sensitivity to growth inhibition *in-vitro* was established *in-vivo* in mice bearing LS174T and HCT116 tumor xenografts. The discoveries of this *in-Vivo* experiments confirm the possible appropriateness of LO for human colon cancer treatment and specify that the advantages shown by LO may interpret into upgraded clinical benefits deprived of increased toxicity. A specific agenda is vigorous for an actual and well-tolerated LO treatment. Optimal possessions of anti-cancer drugs with an enzymatic mechanism of action can be

accomplished by administering them in numerous daily doses, which guarantee sustained exposure to the drug, but this schedule leads to increase in toxicity because of a precarious unceasing decrease of the plasma L-lysine concentration. Perhaps, conservation of doses of LO delivers effective concentrations of L-lysine for the survival of normal cells whereas cancer cells are more susceptible. The poor blood supply in a tumor does not deliver continuous ingestion of indispensable amino acids critical for cycling cells with exhaustive protein synthesis, especially in G₁. This was established by the PK profile of LO [47] (Table 2).

Table 2: Effect of Lysine in different forms (Single, combined and derivatives) on various cancer cells

Molecules	Types of cancer cells on which effects are shown	References
Single forms		
Lysine oxidase	Colorectal cancer cell	[111]
L-lysine	Leukemia cell Bladder cancer cell Pancreatic cancer cell	[112]. [113], [114], [115], [116],[117], [118]. [119].
Poly-lysine	HeLa cell Ehrlich Ascites Carcinoma cell Sarcoma-180, Dalton Ascites Lymphoma cell	[120][121][122][123].
Combined forms		
PEGylated Poly-L-lysine	Solid Walker 256 tumor cell HT-1080 tumor cell C-26 tumor cell B16F10 murine melanoma tumor	[124]. [125][126]. [127]. [128].
Poly-L-lysine dendrimer complexes Doxorubicin	Calu-6 lung cancer cell B16F10 murine melanoma tumor	[129][113]. [128]
Derivatives of L-lysine		
ϵ -Poly-L-lysine (ϵ -PL)	HepG-2 cancer cell	[130][131][132][133].
L-lysine- α -oxidase	HCT116 tumor cell LS174T tumor cell	[134][135][134][136][137][138]. [134][139][140].

Mechanism of Action of L-Lysine as an Anticancer Molecule

L-the lysine regulates tumor necrosis factor alpha (TNF- α) and matrix metalloprotein-3 (MMP-3) expression

L-Lysine is intricate in multiple biological processes including inflammatory regulation. However, rare research has spoken about the effects of Lysine on human chondrocytes. The mRNA levels of tumor necrosis factor- α (TNF- α) and matrix metalloproteinase-9 (MMP-9) reduced when normal chondrocytes treated with Lysine. Lysine down-regulated TNF- α , MMP-3 levels, reinstated aggrecan and collagens expressions, and further increased the aggrecan/type I collagen and type II collagen/type I collagen provisions in IL-1 β -stimulated chondrocytes. In addition, Lysine's treatment reduced the protein fabrications of TNF- α and MMP-3 in encouraging cells. Lysine has been originating to have beneficial effects in diverse pathological circumstances including herpes simplex virus infection, mood disturbances, anxiety, migraine, and osteoporosis [48-51]. Chinese traditional medicine consumptions Lysine in combination with extracts of medicinal plants to constrain acute inflammation [52]. Also, Lysine further decreased TNF- α mRNA expression and protein production. TNF- α is involved in systemic inflammation and related to the acute phase reaction [53]. TNF- α also excites the

announcement of MMP-1, MMP-3, and MMP-13 [54]. This study revealed that the regulations among pro-inflammatory cytokines, MMPs, and TIMPs are predominantly moderate by Lysine. IL-10 modifies the countenance of anti-inflammatory cytokines, manufacture of reactive oxygen species, and a factor of pro-apoptosis of human articular chondrocytes [55, 56]. Since Lysine reduced the TNF- α level in stimulated chondrocytes, Lysine may not change IL-10 production. Further studies displayed that the productions of TNF- α and MMP-3 reduced instantaneously in IL-1 β -stimulated chondrocytes after Lysine's treatment. The decrease of TNF- α and MMP-3, consequently, designates that the anti-inflammatory activities of Lysine encompass the conquest of the inflammatory pathway. An exciting discovery is the modulation of Lysine in inflammatory cytokines to un-stimulated chondrocytes. Lysine did not have substantial effects on IL-1 β but decreased the mRNA levels of TNF- α . It is known that IL-1 interrelates with TNF- α in various physiological and pathological circumstances. Although IL-1 β is an upstream inductor to TNF- α , TNF- α can also initiate IL-1 production in chondrocytes. Also, it was demonstrated that TNF-induced structural combined impairment is mediated by IL-1 [57]. Martel-Pelletier *et al.*, (1998) studied the

effects of diacerein and rhein with Lysine on inflammatory cytokines in lipopolysaccharide-treated synovium and chondrocytes. These representatives produced both dose and time-dependent reduction in IL-1 β protein construction in synoviocytes, and the amount of IL-1 receptors (IL-1R) on chondrocytes [58]. Numerous investigates of randomized measured trials of these TNF- α inhibitors exposed the conceivable risks of cancer [58, 59]. Lysine enhanced hypertrophic conversion of chondrocytes under IL-1 β inspiration. Lysine also reductions the mRNA levels and protein constructions of TNF- α and MMP-3. Generally the imbalance meant of cytokines and growth regulatory proteins by Lysine designate that Lysine may retain anti-inflammatory that may be one of motive on how lysine triggers cell death in cancer cells.

Poly (l-lysine) s/poly (l-lysine)-DNA complexes initiate mitochondrial-mediated apoptosis pathway

Recent studies designated that PLL induces mitochondrial apoptosis in EAC cells, *in-vivo* [60]. Besides this, the molecule was also found to trigger apoptosis which occurred to be facilitated through the intrinsic pathway in DAL and sarcoma-180 induced mice model [61].

On the other hand, synthetic polyamines such as linear and dendritic poly(l-lysine)s, PLL, and variations thereof, have also been

used for DNA compaction and transfection with capricious [62-66]. PLL-DNA complexes do not professionally escape from endosomes, but gene expression still occurs [67]. Explanations of some researchers with poly (ethylenimine) (PEI) [68], and characteristic cytotoxicity of PLLs [64, 69, 70]. Provided the intrinsic mechanisms behind and induced apoptosis in different types of human cell lines (Jurkat T-cells, umbilical vein endothelial cells, and THE-3 hepatocyte-like cell line) by high and low molecular weight PLL (H-PLL and L-PLL) in linear form, which is normally used in transfection protocols. The intracellular levels of endogenous polyamines (e.g., spermine and spermidine) are strongly controlled through several homeostatic mechanisms, whereas their extreme cytoplasmic accretion (e.g., resulting from insults to anionic cellular structures) transduces a death signal to mitochondria by an oxygen-independent mechanism [71, 72]. Incubation of Jurkat T-cells with PLL causes early events in the apoptotic cycle [73, 74]. PS translocation is most rapid (30 min–1h) with H-PLL at concentrations of 20 μ g/ml and above, while PS acquaintance is prominent from 5 h post-incubation with L-PLL. Similar results were also obtained with THE-3 hepatocytes and HUVECs. The activity of this protease accountable for caspase cleavage was thoughtful particularly at 24

h-post-PLL (20 $\mu\text{g/ml}$) treatment, was irrespective of PLL type and could be inhibited by Ac-DVD (a caspase-3 inhibitor). H-PLL was more effective than L-PLL in inducing caspase-3 activation. Also, caspase-3 instigation was noticeable at 24 h post-treatment with lower concentrations of both PLL types (10 $\mu\text{g/ml}$); this was in accord with PS expression in tandem with propidium iodide staining. Researchers also obtained similar results with PLL–DNA complexes, and also in different types of human cancer cell lines. Henceforth, both PLL types induce apoptosis, but the early exposure of PS is the result of plasma membrane damage and phospholipid re-shuffling subsequent electrostatic interaction with PLL and PLL–DNA complexes; early PS expression was not affected in the presence of Ac-DVD and therefore is autonomous of caspase 3 activation. Here, it has been used PLL–DNA complexes that express positive zeta potential values (PLL: DNA of 3:1), thus impersonating previous transfection protocols. Mitochondria are whispered to be a central supervisory element in stress-induced cell death. Stress-induced inducements trigger mitochondrial permeabilization and cause the announcement of proapoptotic proteins such as Cytc and Smac/DIABLO from mitochondrial intermembrane space [75, 76]. The results in show significant

accumulation of Cytc in Jurkat T-cell cytoplasm at 24 h post-PLL (both in free form as well as DNA-bound) treatment. However, more Cytc was released with H-PLL treatment than L-PLL [77, 78]. In the case of H-PLL, the extent of Cytc release was decreased by approximately 50% in the presence of a cell-permeable Bax channel blocker [79], but interestingly inhibition of Bax channel-forming activity had no effect on Cyt *c* release from mitochondria by L-PLL. Similar observations were made with both free and DNA-bound PLLs. As a control experiment, Bcl-2 inhibition [80] accelerated cell death in all cases. Therefore, it has been concluding that H-PLL and L-PLL initiate mitochondrial-mediated apoptosis differently. In line with increased cytoplasmic Cytc accumulation and concomitant with caspase-3 activation, they also detected significant caspase-9 activity thus confirming the involvement of mitochondrially mediated apoptotic pathway. Indeed, MMP in Jurkat T-cells was dramatically altered in a time-dependent manner. Nuclear accumulation of PLL–DNA complexes were previously suggested to occur within a few hours of incubation [81, 82], where PLL may induce genotoxic stress. Cytc release from mitochondria by L-PLL and H-PLL (L-PLL-mediated Bax-independent and partially Bax-dependent H-PLL triggered Cytc release), they turned the attention

towards the direct effect of PLLs on mitochondrial membrane permeabilization and physiological functions, using freshly isolated rat liver mitochondria. H-PLL induced a progressive decline in the rate of DNP uncoupled respiration and partial mitochondrial depolarization. H-PLL may interact directly with the outer mitochondrial membrane, or at contact sites between the outer and inner mitochondrial membranes, where Bcl-2 appears to cluster [83]; this could modulate the concentration or distribution of cardiolipin, hence facilitating Cyt_c release directly, altering mitochondrial energization, which enhances Bax docking [84], and subsequent Bax-mediated mitochondrial destabilization/ permeabilization. Nevertheless, these studies further confirm as to why cell treatment with H-PLL induces significantly more caspase-3 activation than that of L-PLL. Intriguingly, observations of significantly higher Cyt_c release from isolated mitochondria were restricted to H-PLL. The endogenous polyamine spermine was suggested to facilitate the release of a small fraction of Cyt_c from isolated mitochondria through an exchangeable subpopulation of this protein and without incurring mitochondrial damage [72], but in light of some observations in intact cells, this mode of exit cannot explain L-PLL-induced Cyt_c release. However, PLLs and polyarginine are known to stimulate several

cellular kinases and protein phosphatases, particularly phospholipases, even at sub-optimal cytotoxic concentrations [70]. For instance, PLL was shown recently to induce phospholipase D activation in bovine pulmonary artery endothelial cells through the involvement of protein kinase C [70]. Thus, phospholipase activation, presumably through PLL-mediated plasma membrane damage and involvement of protein kinases, may lead towards the insertion of lysolipids into the outer mitochondrial membrane via recently identified lipid transfer capacity of full-length Bid [85]. Indeed, monolysocardiolipin is an inhibitor of mitochondrial phospholipase A2 [86], thus tight binding of full-length Bid to monolysocardiolipin may lead to activation of this phospholipase too. These modifications could collectively alter the physical state of the outer mitochondrial membrane (e.g., a positive change in membrane curvature) leading to the release of a small pool of apoptogenic factors, either directly or through other mediators including PLL. Certainly, this believable mechanism may be functioning with both PLL types, but the rationality of this hypothesis remnant to be verified when mutant cells become available. H-PLL and L-PLL are not only cytotoxic through plasma membrane damage, but moreover, they are accomplished of initiating

apoptosis inversely via the mitochondrial pathway. In the absence of endosomotropic agents, PLL and PLL–DNA complexes could induce late-phase mitochondrial-mediated apoptosis, thus substantiating their cytoplasmic attendance at around 24 h post-challenge. It would appear rational to accomplish that the induction of PLL-induced apoptosis could be contingent on a number of factors, including the cell type that is being pretentious, the intracellular levels of cysteine proteases and Bcl-2-family of proteins, and whether polyamines can afterward induce marked changes in the expression of pro- and anti-apoptotic genes since some PLL–DNA complexes are supposed to localize in the nucleus [81]. As poly (ethyleneglycol)-conjugated PLL [66] and PLL-grafted imidazoleacetic acid [87] can also induce apoptosis in normal and various tumor-derived human cell lines. Understanding of polyamine/polycation-mediated apoptosis is of serious prominence for the rational design of non-toxic delivery vectors for DNA compaction and transfection protocols, predominantly by retaining high-throughput methods [88, 89].

Freebase lysine increases survival and reduces metastasis by altering the tumor microenvironment

Tumor cells are highly glycolytic uniform in the presence of oxygen and henceforth produce free protons (H^+) at a higher rate than normal cells, a spectacle known as the

Warburg effect [90]. As importantly, the microenvironment of solid tumors is acidic and expressively affects tumor growth and invasion. Low extracellular pH principals to the improved statement of Cathepsin B and other proteolytic enzymes that consequence in the dilapidation of the extracellular matrix (ECM) [91, 92]. Fascinatingly tumor cells are comparatively resistant to acidic pH which is most likely due to mutations of the p53 tumor suppressor gene or additional mechanisms of the apoptotic pathway [92]. These explanations have led to the acid-mediated invasion hypothesis, which suggests that H^+ flow movement along absorptions gradients from the tumor into the peritumoral normal tissue instigating normal cell death and ECM degradation. Cancer cells, which are acid-adapted, are then able to invade into the injure neighboring normal tissue. Acidic pH has been exposed to pre-position cancers to augmented aggressive and metastatic phenotypes in animal models. An acquaintance of tumor cells to acidic growth circumstances preceding to intravascular injection considerably upsurges their competence to metastasize [93]. Here scientists investigate the impending role of lysine, a freely obtainable amino acid with a pKa of 10, in buffering tumors and plummeting metastases. In part, this is due to districts of

inadequate perfusion foremost to inadequate oxygen supply (hypoxia), which necessitates the up-regulation of glycolysis to preserve ATP levels. Although the glycolytic phenotype is supposed to be a near-universal phenomenon in cancer cells, it has not been precisely labeled in PC3M. Therefore, It has been first examined the metabolic state of PC3M cells in assessment to a normal prostate cell line (PCS) *in-vitro*. Based on the *in-vitro* results; it has been scrutinized the prospective role of a systemic buffer, lysine, in hindering the expansion of metastases from the PC3M cell *in-vivo*. The researchers found, dependable with our estimates, that lysine considerably prevents the progress of metastases and prolongs endurance. pH plays a significant role in virtually all steps of metastasis [94, 95] and metastatic disease in prostate cancer is consistently fatal. Numerous clarifications and mechanisms could subsidize to the

consequence of buffers on tumor metastasis. Acid-mediated incursion can occur via obliteration of the extracellular matrix, which is endorsed by proteases and glycosidases. Metalloproteinases (MMP-2 and MMP-9) are supposed to be perilous for invasion and extravasation [96, 97]. MMPs are a family of proteolytic enzymes that reduce the extracellular matrix and junctional proteins and further upsurge endothelial permeability [96]. Low pH up-regulates angiogenic factors such as vascular endothelial growth factor (VEGF) and interleukin eight (IL-8) motivating neo-vascularization and indorsing metastasis. Thus, it is conceivable that lysine can converse acidosis and subsequently reduce proteolytic enzyme activity and or angiogenesis, which will central to the reticence of extravasations, and colonization of circulating tumor cells reducing efficacious metastasis (Table 3).

Table 3: Various mechanisms of cancer cell death by Lysine

Types of mechanisms by lysine for apoptosis	References
Regulates TNF- α and MMP-3 expression	[141][142][143][144][145][146][147][148][149][69][150][151][152][153][154][155][156][157][158][151][152][153][154][75][77]
sMitochondrial-mediated apoptosis	[12][121][79][80][81][82][159][84][85][86][87][88][89].
Reduces metastasis by altering the tumor microenvironment	[105][106][160]

CONCLUSION

It is apparent that amino acid treatment against cancer will be in the majority in the 21st century. Thus, the discovery of the efficacy of L-lysine will certainly show its effectiveness against tumors more than the other amino acids. The biggest advantage

that we have that Lysine can be cytotoxic, apoptotic against cancer cells in a proper dose-dependent manner, which alters the metabolism of the cancer cell. On the other hand, it also shows its efficacy in several modes of application. Being single or in derived from and a combinational form,

lysine proves itself to be target-specific, cytotoxic against cancer cells sparing its normal counterparts (Figure 1). However, if it could be coated through nanoparticles then it might show their effectiveness in a more defined way and lower doses. Therefore, further research on the efficacy of Lysine is to be made so that to explore a

new horizon in the field of anticancer research. Thus, this review depicts several efficacies of L-Lysine, which might show a new hope of using Lysine in a broader spectrum. More clinical trials have to be made for proper evaluation of lysine in the battle over cancer then only it can provide new hope against anticancer drug development.

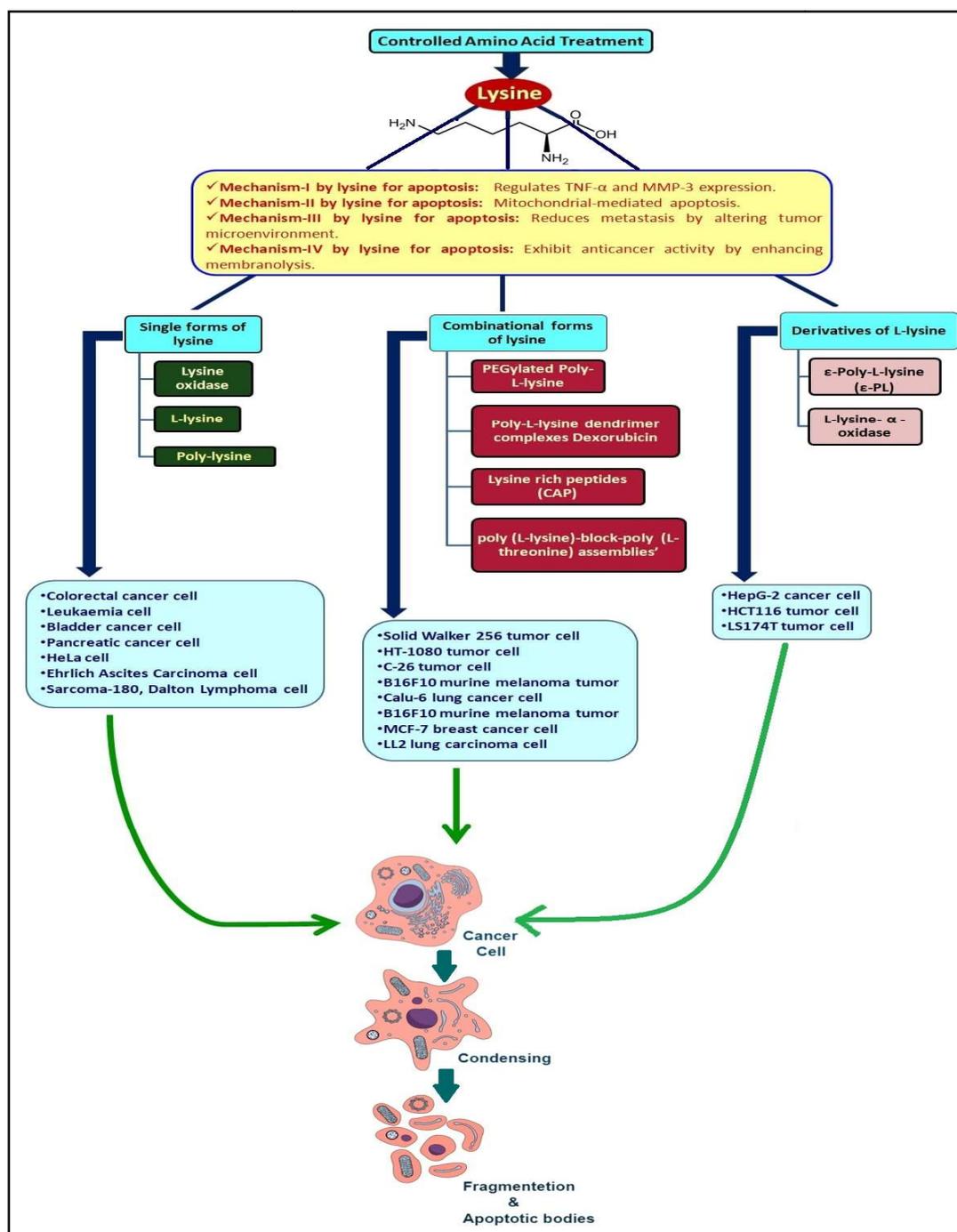


Figure 1: Effectiveness of L-lysine triggering the cancer cell death

Conflict of Interest

The authors declare no conflict of interest.

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REFERENCE

- [1] J. R. Cantor and D. M. Sabatini, "Cancer cell metabolism: One hallmark, many faces," *Cancer Discovery*. 2012.
- [2] P. S. Ward and C. B. Thompson, "Metabolic Reprogramming: A Cancer Hallmark Even Warburg Did Not Anticipate," *Cancer Cell*. 2012.
- [3] D. Hanahan and R. A. Weinberg, "Hallmarks of cancer: The next generation," *Cell*. 2011.
- [4] C. Commisso *et al.*, "Macropinocytosis of protein is an amino acid supply route in Ras-transformed cells," *Nature*, 2013.
- [5] P. S. Ward and C. B. Thompson, "Metabolic Reprogramming: A Cancer Hallmark Even Warburg Did Not Anticipate," *Cancer Cell*, vol. 21, no. 3, pp. 297–308, 2012.
- [6] L. J. Arnold, A. Dagan, J. Gutheil, and N. O. Kaplan, "Antineoplastic activity of poly(L-lysine) with some ascites tumor cells.," *Proc. Natl. Acad. Sci.*, 1979.
- [7] J. KAWADA, Y. YOSHIMURA, and T. MINAMI, "Some Properties of Thyroidal Membrane Adenosinetriphosphatase and Iodide Uptake: Effects of Basic Polyamino Acids," *Endocrinol. Jpn.*, vol. 23, no. 3, pp. 221–225, 2011.
- [8] L. J. Anghileri, M. Heidbreder, and R. Mathes, "Accumulation of ⁵⁷Co-poly-L-lysine by tumors: an effect of the tumor electrical charge.," *J. Nucl. Biol. Med.*, vol. 20, no. 2, pp. 79–83, 1976.
- [9] S. E. Kornguth and M. A. Stahmann, "Effect of Polylysine on the Leakage and Retention of Compounds by Ehrlich Ascites Tumor Cells," *Cancer Res.*, 1961.
- [10] S. Debnath, S. Karan, T. K. Chatterjee, M. Debnath, and J. Dash, "Poly-L-lysine inhibits tumor angiogenesis and induces apoptosis in ehrlich ascites carcinoma and in sarcoma s-180 tumor.," *Asian Pacific J. Cancer Prev.*, vol. 18, no. 8, pp. 2255–2268, 2017.
- [11] K. Ogawa and A. Ichihara, "Inhibition of protein synthesis in mouse L cells by poly-L-ornithine," *J. Biochem.*, vol. 83, no. 2, pp. 519–525, 1978.
- [12] D. Duksin, E. Katchalski, and L. Sachs, "Specific Aggregation of SV40-Transformed Cells by Ornithine, Leucine Copolymers," *Proc. Natl. Acad. Sci.*, vol. 67, no. 1, pp. 185–192, 1970.
- [13] L. J. Arnold, A. Dagan, J. Gutheil, and N. O. Kaplan, "Antineoplastic activity of poly(L-lysine) with some ascites tumor cells.," *Proc. Natl. Acad. Sci.*, vol. 76, no. 7, pp. 3246–3250, 1979.
- [14] L. M. Kaminskas *et al.*, "Pharmacokinetics and tumor disposition of PEGylated, methotrexate

- conjugated poly-L-lysine dendrimers,” *Mol. Pharm.*, vol. 6, no. 4, pp. 1190–1204, 2009.
- [15] M. E. Fox, S. Guillaudeu, J. M. J. Fréchet, K. Jerger, N. Macaraeg, and F. C. Szoka, “Synthesis and *In Vivo* Antitumor Efficacy of PEGylated Poly(L-lysine) Dendrimer–Camptothecin Conjugates,” *Mol. Pharm.*, vol. 6, no. 5, pp. 1562–1572, Oct. 2009.
- [16] V. P. Torchilin *et al.*, “Cationic charge determines the distribution of liposomes between the vascular and extravascular compartments of tumors,” *Cancer Res.*, vol. 62, no. 23, pp. 6831–6, 2002.
- [17] M. Eichhorn, S. Strieth, S. Krasnici, B. Sauer, M. T.- Angiogenesis, and U. 2004, “Protamine enhances uptake of cationic liposomes in angiogenic microvessels of solid tumours,” *Angiogenesis*, vol. 7, no. 2, pp. 133–141, 2004.
- [18] M. Dellian, B. P. Witwer, H. A. Salehi, F. Yuan, and R. K. Jain, “Quantitation and physiological characterization of angiogenic vessels in mice: Effect of basic fibroblast growth factor, vascular endothelial growth factor/vascular permeability factor, and host microenvironment,” *Am. J. Pathol.*, vol. 149, no. 1, pp. 59–71, 1996.
- [19] M. Leunig *et al.*, “Microhemodynamics, and Interstitial Fluid Pressure during Early Growth of Human Adenocarcinoma LS174T in SCID Mice in SCID Mice 1,” *Cancer Res.*, vol. 52, pp. 6553–6560, 1992.
- [20] B. Ren, N. Hoti, X. Rabasseda, Y. Wang, and M. Wu, “The antiangiogenic and therapeutic implications of endostatin,” *Methods Find. Exp. Clin. Pharmacol.*, vol. 25, no. 3, p. 215, 2005.
- [21] K. T. Al-Jamal *et al.*, “Systemic antiangiogenic activity of cationic poly-L-lysine dendrimer delays tumor growth,” *Proc. Natl. Acad. Sci.*, vol. 107, no. 9, pp. 3966–3971, 2010.
- [22] K. T. Al-Jamal *et al.*, “Cationic poly-L-Lysine dendrimer complexes doxorubicin and delays tumor growth in vitro and in vivo,” *ACS Nano*, vol. 7, no. 3, pp. 1905–1917, Mar. 2013.
- [23] K. T. Al-Jamal *et al.*, “Cationic poly-L-Lysine dendrimer complexes doxorubicin and delays tumor growth in vitro and in vivo,” *ACS Nano*, vol. 7, no. 3, pp. 1905–1917, Mar. 2013.
- [24] M. W. Roomi, V. Ivanov, T. Kalinovsky, A. Niedzwiecki, and M. Rath, “Antitumor effect of ascorbic acid, lysine, proline, arginine, and green tea extract on bladder cancer cell line T-24,” *Int. J. Urol.*, vol. 13, no. 4, pp. 415–419, Apr. 2006.
- [25] H. F. Liao *et al.*, “Inhibitory Effect of Caffeic Acid Phenethyl Ester on Angiogenesis, Tumor Invasion, and Metastasis,” *J. Agric. Food Chem.*, vol. 51, no. 27, pp. 7907–7912, Dec. 2003.
- [26] M. J. Duffy, “The role of proteolytic enzymes in cancer invasion and metastasis,” *Clin. Exp. Metastasis*, vol. 10, no. 3, pp. 145–155, May 1992.
- [27] J. E. Jung *et al.*, “Caffeic acid and its

- synthetic derivative CADPE suppress tumor angiogenesis by blocking STAT3-mediated VEGF expression in human renal carcinoma cells,” *Carcinogenesis*, vol. 28, no. 8, pp. 1780–1787, 2007.
- [28] S. P. Netke, M. W. Roomi, V. Ivanov, and M. Rath, “A Specific Combination Of Ascorbic Acid , Lysine , Proline And Epigallocatechin Gallate Inhibits Proliferation And Extracellular Matrix Invasion Of Various Human Cancer Cell Lines,” *Emerg. Drugs*, pp. 1–11, 2003.
- [29] M.-R. Ahn *et al.*, “Suppression of tumor-induced angiogenesis by Brazilian propolis: Major component artepillin C inhibits in vitro tube formation and endothelial cell proliferation,” *Cancer Lett.*, vol. 252, no. 2, pp. 235–243, 2007.
- [30] M. W. Roomi, V. Ivanov, T. Kalinovsky, A. Niedzwiecki, and M. Rath, “Anti-tumor effect of ascorbic acid, lysine, proline, arginine, and epigallocatechin gallate on prostate cancer cell lines PC-3, LNCAP, and DU 145,” *Res. Commun. Mol. Pathol. Pharmacol.*, vol. 115–116, no. 2, pp. 251–264, 2004.
- [31] N. Weidner, “Intratumor microvessel density as a prognostic factor in cancer.,” *Am. J. Pathol.*, vol. 147, no. 1, pp. 9–19, 1995.
- [32] M. W. Roomi, V. Ivanov, T. Kalinovsky, A. Niedzwiecki, and M. Rath, “In vivo antitumor effect of ascorbic acid, lysine, proline and green tea extract on human prostate cancer PC-3 xenografts in nude mice: Evaluation of tumor growth and immunohistochemistry,” *In Vivo (Brooklyn)*, vol. 19, no. 1, pp. 179–184, Mar. 2005.
- [33] C. A. Dornelas *et al.*, “Angiogenesis inhibition by green propolis and the angiogenic effect of L-lysine on bladder cancer in rats,” *Acta Cir. Bras.*, vol. 27, no. 8, pp. 529–536, 2012.
- [34] M. W. Roomi, N. W. Roomi, V. Ivanov, T. Kalinovsky, A. Niedzwiecki, and M. Rath, “Modulation of N-methyl-N-nitrosourea induced mammary tumors in Sprague-Dawley rats by combination of lysine, proline, arginine, ascorbic acid and green tea extract,” *Breast Cancer Res.*, vol. 7, no. 3, p. R291, Jun. 2005.
- [35] M. W. Roomi, V. Ivanov, T. Kalinovsky, A. Niedzwiecki, and M. Rath, “Antitumor Effect of a Combination of Lysine, Proline, Arginine, Ascorbic Acid, and Green Tea Extract on Pancreatic Cancer Cell Line MIA PaCa-2,” *Int. J. Gastrointest. Cancer*, vol. 35, no. 2, pp. 097–102, 2005.
- [36] C. Shi, X. Zhao, Z. Liu, R. Meng, X. Chen, and N. Guo, “Antimicrobial, antioxidant, and antitumor activity of epsilon-poly-L-lysine and citral, alone or in combination,” *Food Nutr. Res.*, vol. 60, no. 1, p. 31891, Jan. 2016.
- [37] J. Hiraki, “epsilon-Polylysine, its development and utilization.,” *Fine*

- Chem.*, vol. 29, pp. 18–25, 2000.
- [38] I. Geornaras, Y. Yoon, K. E. Belk, G. C. Smith, and J. N. Sofos, “Antimicrobial activity of ϵ -polylysine against *Escherichia coli* O157:H7, *Salmonella typhimurium*, and *Listeria monocytogenes* in various food extracts,” *J. Food Sci.*, vol. 72, no. 8, pp. M330–M334, Oct. 2007.
- [39] T. Greten, F. Papendorf, J. Bleck, ... T. K.-B. journal of, and U. 2005, “Survival rate in patients with hepatocellular carcinoma: A retrospective analysis of 389 patients,” *Br. J. Cancer*, vol. 92, no. 10, pp. 1862–1868, 2005.
- [40] B. Szende *et al.*, “Antitumor effect of lysine-isopeptides,” *Cancer Cell Int.*, vol. 2, 2002.
- [41] E. V. Lukasheva and T. T. Berezov, “L-lysine α -oxidase: Physicochemical and biological properties,” *Biochemistry (Moscow)*, vol. 67, no. 10. pp. 1152–1158, 2002.
- [42] H. Kusakabe, K. Kodama, A. Kuninaka, and ... H. Y., “A new antitumor enzyme, L-lysine α -oxidase from *Trichoderma viride*: Purification and enzymological properties,” *J. Biol. Chem.*, vol. 255, no. 3, pp. 976–981, 1980.
- [43] H. Kusakabe, K. Kodama, A. Kuninaka, H. Yoshino, and K. Soda, “Effect of L-lysine α -oxidase on growth of mouse leukemic cells,” *Agric. Biol. Chem.*, vol. 44, no. 2, pp. 387–392, 1980.
- [44] N. Y. Anisimova, “Cloning, expression and characterization of the recombinant *Yersinia pseudotuberculosis* l-asparaginase,” *Protein Expr. Purif.*, vol. 82, no. 1, pp. 150–154, 2012.
- [45] M. KOZAI, E. SASAMORI, M. FUJIHARA, T. YAMASHITA, H. TAIRA, and R. HARASAWA, “Growth Inhibition of Human Melanoma Cells by a Recombinant Arginine Deiminase Expressed in *Escherichia coli*,” *J. Vet. Med. Sci.*, vol. 71, no. 10, pp. 1343–1347, 2009.
- [46] C. Schulenburg *et al.*, “The interdependence between catalytic activity, conformational stability, and cytotoxicity of onconase,” *Cancer Biol. Ther.*, vol. 6, no. 8, pp. 1233–1239, Aug. 2007.
- [47] V. S. Pokrovsky *et al.*, “Enzymatic properties and anticancer activity of L-lysine α -oxidase from *Trichoderma cf. aureoviride* Rifai BKMF-4268D,” *Anticancer. Drugs*, vol. 24, no. 8, pp. 846–851, 2013.
- [48] M. Singh *et al.*, “Medicinal uses of L-Lysine: Past and future,” *International Journal of Research in Pharmaceutical Sciences*, vol. 2, no. 4. pp. 637–642, 2011.
- [49] M. SMRIGA, T. ANDO, M. AKUTSU, Y. FURUKAWA, K. MIWA, and Y. MORINAGA, “Oral treatment with L-lysine and L-arginine reduces anxiety and basal cortisol levels in healthy humans,” *Biomed. Res.*, vol. 28, no. 2, pp. 85–90, 2007.
- [50] S. M., M. R. D., P. S., B. S., M. K., and R. D. K., “Medicinal uses of L-Lysine:

- Past and future,” *Int. J. Res. Pharm. Sci.*, vol. 2, no. 4, pp. 637–642, 2011.
- [51] Y. Mine and H. Zhang, “Anti-inflammatory Effects of Poly-L-lysine in Intestinal Mucosal System Mediated by Calcium-Sensing Receptor Activation,” *J. Agric. Food Chem.*, vol. 63, no. 48, pp. 10437–10447, Dec. 2015.
- [52] Y. JIA, R. HU, B. K.-C. J. of E. Traditional, and U. 2004, “Study on Anti-inflammatory Effects of the Complex (LaiJinsi) of EHPL and Lysine Hydrochloride,” *en.cnki.com.cn*.
- [53] K. Dudek, “Role of pro-inflammatory cytokines in the etiology of osteoarthritis,” *Med. Weter.*, vol. 60, no. 10, pp. 1039–1041, 2004.
- [54] C. Westacott, A. Barakat, L. Wood, M. P.-O. and, and U. 2000, “Tumor necrosis factor alpha can contribute to focal loss of cartilage in osteoarthritis,” *Osteoarthr. Cartil.*, vol. 8, no. 3, pp. 213–221, 2000.
- [55] T. John, R. Müller, A. Oberholzer, H. Zreiqat, B. K.- Cytokine, and U. 2007, “Interleukin-10 modulates pro-apoptotic effects of TNF- α in human articular chondrocytes in vitro,” *Cytokine*, vol. 40, no. 3, pp. 226–234, 2007.
- [56] R. Müller, T. John, B. Kohl, A. Oberholzer, T. G.- Cytokine, and U. 2008, “IL-10 overexpression differentially affects cartilage matrix gene expression in response to TNF- α in human articular chondrocytes in vitro,” *Cytokine*, vol. 44, no. 3, pp. 377–385, 2008.
- [57] J. Zwerina et al., “TNF-induced structural joint damage is mediated by IL-1,” *Proc. Natl. Acad. Sci.*, vol. 104, no. 28, pp. 11742–11747, 2007.
- [58] J. Martel-Pelletier, F. Mineau, F. C. Jolicoeur, J. M. Cloutier, and J. P. Pelletier, “In vitro effects of diacerhein and rheim on IL-1 and TNF-alpha systems in human osteoarthritic tissues,” *J. Rheumatol.*, vol. 25, pp. 753-762., 1998.
- [59] G. Moulis et al., “Cancer Risk of Anti-TNF- α at Recommended Doses in Adult Rheumatoid Arthritis: A Meta-Analysis with Intention to Treat and per Protocol Analyses,” *PLoS One*, vol. 7, no. 11, p. e48991, Nov. 2012.
- [60] S. Debnath, S. Karan, T. K. Chatterjee, M. Debnath, and J. Dash, “Poly-L-lysine inhibits tumor angiogenesis and induces apoptosis in ehrlich ascites carcinoma and in sarcoma s-180 tumor,” *Asian Pacific J. Cancer Prev.*, vol. 18, no. 8, pp. 2255–2268, 2017.
- [61] S. Debnath, A. Mukherjee, S. Karan, M. Debnath, and T. K. Chatterjee, “Induction of apoptosis, anti-proliferation, tumor-angiogenic suppression and down-regulation of Dalton’s Ascitic Lymphoma (DAL) induced tumorigenesis by poly-L-lysine: A mechanistic study,” *Biomed. Pharmacother.*, vol. 102, pp. 1064–1076, 2018.
- [62] G. Y. Wu and C. H. Wu, “Receptor-mediated in vitro gene transformation

- by a soluble DNA carrier system,” *J. Biol. Chem.*, vol. 262, no. 10, pp. 4429–4432, 1987.
- [63] W. C. M., R. M. L., and S. L. W., “Systemic Circulation of Poly(L-lysine)/DNA Vectors is Influenced by Polycation Molecular Weight and Type of DNA: Differential Circulation in Mice and Rats and the Implications for Human Gene Therapy,” *Blood*, vol. 97, no. 8, p. 2221, 2001.
- [64] M. A. Wolfert *et al.*, “Polyelectrolyte vectors for gene delivery: Influence of cationic polymer on biophysical properties of complexes formed with DNA,” *Bioconjug. Chem.*, vol. 10, no. 6, pp. 993–1004, Nov. 1999.
- [65] M. Ohsaki, T. Okuda, A. Wada, T. Hirayama, T. Niidome, and H. Aoyagi, “In vitro gene transfection using dendritic poly(L-lysine),” *Bioconjug. Chem.*, vol. 13, no. 3, pp. 510–517, May 2002.
- [66] C. M. Ward, M. Pechar, D. Oupicky, K. Ulbrich, and L. W. Seymour, “Modification of pLL/DNA complexes with a multivalent hydrophilic polymer permits folate-mediated targeting in vitro and prolonged plasma circulation in vivo,” *J. Gene Med.*, vol. 4, no. 5, pp. 536–547, Sep. 2002.
- [67] M. L. Forrest and D. W. Pack, “On the kinetics of polyplex endocytic trafficking: Implications for gene delivery vector design,” *Mol. Ther.*, vol. 6, no. 1, pp. 57–66, 2002.
- [68] S. M. Moghimi, P. Symonds, J. C. Murray, A. C. Hunter, G. Debska, and A. Szewczyk, “A two-stage poly(ethylenimine)-mediated cytotoxicity: Implications for gene transfer/therapy,” *Mol. Ther.*, vol. 11, no. 6, pp. 990–995, 2005.
- [69] W. T. Shier, D. J. Dubourdieu, and J. P. Durkin, “Polycations as prostaglandin synthesis inducers. Stimulation of arachidonic acid release and prostaglandin synthesis in cultured fibroblasts by poly(L-lysine) and other synthetic polycations,” *Biochim. Biophys. Acta (BBA)/Lipids Lipid Metab.*, vol. 793, no. 2, pp. 238–250, 1984.
- [70] S. Vepa, W. M. Scribner, and V. Natarajan, “Activation of endothelial cell phospholipase D by polycations,” *Am. J. Physiol. Cell. Mol. Physiol.*, vol. 272, no. 4, pp. L608–L613, Apr. 2017.
- [71] C. Stefanelli *et al.*, “Spermine triggers the activation of caspase-3 in a cell-free model of apoptosis,” *FEBS Lett.*, vol. 451, no. 2, pp. 95–98, May 1999.
- [72] C. STEFANELLI *et al.*, “Polyamines directly induce release of cytochrome c from heart mitochondria,” *Biochem. J.*, vol. 347, no. 3, pp. 875–880, 2000.
- [73] S. J. Martin, D. M. Finucane, G. P. Amarante-Mendes, G. A. O’Brien, and D. R. Green, “Phosphatidylserine externalization during CD95-induced apoptosis of cells and cytoplasts requires ICE/CED-3 protease activity,” *J. Biol. Chem.*, vol. 271, no. 46, pp. 28753–28756, 1996.

- [74] S. J. Martin, "Early redistribution of plasma membrane phosphatidylserine is a general feature of apoptosis regardless of the initiating stimulus: inhibition by overexpression of Bcl-2 and Abl," *J. Exp. Med.*, vol. 182, no. 5, pp. 1545–1556, 1995.
- [75] M. O. Hengartner, "The biochemistry of apoptosis," *Nature*, vol. 407, no. 6805, pp. 770–776, 2000.
- [76] R. A. Gottlieb, "Mitochondria and apoptosis," *NeuroSignals*, vol. 10, no. 3–4, pp. 147–161, 2001.
- [77] D. R. Green and J. C. Reed, "线粒体和细胞凋亡 Mitochondria and apoptosis.," *Science*, vol. 281, no. 5381, pp. 1309–12, 1998.
- [78] D. D. Newmeyer, S. Ferguson-miller, S. Diego, and B. Building, "Mitochondria: Releasing Power for Life and Unleashing the Machineries of Death We apologize for an error in citation in," *Cell*, vol. 112, no. 1988, p. 2003, 2003.
- [79] A. Bombrun, P. Gerber, G. Casi, O. Terradillos, B. Antonsson, and S. Halazy, "3,6-Dibromocarbazole piperazine derivatives of 2-propanol as first inhibitors of cytochrome C release via bax channel modulation," *J. Med. Chem.*, vol. 46, no. 21, pp. 4365–4368, Oct. 2003.
- [80] I. J. Enyedy et al., "Discovery of small-molecule inhibitors of Bcl-2 through structure-based computer screening," *J. Med. Chem.*, vol. 44, no. 25, pp. 4313–4324, Dec. 2001.
- [81] K. Fisher, K. Ulbrich, V. Subr, C. Ward, V. M.-G. Therapy, and U. 2000, "A versatile system for receptor-mediated gene delivery permits increased entry of DNA into target cells, enhanced delivery to the nucleus and elevated rates of transgene expression," *Gene Ther.*, vol. 7, no. 15, pp. 1337–1343, 2000.
- [82] S. Grosse, A. Tremeau-Bravard, Y. Aron, P. B.-G. Therapy, and U. 2002, "Intracellular rate-limiting steps of gene transfer using glycosylated polylysines in cystic fibrosis airway epithelial cells," *Gene Ther.*, vol. 9, no. 15, pp. 1000–1007, 2002.
- [83] J. C. Reed, S. Tanaka, S. Takayama, M. J. Schibler, and W. Fenton, "Investigation of the Subcellular Distribution of the bcl-2 Oncoprotein: Residence in the Nuclear Envelope, Endoplasmic Reticulum, and Outer Mitochondrial Membranes," *Cancer Res.*, vol. 53, no. 19, pp. 4701–4714, 1993.
- [84] S. Smaili, Y. Hsu, K. Sanders, ... J. R.-C. death and, and U. 2001, "Bax translocation to mitochondria subsequent to a rapid loss of mitochondrial membrane potential," *Cell Death Differ.*, vol. 8, no. 9, pp. 909–920, 2001.
- [85] A. GOONESINGHE, E. S. MUNDY, M. SMITH, R. KHOSRAVI-FAR, J.-C. MARTINO, and M. D. ESPOSTI, "Pro-apoptotic Bid induces membrane

- perturbation by inserting selected lysolipids into the bilayer,” *Biochem. J.*, vol. 387, no. 1, pp. 109–118, 2005.
- [86] M. Reers and D. R. Pfeiffer, “Inhibition of Mitochondrial Phospholipase A2 by Mono- and Dilyscardiolipin,” *Biochemistry*, vol. 26, no. 25, pp. 8038–8041, Dec. 1987.
- [87] D. Putnam, C. A. Gentry, D. W. Pack, and R. Langer, “Polymer-based gene delivery with low cytotoxicity by a unique balance of side-chain termini,” *Proc. Natl. Acad. Sci.*, vol. 98, no. 3, pp. 1200–1205, 2012.
- [88] D. G. Anderson, A. Akinc, N. Hossain, and R. Langer, “Structure/property studies of polymeric gene delivery using a library of poly(β -amino esters),” *Mol. Ther.*, vol. 11, no. 3, pp. 426–434, 2005.
- [89] P. Symonds, J. C. Murray, A. C. Hunter, G. Debska, A. Szewczyk, and S. M. Moghimi, “Low and high molecular weight poly(L-lysine)s/poly(L-lysine)-DNA complexes initiate mitochondrial-mediated apoptosis differently,” *FEBS Lett.*, vol. 579, no. 27, pp. 6191–6198, Nov. 2005.
- [90] Otto Warburg, “On the Origin of Cancer Cells,” *Science (80-.)*, vol. 123, no. 3191, pp. 309–314, 1956.
- [91] P. Montcourrier et al., “Characterization of very acidic phagosomes in breast cancer cells and their association with invasion,” *J. Cell Sci.*, vol. 107 (Pt 9, pp. 2381–91, 1994.
- [92] C. Cuvier, A. Jang, and R. P. Hill, “Exposure to hypoxia, glucose starvation and acidosis: Effect on invasive capacity of murine tumor cells and correlation with cathepsin (L + B) secretion,” *Clin. Exp. Metastasis*, vol. 15, no. 1, pp. 19–25, 1997.
- [93] R. E.K., M. B., K. K., and G. K., “Acidic extracellular pH promotes experimental metastasis of human melanoma cells in athymic nude mice,” *Cancer Res.*, vol. 66, no. 13, pp. 6699–6707, 2006.
- [94] A. I. Hashim, X. Zhang, J. W. Wojtkowiak, G. V. Martinez, and R. J. Gillies, “Imaging pH and metastasis,” *NMR in Biomedicine*, vol. 24, no. 6. pp. 582–591, 2011.
- [95] mahsa, “Breast cancer metastasis: markers and models. Nature reviews cancer,” *breast*, 2005.
- [96] J. H. Nguyen et al., “Matrix metalloproteinase-9 contributes to brain extravasation and edema in fulminant hepatic failure mice,” *J. Hepatol.*, vol. 44, no. 6, pp. 1105–1114, 2006.
- [97] A. I.- Hashim, J. W. W. Ojtkowiak, and M. de L. C. Ribeiro, “Free Base Lysine Increases Survival and Reduces Metastasis in Prostate Cancer Model,” *J. Cancer Sci. Ther.*, vol. 01, no. 01, 2011.
- [98] P. Gao et al., “C-Myc suppression of miR-23a/b enhances mitochondrial glutaminase expression and glutamine metabolism,” *Nature*, vol. 458, no. 7239, pp. 762–765, 2009.
- [99] R. Possemato et al., “Functional genomics reveal that the serine synthesis pathway is essential in breast cancer,”

- Nature*, vol. 476, no. 7360, pp. 346–350, 2011.
- [100] C. Link, “Phosphoglycerate dehydrogenase diverts glycolytic flux and contributes to oncogenesis Citation Accessed,” *nature.com*, 2013.
- [101] J. Ye *et al.*, “Serine catabolism regulates mitochondrial redox control during hypoxia,” *Cancer Discov.*, vol. 4, no. 12, pp. 1406–1417, 2014.
- [102] D. Kim *et al.*, “SHMT2 drives glioma cell survival in ischaemia but imposes a dependence on glycine clearance,” *Nature*, vol. 520, no. 7547, pp. 363–367, 2015.
- [103] W. C. Zhang *et al.*, “Glycine decarboxylase activity drives non-small cell lung cancer tumor-initiating cells and tumorigenesis,” *Cell*, vol. 148, no. 1–2, pp. 259–272, 2012.
- [104] J. Zhang *et al.*, “Asparagine plays a critical role in regulating cellular adaptation to glutamine depletion,” *Mol. Cell*, vol. 56, no. 2, pp. 205–218, 2014.
- [105] B. C. Grabiner *et al.*, “A diverse array of cancer-associated MTOR mutations are hyperactivating and can predict rapamycin sensitivity,” *Cancer Discov.*, vol. 4, no. 5, pp. 554–563, 2014.
- [106] T. Sato, A. Nakashima, L. Guo, K. Coffman, and F. Tamanoi, “Single amino-acid changes that confer constitutive activation of mTOR are discovered in human cancer,” *Oncogene*, vol. 29, no. 18, pp. 2746–2752, 2010.
- [107] W. H. Fujita, R. J. Barr, and J. L. Headley, “Multiple Fibrofolliculomas With Trichodiscomas and Acrochordons,” *Arch. Dermatol.*, vol. 117, no. 1, pp. 32–35, 1981.
- [108] Z.-Y. Y, “The Folliculin Tumor Suppressor Is a GAP for the RagC/D GTPases That Signal Amino Acid Levels to mTORC1,” *Elsevier*, 2016.
- [109] M. L. Nickerson *et al.*, “Mutations in a novel gene lead to kidney tumors, lung wall defects, and benign tumors of the hair follicle in patients with the Birt-Hogg-Dubé syndrome,” *Cancer Cell*, vol. 2, no. 2, pp. 157–164, 2002.
- [110] T. Flower and T. O. F. Eat, “A Tumor Suppressor Complex with GAP Activity for the Rag GTPases That Signal,” *Science (80-.)*, no. May, pp. 1100–1106, 2013.
- [111] T. Richardson, J. Hodgett, A. Lindner, and M. A. Stahmann, “Action of Polylysine on Some Ascites Tumors in Mice,” *Exp. Biol. Med.*, vol. 101, no. 2, pp. 382–386, Jun. 2013.
- [112] J. KAWADA, Y. YOSHIMURA, and T. MINAMI, “Some Properties of Thyroidal Membrane Adenosinetriphosphatase and Iodide Uptake: Effects of Basic Polyamino Acids,” *Endocrinol. Jpn.*, vol. 23, no. 3, pp. 221–225, 2011.
- [113] J. E. Jung *et al.*, “Caffeic acid and its synthetic derivative CADPE suppress tumor angiogenesis by blocking STAT3-mediated VEGF expression in human renal carcinoma cells,” *Carcinogenesis*, vol. 28, no. 8, pp.

- 1780–1787, 2007.
- [114] S. P. Netke, M. W. Roomi, V. Ivanov, and M. Rath, “A Specific Combination Of Ascorbic Acid , Lysine , Proline And Epigallocatechin Gallate Inhibits Proliferation And Extracellular Matrix Invasion Of Various Human Cancer Cell Lines,” *Emerg. Drugs*, pp. 1–11, 2003.
- [115] M.-R. Ahn *et al.*, “Suppression of tumor-induced angiogenesis by Brazilian propolis: Major component artepillin C inhibits in vitro tube formation and endothelial cell proliferation,” *Cancer Lett.*, vol. 252, no. 2, pp. 235–243, 2007.
- [116] S. Brown, “Handling Chemical Carcinogens in the Laboratory--Problems of Safety,” *J. Clin. Pathol.*, vol. 34, no. 1, pp. 108–108, 2007.
- [117] C. A. Dornelas *et al.*, “Angiogenesis inhibition by green propolis and the angiogenic effect of L-lysine on bladder cancer in rats,” *Acta Cir. Bras.*, vol. 27, no. 8, pp. 529–536, 2012.
- [118] C. Shi, X. Zhao, Z. Liu, R. Meng, X. Chen, and N. Guo, “Antimicrobial, antioxidant, and antitumor activity of epsilon-poly-L-lysine and citral, alone or in combination,” *Food Nutr. Res.*, vol. 60, no. 1, p. 31891, Jan. 2016.
- [119] I. Geornaras, Y. Yoon, K. E. Belk, G. C. Smith, and J. N. Sofos, “Antimicrobial activity of ϵ -polylysine against *Escherichia coli* O157:H7, *Salmonella typhimurium*, and *Listeria monocytogenes* in various food extracts,” *J. Food Sci.*, vol. 72, no. 8, pp. M330–M334, Oct. 2007.
- [120] D. Duksin, E. Katchalski, and L. Sachs, “Specific Aggregation of SV40-Transformed Cells by Ornithine, Leucine Copolymers,” *Proc. Natl. Acad. Sci.*, vol. 67, no. 1, pp. 185–192, 1970.
- [121] D. D. Newmeyer, S. Ferguson-miller, S. Diego, and B. Building, “Mitochondria: Releasing Power for Life and Unleashing the Machineries of Death We apologize for an error in citation in,” *Cell*, vol. 112, no. 1988, p. 2003, 2003.
- [122] L. J. Arnold, A. Dagan, J. Gutheil, and N. O. Kaplan, “Antineoplastic activity of poly(L-lysine) with some ascites tumor cells,” *Proc. Natl. Acad. Sci.*, vol. 76, no. 7, pp. 3246–3250, 1979.
- [123] L. M. Kaminskas *et al.*, “Pharmacokinetics and tumor disposition of PEGylated, methotrexate conjugated poly-L-lysine dendrimers,” *Mol. Pharm.*, vol. 6, no. 4, pp. 1190–1204, Aug. 2009.
- [124] M. E. Eichhorn *et al.*, “Protamine enhances uptake of cationic liposomes in angiogenic microvessels of solid tumours,” *Angiogenesis*, vol. 7, no. 2, pp. 133–141, 2004.
- [125] M. Dellian, B. P. Witwer, H. A. Salehi, F. Yuan, and R. K. Jain, “Quantitation and physiological characterization of angiogenic vessels in mice: Effect of basic fibroblast growth factor, vascular endothelial growth factor/vascular permeability factor, and host

- microenvironment,” *Am. J. Pathol.*, vol. 149, no. 1, pp. 59–71, 1996.
- [126] M. Leunig *et al.*, “Microhemodynamics, and Interstitial Fluid Pressure during Early Growth of Human Adenocarcinoma LS174T in SCID Mice in SCID Mice 1,” *Cancer Res.*, vol. 52, pp. 6553–6560, 1992.
- [127] B. Ren, N. Hoti, X. Rabasseda, Y. Wang, and M. Wu, “The antiangiogenic and therapeutic implications of endostatin,” *Methods Find. Exp. Clin. Pharmacol.*, vol. 25, no. 3, p. 215, 2005.
- [128] H. F. Liao *et al.*, “Inhibitory Effect of Caffeic Acid Phenethyl Ester on Angiogenesis, Tumor Invasion, and Metastasis,” *J. Agric. Food Chem.*, vol. 51, no. 27, pp. 7907–7912, Dec. 2003.
- [129] M. J. Duffy, “The role of proteolytic enzymes in cancer invasion and metastasis,” *Clin. Exp. Metastasis*, vol. 10, no. 3, pp. 145–155, May 1992.
- [130] M. V. Pokrovskaya *et al.*, “Cloning, expression and characterization of the recombinant *Yersinia pseudotuberculosis* l-asparaginase,” *Protein Expr. Purif.*, vol. 82, no. 1, pp. 150–154, 2012.
- [131] M. KOZAI, E. SASAMORI, M. FUJIHARA, T. YAMASHITA, H. TAIRA, and R. HARASAWA, “Growth Inhibition of Human Melanoma Cells by a Recombinant Arginine Deiminase Expressed in *Escherichia coli*,” *J. Vet. Med. Sci.*, vol. 71, no. 10, pp. 1343–1347, 2009.
- [132] C. Schulenburg *et al.*, “The interdependence between catalytic activity, conformational stability, and cytotoxicity of onconase,” *Cancer Biol. Ther.*, vol. 6, no. 8, pp. 1233–1239, Aug. 2007.
- [133] H. Kusakabe, K. Kodama, A. Kuninaka, H. Yoshino, and K. Soda, “Effect of L-lysine α -oxidase on growth of mouse leukemic cells,” *Agric. Biol. Chem.*, vol. 44, no. 2, pp. 387–392, 1980.
- [134] M. Singh *et al.*, “Medicinal uses of L-Lysine: Past and future,” *Int. J. Res. Pharm. Sci.*, vol. 2, no. 4, pp. 637–642, 2011.
- [135] M. SMRIGA, T. ANDO, M. AKUTSU, Y. FURUKAWA, K. MIWA, and Y. MORINAGA, “Oral treatment with L-lysine and L-arginine reduces anxiety and basal cortisol levels in healthy humans,” *Biomed. Res.*, vol. 28, no. 2, pp. 85–90, 2007.
- [136] Y. Mine and H. Zhang, “Anti-inflammatory Effects of Poly-L-lysine in Intestinal Mucosal System Mediated by Calcium-Sensing Receptor Activation,” *J. Agric. Food Chem.*, vol. 63, no. 48, pp. 10437–10447, Dec. 2015.
- [137] Y. JIA, R. HU, and B. K.- Traditional, “Study on Anti-inflammatory Effects of the Complex (LaiJinsi) of EHPL and Lysine Hydrochloride,” *Chinese J. Exp.*, vol. 2004.
- [138] C. Westacott, A. Barakat, L. Wood, ... M. P.-O. and, and U. 2000, “Tumor necrosis factor alpha can contribute to focal loss of cartilage in osteoarthritis,”

- Osteoarthr. Cartil.*, vol. 8, no. 3, pp. 213–221, 2000.
- [139] K. Dudek, “Role of pro-inflammatory cytokines in the etiology of osteoarthritis,” *Med. Weter.*, vol. 60, no. 10, pp. 1039–1041, 2004.
- [140] C. I. Westacott *et al.*, “Tumor necrosis factor alpha can contribute to focal loss of cartilage in osteoarthritis,” *Osteoarthr. Cartil.*, vol. 8, no. 3, pp. 213–221, 2000.
- [141] T. John *et al.*, “Interleukin-10 modulates pro-apoptotic effects of TNF- α in human articular chondrocytes in vitro,” *Cytokine*, vol. 40, no. 3, pp. 226–234, 2007.
- [142] R. D. Müller *et al.*, “IL-10 overexpression differentially affects cartilage matrix gene expression in response to TNF- α in human articular chondrocytes in vitro,” *Cytokine*, vol. 44, no. 3, pp. 377–385, 2008.
- [143] J. Zwerina *et al.*, “TNF-induced structural joint damage is mediated by IL-1,” *Proc. Natl. Acad. Sci.*, vol. 104, no. 28, pp. 11742–11747, 2007.
- [144] G. Y. Wu and C. H. Wu, “Receptor-mediated in vitro gene transformation by a soluble DNA carrier system,” *J. Biol. Chem.*, vol. 262, no. 10, pp. 4429–4432, 1987.
- [145] C. M. Ward, M. Pechar, D. Oupicky, K. Ulbrich, and L. W. Seymour, “Modification of pLL/DNA complexes with a multivalent hydrophilic polymer permits folate-mediated targeting in vitro and prolonged plasma circulation in vivo,” *J. Gene Med.*, vol. 4, no. 5, pp. 536–547, Sep. 2002.
- [146] M. A. Wolfert *et al.*, “Polyelectrolyte Vectors for Gene Delivery: Influence of Cationic Polymer on Biophysical Properties of Complexes Formed with DNA,” *Bioconjug. Chem.*, vol. 10, no. 6, pp. 993–1004, Nov. 1999.
- [147] M. Ohsaki, T. Okuda, A. Wada, T. Hirayama, T. Niidome, and H. Aoyagi, “In vitro gene transfection using dendritic poly(L-lysine),” *Bioconjug. Chem.*, vol. 13, no. 3, pp. 510–517, May 2002.
- [148] M. Forrest, D. P.-M. Therapy, and U. 2002, “On the kinetics of polyplex endocytic trafficking: implications for gene delivery vector design,” *Elsevier*.
- [149] S. Moghimi, P. Symonds, J. Murray, A. H.-M. Therapy, and U. 2005, “A two-stage poly (ethylenimine)-mediated cytotoxicity: implications for gene transfer/therapy,” *Elsevier*.
- [150] S. Vepa, W. M. Scribner, and V. Natarajan, “Activation of endothelial cell phospholipase D by polycations,” *Am. J. Physiol. Cell. Mol. Physiol.*, vol. 272, no. 4, pp. L608–L613, Apr. 1997.
- [151] C. Stefanelli *et al.*, “Spermine triggers the activation of caspase-3 in a cell-free model of apoptosis,” *FEBS Lett.*, vol. 451, no. 2, pp. 95–98, May 1999.
- [152] C. STEFANELLI *et al.*, “Polyamines directly induce release of cytochrome c from heart mitochondria,” *Biochem. J.*, vol. 347, no. 3, pp. 875–880, 2000.
- [153] S. J. Martin, D. M. Finucane, G. P.

- Amarante-Mendes, G. A. O'Brien, and D. R. Green, "Phosphatidylserine externalization during CD95-induced apoptosis of cells and cytoplasts requires ICE/CED-3 protease activity," *J. Biol. Chem.*, vol. 271, no. 46, pp. 28753–28756, 1996.
- [154] S. J. Martin, "Early redistribution of plasma membrane phosphatidylserine is a general feature of apoptosis regardless of the initiating stimulus: inhibition by overexpression of Bcl-2 and Abl," *J. Exp. Med.*, vol. 182, no. 5, pp. 1545–1556, 1995.
- [155] M. O. Hengartner, "The biochemistry of apoptosis," *Nature*, vol. 407, no. 6805, pp. 770–776, 2000.
- [156] R. A. Gottlieb, "Mitochondria and apoptosis," *NeuroSignals*, vol. 10, no. 3–4, pp. 147–161, 2001.
- [157] W. Shier, D. Dubourdieu, J. D. B. A. (BBA)-L. and, and undefined 1984, "Polycations as prostaglandin synthesis inducers: stimulation of arachidonic acid release and prostaglandin synthesis in cultured fibroblasts by poly (l-lysine) and other," *Elsevier*.
- [158] S. Vepa, W. M. Scribner, and V. Natarajan, "Activation of endothelial cell phospholipase D by polycations," *Am. J. Physiol. Cell. Mol. Physiol.*, vol. 272, no. 4, pp. L608–L613, Apr. 2017.
- [159] J. C. Reed, S. Tanaka, S. Takayama, M. J. Schibler, and W. Fenton, "Investigation of the Subcellular Distribution of the bcl-2 Oncoprotein: Residence in the Nuclear Envelope, Endoplasmic Reticulum, and Outer Mitochondrial Membranes," *Cancer Res.*, vol. 53, no. 19, pp. 4701–4714, 1993.
- [160] W. H. Fujita, R. J. Barr, and J. L. Headley, "Multiple Fibrofolliculomas With Trichodiscomas and Acrochordons," *Arch. Dermatol.*, vol. 117, no. 1, pp. 32–35, 1981.