



**PHARMACOLOGICAL ACTIVITIES FOR THE EXTRACT AND ISOLATED
COMPOUNDS OF *EUPHORBIA CUNEATA* VHL**

**AMANI S AWAAD¹, ABDALQAWI M², SANAA A KENAWI³, MONERAH R
ALOTHMAN⁴, REHAB S⁵, AND ZAIN ME⁶**

1: Pharmacognosy Department, College of Pharmacy, Prince Sattam bin Abdul-Aziz
University, Al-Kharj, Saudi Arabia

2: Pharmacognosy Department, College of Pharmacy, Cairo University, Egypt

3: Pharmacology Department, College of Pharmacy, Cairo University, Egypt

4: Botany and Microbiology Department, College of Science, King Saud University, Riyadh,
KSA

5: Aromatic and Medicinal Plant Dept., Desert Research Center, Elmataryah, Cairo, Egypt

6: Botany and Microbiology Department, Faculty of Science, Al-Azhar University, Cairo,
Egypt

***Corresponding Author: Amani S. Awaad: Tel: +966565941707;**

Email: amaniawaad@hotmail.com; Website: <http://https://faculty.psau.edu.sa/a.awaad>

ABSTRACT

The present work explored the chemical profile and pharmacological activities of *Euphorbia cuneata* Vahl. Eight phenolic compounds were isolated and identified using IR, ¹³C-NMR and ¹H-NMR. as; naringenin¹, aromadendrin², apigenin³, 3'-O-methoxy-luteolin-7-O-rhamnoglucoside ⁴, Apigenin 7- galactoside⁵, kaempferol⁶, quercetin⁷ and gallic acid ⁸. The extract showed variable pharmacological activities on both *in-vitro* (on smooth and skeletal muscles) and *in-vivo* (analgesic, antipyretic and anti-inflammatory activity). No side effects were reported on liver and kidney functions.

Keywords: Antimicrobial activity, *Euphorbia cuneata*, gallic acid, kaempferol, naringenin, phenolic compounds

1. INTRODUCTION

Family of *Euphorbiaceae* consists of almost 2000 species. The genus *Euphorbia* is the largest genus of medicinal plants widely distributed [1]. Forty species are

known to occur in Egypt[2,3]The members of this family contain many phytochemical groups and have grate economic importance and medicinal use. This genus is very important due to its various phytochemical constituents as; phenolic compounds[4-6], terpenoids [7], tannins [8], etc.

It is also well known due to its important medicinal uses. The extract of *E. prostrata* has been found to have significant anti-inflammatory, analgesic, haemostatic, wound healing properties and anti-hemorrhoid agent. The major flavonoids of *E. prostrata* extract are apigenin, luteolin, apigenin-7-glucoside and luteolin-7-glucoside. The major phenolic compounds present are gallic acid, ellagic acid and tannins [9]. Several researchers studied the anti-inflammatory activity of some *Euphorbia* species such as; *E. acaulis*[10], *E. peplus*[11],*E. prostrate* [12], *E. heterophylla*[13] and *E. Royleana*[14] they referred their anti-inflammatory activity to the flavonoids present in these species.

Others reported that there are anti-leukemic and antitumor activities for some species such as; *E. esula*[15],*E. lagasa*[16],*E. kansui*[17], *E. fischeriana*[18] and *E. serpense*[19]. There are many other reported biological activities for *Euphorbia* such as;

acetylcholine like action with muscarinic and nicotinicactivities (*E. dracunculoides*, Lam)[20,21], diuretic effect(*E. palustris* and *E. serpens*)[22,23] *Euphorbia cuneata*Vahl growing in the southern coast of the Red Sea in the area of Gabal Elba in Egypt[2,3]. This plant was in use by native in treatment of Gastric disorder but no scientific reported data approved this activity. So, our work aimed at isolation and identification of the biologically active compounds and some other pharmacological activities of *Euphorbiacuneata*.

2.MATERIAL AND METHODS

2.1.Plant materials: The aerial parts of *Euphorbiacuneata*, Vahl.(*Euphorbiaceae*) were collected during flowering stage in 2006, from Gabal Elba in Halayeb and Shalatin. The sample was kindly identified by Dr. Dr. M. Gebali, former researcher of Botany, and by comparison with plant description Flora of Egypt[2,3]. A voucher specimen (19356 SH) of the titled plant is kept in the herbarium of the Desert Research Center. Plant materials were air-dried in shade, reduced to fine powder, packed in tightly closed containers and stored for phytochemical and biological studies.

2.2. Extraction &Isolation of the active compounds

The air dried powder of *E.cuneata*, was extracted by percolation in 90% ethanol at room temperature for two days and then filtered. The residue was re-percolated again; for four times. The combined ethanol extracts were concentrated under reduced pressure at a temperature not exceeding 35°C till a dry extract was yielded. Water was added and the resultant mixture successively extracted with diethyl ether, chloroform, ethyl acetate, and *n*-butanol respectively. Each extract was dried over anhydrous sodium sulphate, and concentrated under reduced pressure at a temperature not exceeding 35°C and dry extracts were obtained.

Chromatographic examinations of the successive extracts were carried out using Silica Gel G (stationary phase) and solvent systems; (a), ethyl acetate-methanol-water (30:5:4); (b), ethyl acetate-methanol-acetic acid-water (65:15:10:10); (c) chloroform-methanol (95:10); (d) butanol-acetic acid- water (4:1:5) were used for developing the chromatoplates. Visualization of chromatograms was achieved under UV before and after exposure to ammonia vapor or by spraying with aluminum chloride [24]. TLC examination of the different extractives adopting solvent systems (a and c) and visualizing reagent revealed the presence of same 2 spots in ether and chloroform

extracts while ethyl acetate and *n*-butanol extracts were found to have 5 similar spots.

Accordingly, the ether and chloroform extracts combined together and applied on the top of column chromatography packed with silica gel and eluted gradually with chloroform - methanol, 90 fractions (100 ml each) were collected and reduced to 3 main sub-fractions in different yields 2.0, 3.7 and 4.6 respectively (according to the number, color and R_f values of the spots), these sub-fractions were reapplied on preparative thin layer chromatography (system c) for final purifications from which compound **1-3** were isolated.

The ethyl acetate and butanol extracts were combined together and subjected to column chromatography packed with silica gel and eluted gradually with ethyl acetate -methanol, from this column 150 fractions (100 ml each) were collected and reduced into 6 mean fractions (according to number, colure and R_f of the spots), each fraction was concentrated under reduced pressure to yield 6.11, 4.55, 3.22, 4.87, 3.11 and 5.98 respectively. The residue of each fraction was separately reapplied on preparative paper chromatography (using systems d). Bands corresponding to each flavonoid were separately extracted with methanol, concentrated and re-purified on a column of

Sephadex LH-20 eluted with methanol – water where compounds 4-8 were isolated.

2.4. Biological Activities

2.4.1. Plant extract

Dried aerial parts of *Euphorbiacuneata*, Vahl were extracted as mentioned before apparatus with ethanol 95%. The ethanol extract was completely dried under vacuum, weighed and the residue was used in testing. The dried plant extract was freshly suspended in distilled water just before administration.

2.4.2. In-vitro investigation

2.4.2.1. Isolated smooth and skeletal muscles:

The effect of the alcoholic extract on rabbit intestine motility and frog rectus abdominals were studied using the glass organ bath as described by the staff of Department of Pharmacology, University of Edinburgh [25, 26].

2.4.3. In- vivo investigation

2.4.3.1. Determination of median lethal dose (LD50)

The oral LD50 of the total alcohol extract of *E. cuneata* was determined as described by Soliman et al., [27]. Groups of six mice, received one of 500, 1000, 2000, or 4000 mg/kg doses of the tested extract. Control animals were received the vehicle and kept under the same conditions. Signs of acute toxicity and number of deaths per dose within 24h were recorded.

2.4.3.2. Antinociceptive activity

Anti-nociceptive were determined using acetic acid induced writhing test [28]. Six groups of mice each of six were used. Three groups received the total alcohol extract of the investigated plant at doses 100, 200 & 400 mg/g, control group received the vehicle, standard group received diclofenac sodium (30 mg/kg) and the last group received the isolated narigenin (100 mg/kg).

2.4.3.3. Anti-inflammatory activity

Anti-inflammatory activity was assessed using carrageenan induced paw edema method [29]. Thirty six rats were distributed equally into six groups. Groups 1, 2 & 3 received the total alcohol extract of the investigated plant (100, 200 & 400 mg/g), group 4 (control) received the vehicle, group 5 (standard) received diclofenac sodium (30 mg/kg) and the last group received the isolated narigenin (100 mg/kg).

2.4.3.4. Antipyretic activity:

Antipyretic were assayed using yeast-induced hyperpyrexia method described by Loux et al., [30]. Six groups each of six rats were used. Three groups received the total alcohol extract of the investigated plant at doses 100, 200 & 400 mg/g, control group received the vehicle, standard group received paracetamol (100 mg/kg)

and the last group received the isolated naringenin (100 mg/kg).

2.4.3.5. Measurement of liver and kidney functions

Rats were randomly divided into two groups each of 10. Rats of the 1st group received the vehicle in a dose of 5 mL/kg and left as normal control. Rats of the 2nd, group were administered the alcohol extract of *E. cuneata* (400 mg/kg). All medications were administered orally daily for 35 consecutive days. After the treatment period, serums were collected and many markers were measured. Serum activity of alanine transferase (ALT) and aspartate aminotransferase (AST) following the method of Christoph et al., [31]. Serum concentrations of urea and creatinine were determined [32,33].

Statistical analysis: All parametric values are given as Mean \pm S.E.M and were analyzed by One-way ANOVA followed by Student Newman-Keuls test for significance at $p < 0.05$.

3. RESULT AND DISCUSSION

3.1. Chemical investigation

3.1.1.2. Isolated active compounds: Eight phenolic compounds were isolated and identified as;

Naringenin¹; Aromadendrin²; Apigenin³; 3'-O-methoxy-luteolin-7-O-rhamnoglucoside **4**, Apigenin-7-galactoside⁵; Kaempferol⁶; Quercetin⁷ and

Gallic acid **8** (Figure 1). Identification was carried out by comparing their TLC chromatograms, UV spectra in methanol and with different shift reagents, EI-MS, ¹H NMR and ¹³C NMR spectra with authentic samples [34].

3.2. In-Vitro Activities

3.2.1. Effect on isolated smooth and skeletal muscles: The alcohol extract in different concentrations (5, 10, 25, 50, 100, 200 and 300 mg/ml) showed initial stimulation to the rabbit intestinal followed by slight inhibition and then causes blocking of the ganglia. The effect of the extract seems reversible by washing on the motility and study of the possible site of action.

3.2.2. Effect on rectus abdominals:

No effect was observed on the muscle at doses up to 300 mg/ml.

3.3. In-Vivo Activities

3.3.1. Determination of median Lethal Dose (LD₅₀): The total alcohol extract *E. Cuneata* did not produce any behavioral changes and mortality in mice in doses up to 5000 mg/kg

3.3.2. Analgesic activity: the alcohol extract has a significant analgesic activity at the dose starting from 1g/ kg body weight when using both acid-induced pain and Eddy's hot plate method (Tables 1&2). The isolated compound naringenin had no significant analgesic activity.

3.3.3. Anti-inflammatory activity

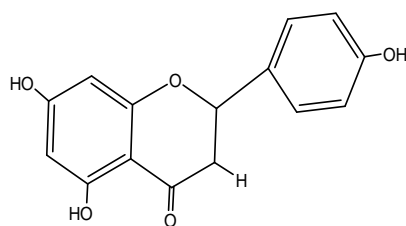
The obtained results (Table 3) revealed that there is no significant effect with the extract (1g/kg).

3.3.4. The antipyretic activity

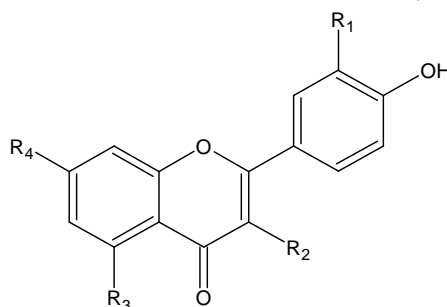
There is no antipyretic activity for the alcohol extract or the isolated compound naringenin (Table 4).

3.3.5. Measurement of liver and kidney functions

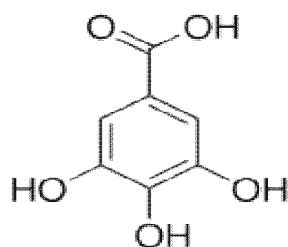
Daily administration of the alcohol extract (2 g/kg) for 15 days did not produce any observable side effects related to liver or kidney functions (Table 5).



1. R = OH (Naringenin).
2. R = OH (Aromadendrin).



	R1	R2	R3	R4	
3.	H	H	OH	OH	(Apigenin)
4.	OCH ₃	H	OH	rha.gluc.	(3'-O-methoxy-luteolin-7-O-rhamnoglucoside)
5.	H	H	OH	gal.	(Apigenin-7- galactoside)
6.	H	OH	OH		(Kaempferol)
7.	OH		OH	OH	(Quercetin)



8. Gallic acid

Figure 1: The compounds isolated from *Euphorbia cuneata* Vahl

Table 1: The analgesic effect of different doses of the alcoholic extract and naringenin on acetic acid-induced pain

Groups	No. of writhing/20 minutes (mean \pm S.E.)	Percentage of Inhibition
Control	20.714 \pm 1.222	
Diclofenac	15.600 \pm 1.284	64.12
0.5 g/ kg	7.600 \pm 0.5640	24.69
1 g/ kg	2.100 \pm 0.640 *	63.32
2 g/ kg	3.100 \pm 0.767*	89.88
4 g/ kg	7.430 \pm 0.400*	85.05
Naringenin (100 mg/ml)	19.514 \pm 0.988	5.79

* Significant at P<0.05

Table 2: The analgesic effect of different doses of the alcoholic extract and naringenin on by writhing method in mice

Treatments	Dose (mg/kg)	No. of writhing / 20 minutes (mean \pm S.E.)	Percentage of Inhibition
Control	400	35.83 \pm 0.87	01.83
Diclofenac	200	20.66 \pm 1.08*	43.39
0.5 g/ kg	400	35.33 \pm 1.02	03.20
1 g/ kg	400	35.50 \pm 1.05	02.73
2 g/ kg	400	25.00 \pm 0.73	31.50
4 g/ kg	400	36.00 \pm 0.73*	01.36
Naringenin (100 mg/ml)	400	36.00 \pm 0.81	01.36

* Significant at P<0.05

Table 3: The anti-inflammatory effect of different doses of the alcoholic extract and Naringenin on carrageenan-induced oedema

Treatments	Difference in paw thickness (mm) after		
	1h	2h	3h
Control (saline)	1.375 \pm 0.249	2.067 \pm 0.133	2.260 \pm 0.161
Extract (1 g/ kg)	1.362 \pm 0.191	1.657 \pm 0.265	2.557 \pm 0.431
Extract (2g/ kg)	1.155 \pm 0.171	0.555 \pm 0.166*	0.703 \pm 0.172*
Extract (4 g/ kg)	0.948 \pm 0.135	1.477 \pm 0.199	3.057 \pm 0.308
Diclofenac(50mg/kg)	1.300 \pm 0.120	0.610 \pm 0.160*	0.200 \pm 0.210*
Naringenin (100 mg/kg)	1.358 \pm 0.262	2.136 \pm 0.153	2.311 \pm 0.182

* Significant at P<0.05

Table 4: The antipyretic effect of different doses of the alcoholic extract on yeast -induced hyperthermia

Treatments	Difference in rectal temp ($^{\circ}$ C) after			
	0 time	1 h	2 h	3 h
Control (saline)	2.833 \pm 0.325	3.317 \pm 0.282	2.900 \pm 0.459	3.233 \pm 0.406
Extract (1 g/ kg)	2.800 \pm 0.403	2.833 \pm 0.358	3.083 \pm 0.426	2.950 \pm 0.409
Extract (2g/ kg)	1.917 \pm 0.227	2.650 \pm 0.167	2.650 \pm 0.281	2.500 \pm 0.250
Extract (4 g/ kg)	2.083 \pm 0.294	2.317 \pm 0.341	2.883 \pm 0.227	3.017 \pm 0.287
Paracetamol (100mg/kg)	3.000 \pm 0.703	1.500 \pm 0.500*	1.100 \pm 0.521*	0.700 \pm 0.410*
Naringenin (100 mg /kg)	2.743 \pm 0.331	3.421 \pm 0.253	3.101 \pm 0.988	3.324 \pm 0.288

* Significant at P< 0.05

Table 5: Effect of fifteen daily administrations of alcohol extract on kidney and liver functions

Treatment	Liver functions		Kidney functions	
	ALT	AST	Urea	Creatinine
Control	10.800 \pm 0.583	13.000 \pm 0.707	50.812 \pm 3.568	0.970 \pm 0.016
Extract	11.600 \pm 3.999	13.800 \pm 1.019	66.556 \pm 5.208	1.078 \pm 0.045

DISCUSSION

Upon investigation of the active compounds in *Euphorbia cuneata* 8 compounds were isolated, only naringenin obtained in a higher amount which allowed us to investigate some of its biological activities with the total extract. It seems that neither the alcoholic extract nor the isolated compound naringenin has antipyretic activity. On the other hand the use of the standard antipyretic paracetamol produced significant antipyretic effect [35, 36].

The biological activities of the plant extract showed very promising activities *in-vitro* and *in-vivo*. The effect of the extract on isolated smooth and skeletal muscles showed stimulation of the contraction followed by slight inhibition was recorded following the addition of 200 mg/ ml, same result was achieved in 10 separate experiments. In order to identify the site of action, dilute nicotine (1 µg / ml) was applied for 30 seconds then washed. The alcoholic extract was applied (1 mg / 1 ml organ bath) for 5 minutes followed without wash by the same dose of dilute nicotine (1 µg / ml). This time dilute nicotine did not produce any stimulation this means that the alcoholic extract possesses a ganglion blocking effect. Washing then addition of the same dose of dilute nicotine to the contracting muscle produced the same stimulation. It

could be concluded that the alcoholic extract contain substances that initially stimulates and then causes blocking of the ganglia. The effect of the extract seems reversible by washing. On the other hand it seems that the alcohol extract of *E. cuneata* is devoid of substances that can stimulate skeletal muscle because it didn't show any effect on rectus abdominals.

The variation of antimicrobial activities which were observed from alcohol extract when it tested on 10 different microorganisms revealed the possible use of this plant for treating such organisms as total better than successive fractions.

The *in-vivo* activities of the plant extract proved that this plant is very safe for human use because No deaths were recorded in mice treated with the alcohol extracts in doses up to 5000 mg/kg. This means that, the investigated extract was safe for use, since substances possessing safety test higher than 0.5 g/kg are considered non-toxic [37]. It seems that the alcoholic extract has a significant analgesic activity at the dose starting from 1g/ kg body weight. The isolated compound naringenin had no significant analgesic activity. No significant effect was produced with the extract (1g / kg). The alcoholic extract started to have an anti-inflammatory activity at a dose of 2 g/ kg after 2 and 3 hours. However increasing the dose to 4 g /

kg did not produce any significant anti-inflammatory effect. Furthermore the isolated compound naringenin had no significant anti-inflammatory activity.

Side effects of total alcohol extract showed that this plant has no adverse effect on both liver and kidney function because the results were very close to that reported to the control group of animal [38, 39].

ACKNOWLEDGMENT

This research project was supported by a grant from the “Research Center of the Female Scientific and Medical Colleges”, Deanship of Scientific Research, King Saud University.

REFERENCES

- [1] Singla, A.K. and K. Pathak. 1989. Anti-inflammatory studies on *Euphorbia prostrata*. *Journal of Ethnopharmacology*. 27:1-2,55-61.
- [2] Bolus, L. 2002. *Medicinal plant of North Africa*. Reference PubnsInc, Cairo, Egypt. 268 pp.
- [3] Tackholm, V. 1974. *Student's flora of Egypt*. 2nd edition. Cairo University publisher. pp 532.
- [4] Galarza, S. Del V.; Gabreya, J. L. and Juliani, H. R. 1984. Constituents of *Euphorbiaceae* from Argentina, Part I. *An. Assoc. Quim. Argent.*, 71 (4), 505 – 8.
- [5] Khamzinova, V. B.; Erzhanova, M. S.; Kukenov, M. K. and Nurkianova, K. M. 1989. Polyphenols from *Euphorbia songarica* Boiss and *Euphorbia alata* Boiss. F. E. C. S. Int. conf. Chem. *Biotecnol. Biol. Act. Nat. Prod.* (Proc.), 3rd 1985 (Pub. 1987); 5, 487 – 91. From C. A. 110: 4071g.
- [6] Liu, L-G; Meng, J- C.; Wu, S-X; Li, X-y; Zhoo, X-C and Tan, R-X. 2002. New macrocyclic diterpenoids from *Euphorbia esula*. *Planta Med.*, 68 (3), 244-8.
- [7] Ahn, B. T.; Zhang, B. K.; Lee, S. C.; Kim, J. G.; Jai, S. and Lee, K.S. 1996. Flavonoid constituents of *Euphorbia pekinensis*. *Yakihak Hoechi*, 40 (2), 170 – 6.
- [8] Yashida, T.; Namba, O.; Kurokawa, K.; Amakuka, Y.; Liu, Y. and Okuda, T. 1994. Tannins and related polyphenol of *Euphorbiaceae* plants XII, Euphorbin G and H, New dimeric hydrolysable tannin from *E. chamaesyce*. *Chem. Pharm. Bull.*, 42 (10), 2005- 2010.
- [9] Bakshi, D.G., D.G. Langade and V. Desai. 2008. Prospective open label study of *Euphorbia prostrata* extract 100 mg in the treatment of bleeding hemorrhoids. *Bombay Hospital Journal*, 50 (4): 577-583.

- [10] Parmar, N.S.; Mossa, J.S.; Al-Yahya, M.A.; Al-Said, M.S. and Tarig, M. 1989. Anti-inflammatory, analgesic and antipyretic activity of *E. acaulis*, *RoxbInt J. Crude Drug Res.*, 27, 9-13.
- [11] Singh, G. B.; Kaur, S.; Satti, N. K.; Atal, C. K. and Maheshwari, J. K. 1984. Anti-inflammatory activity of the aerial parts of *E. peplus*. *J. Ethnopharmacol.*, 10, 233- 55.
- [12] Singla, A.K. and Pathak, K.1990. Topical anti-inflammatory effects on *Euphorbia pcostrata* on carrageenan- induced foot pad aedema in mice. *J. Ethnopharmacol*, 29(3), 291-4.
- [13] Vamsidhar, I.; Mohammed A. H.; Nataray B.; Roe C. and Madhusudana, R.M. .2000. Antinociceptive activities of *Euphorbia heterophylla* roots. *Fitoterapia*, 71 (5), 562-3.
- [14] Bani, S; Kaul, A; Jaggi, B-S.; Suri, K-A; Suri, O-p. and Shormo, O-P. 2000. Anti-inflammatory activity of the hydrosoluble fraction of *Euphorbia royleana* latex. *Fitoterapia*, 71 (8), 655-62.
- [15] Whelan, L-C and Ryans, M-F. .2003. Ethanolic extracts of *Euphorbia* and other ethobotanical species as inhibitors of human tumour growth. *Phytomedicine J.*; 10 (1), 53-8.
- [16] Ferigri, N.R.; Mclaughlin, J.L.; Powell, R.G. and Simth, C.R. 1984. Use of potato disc and brine shrimp bioassay to detect activity and isolate piceatannol as the antileukemic principle from the seeds of *Euphorbia lagascese*. *J. Nat. Prod.*, 47, 347-52.
- [17] Kupchan, S. M.; Uchida, I. and Branfman, A. R. 1976. Antileukemic principles isolated from *Euphorbia* plants Daily, R. G. , Jr. and Fei, B. Yu.,. *Science*, 191, 571 – 2
- [18] Cui, X.; Yi, L.; Wang, S.; Wang, W.; Kong, G. and Liu, P. 1998. Inhibition of L615 leukemia cell proliferation and recovery of GSH- PX and sod activation in L615 mice *Euphorbia fiseheriana* stead. *Shan dong yike Daxue Xuebao*, 36 (4), 289-92.
- [19] Cheruse, J.J.; Baldini, O.A.N. and Blanch, L.E.B. 1983. Investigation of the biological activity of *E. serpense*. *Acta Farmaceutica Bonaerense*, 2 (1), 12-22.
- [20] Bondarenko, O. M.; Chogovets, R. K.; Litvinenko, V. I. ;

- Obolentseva, G. V. ; Sila, V. I. and Kigel, T. B.; (1972). *Euphorbia palustris* and *Euphorbia stepposaflavonoides* and their pharmacological properties. *Farm, Zh.*, 26 (6), 46 – 8 (1971). *From C. A. 76 : 121697k.*
- [21] Prasad, D. N.; Gode, K. D. ; Singh, R. S. and Das, P. K.; (1967). Preliminary phytochemical and pharmacological studies on *Euphorbia dracunculoides*, *Lam.* *Indian J. Med. Res.* 55 (1), 73 – 9.
- [22] AwaadAmani S., Nabilah A. Al-Jaber, Gamal A. Soliman, Mounerah R. Al-Outhman, Mohamed E. Zain, John E. Moses and Reham M. El-Meligy. 2012. New Biological Activities of *Casimiroaedulis* Leaf Extract and Isolated Compounds. *Phytother. Res.* 26: 452–457.
- [23] AwaadAmani S.; Nawal H. Mohamed; N.H. El-Sayed; G. A. Soliman and T.J. Mabry (2004). "Flavonoids, carbohydrates and some pharmacological screening of *Convolvulus arvensis*. *Journal of Desert Research Centre*, 54(1), 15-33.
- [24] Stahl, E.1969. Thin-layer chromatography, 2nd ed. GeorgeAllen andUnwinlid London, springer-verlag Berlin.pp.960.
- [25] Staff of the department of Pharmacology, University of Edinburgh. 1970. *Pharmacological Experiments on isolated preparations.* 2nd ed., 54, Livingstone, E. and S. ltd., Edinburgh and London.
- [26] NCCLS. 2000. *Methods for Dilution Antimicrobial Susceptibility Tests for Bacteria That Grow Aerobically; Approved Standard Fifth Edition.* NCCLS Document M7-A5, NCCLS: Wayne, PA, USA.
- [27] Soliman G. A., Donia A.M., Awaad A.S., Alqasoumi S. I., and Yusufoglu H. 2012. Effect of *Emexspinosa*, *Leptadeniapyrotechnica*, *Haloxylonsalicornicum* and *Ochradenusbaccatus* extracts on the reproductive organs of adult male rats. *Pharmaceutical Biology*, 50(1): 105–112.
- [28] AlqasoumiSaleh Ibrahim, GamalAbd El Hakim Soliman, AmaniShafeekAwaad, Abd El Raheim Mohammed Donia. 2012. Anti-inflammatory activity, safety and protective effects of *Leptadeniapyrotechnica*,

- Haloxylonsalicornicum* and *Ochradenusbaccatus* in ulcerative colitis. *Phytopharmacology*, 2(1) 58-71.
- [29] Winter, C.A.; Risely, E.A., and Nuss, G.V.1962. Carragennin-induced oedema in hind paw of the rat as an assay for anti-inflammatory drugs. Proceeding of the society of experimental *Biology and Medicine*, 111, 544-7.
- [30] Loux J.J., DePalma P.D. and Yankell S.L. 1972. Antipyretic testing of aspirin in rats. *Toxicol. Appl. Pharmacol.*, 22:672.
- [31] Christoph K. Meinecke, Justus Schottelius, Linda Oskam, and Bernhard Fleischer (1999). Congenital Transmission of Visceral Leishmaniasis (Kala Azar). From an Asymptomatic Mother to Her Child. *Pediatrics*, Vol. 104 (5): 1-5.
- [32] Wills MR, Savory J. 1981. Biochemistry of renal failure. *Annals of Clinical and Laboratory Science*. 11(4), 292-299.
- [33] Zain M. E., Amani S. Awaad, Razak, A.A., Maitland D.J., Khamis N.E. and Sakhawy M.A. 2009. Secondary Metabolites of *Aureobasidium Pullulans* Isolated from Egyptian Soil and Their Biological Activity. *Journal of Applied Sciences Research*, 5(10): 1582-1591.
- [34] Mabry TJ, Markham KR, Thomas MB. 1970. The Systematic Identification of Flavonoids. Springer-Verlag, Berlin. P, 345.
- [35] Kalyana Sundram, Tilakavati Karupiah and KC Hayes. 2007. Stearic acid-rich interesterified fat and trans-rich fat raise the LDL/HDL ratio and plasma glucose relative to palm olein in humans. *Nutrition & Metabolism*, 4:3. pp123-234
- [36] BEEF FACTS • Human Nutrition Research Stearic Acid—A Unique Saturated Fat. Online at; <http://www.beefnutrition.org/CMDocs/BeefNutrition/StearicAcid.pdf>
- [37] Awaad, A.S., El-Meligy, R.M., Al-Jaber, N.A., Al-Muteeri, H.S., Zain, M.E., Alqasoumi, S.I., Alafeefy, A.M., Donia, A.M., 2013. Anti-ulcerative colitis activity of the total alcohol extracts and isolated compounds from *Euphorbia granuleta* Forssk. *Phytother. Res.* 11, 188–200.
- [38] Kroll, MH, Roach NA, Poe B, Elin RJ. 1987. Mechanism of interference with Jaffé reaction for

creatinine. *Clinical Chemistry*. 33, 1129-1132.

- [39] Buck D. R., Arthur W. Mahoney, Deloy G. Hendricks. 1976. Effect of magnesium deficiency on nonspecific excitability level (NEL) and audiogenic seizure susceptibility. *Pharmacol. Biochem. Behav.*, 5(5): 529.