



**International Journal of Biology, Pharmacy
and Allied Sciences (IJBPAS)**

'A Bridge Between Laboratory and Reader'

www.ijbpas.com

RECENT ADVANCES AND BENEFICIAL ROLES OF *BACILLUS MEGATERIUM* IN AGRICULTURE AND OTHEIRS FIELDS

DJADOUNI FATIMA^{*1} AND MADANI ZOHRA²

^{*1}Departement of Biology, Faculty of Natural Sciences and Life, Mascara University,
Mascara 29000, Algeria, Telephone: 00213781879665, Email: Houarisalwa@yahoo.fr

²Departement of Biology, Faculty of Natural Sciences and Life, Mascara University, Mascara,
29000, Algeria, Telephone: 00213559447311, Email: zohramadani14@gmail.com

Received 10th Aug. 2016; Revised 20th Sept. 2016; Accepted 29th Oct. 2016; Available online 1st Dec. 2016

ABSTRACT

Bacteria spread in nature almost every where, they exist in the soil, fresh water, saltwater, deep-sea, hot springs water, and in polar snow as airborne into the upper atmosphere. It is more prevalent in places where food is available, humidity and temperature suitable for their growth and reproduction, and since this is the conditions the same in which man lives, so we find large numbers of bacteria in air, food, humans and animals skin, in the channel tract.....etc. Due to the presence of bacteria in large numbers in most natural circles, they occurs beneficial changes in the earth. Expansion bacterial activity and multi-ranging impact on soil fertilitization, biomethylation, oxidation and reduction, eutrophication, bioremediation, mineralization, and interactions plants with soil microorganisms, so we'll show in this mini review some of the recent advances and beneficial roles of *Bacillus megaterium* in soil and agriculture and others fields.

Keywords: *Bacillus megaterium*, applications, agriculture, environment, industry, medicine, pharmacy.

INTRODUCTION

Evolution of applied bacteriology, *megaterium*, these bacteria was found in biotechnology, genetic engenering, many places including soil, water, wastes, biochemistry and molecular biology plants, animals, insects, and in very harsh reinforced the wides pread use of *B.* conditions. The continuous research on

improving soil fertility with its multiple kinds, different climates, water shortages in many regions around the world, and try to reduce the soilborne plants pathogens and diseases, and the use of chemical fertilizers make scientists look for alternative in the use of microorganisms in agriculture specially *Bacillus* sp.

Application of *B. megaterium* in agriculture and horticulture

Many of the world is affected by the scarcity of rain and not good for agriculture and water availability is affected as well as the bad effects produced as a result of the solubility of the salts deposited in the soil were added whenever water during irrigation. Most of the land is characterized by (high pH pH 8 and more), as well as its proximity to the very high percentage of calcium carbonate.

The high alkalinity scale in this land and the presence of a high percentage of calcium carbonate has a major negative impact on the availability and the availability of major and trace elements important and necessary to feed the plants, which also affects the fertility of the soil and thus on agricultural land efficiently and from then on the agricultural production [1-2-3]. Phosphorus component of the major elements important for the growth and plant nutrition in all its stages, and is considered the second component after the nitrogen in

terms of importance. Phosphorus is added to the soil in the form of inorganic phosphate fertilizers such as single super phosphate and triple super phosphate and rock phosphorus [4-5].

In most of the agricultural land lives microorganisms beneficial bacteria and fungi and is known organisms analyst minute and dissolving phosphorus where these organisms play a very important role in the analysis and dissolving phosphorus installer and non slushy and non-available and non-absorbable by plants the image dissolved and absorbable, leading to increased availability of phosphorus dissolved element in the soil [6].

And therefore the presence of these microorganisms dissolving phosphorus increases the soil fertility and agricultural production of crops and by increasing the solubility of phosphorus. The most important types of bacteria dissolving phosphorus is that follow the genus *B. megaterium*, *B. subtilis*, *Serratia* spp., *Proteus* spp., *Arthrobacter* spp., enterobacters, *Pseudomonas* spp., and *Streptomyces* spp. and fungi such as *Aspergillus* spp., *Penicillium* spp., *Rhizopus* spp., *Fusarium* spp., and *Cunninghamella* spp. [7-8-9].

These bacteria was involved in the cycle of phosphorus and microbial mineralization of organic phosphorus (biofertilizers),

necessary for the plants can use this same organic phosphorus and nitrogen fixation (*B. megaterium* var. *phosphaticum*) [10-11]. Due to its ability to produce a range of

enzymes, solubilization pounded nutrients and degrade organic wastes and along with the N₂-fixing ability of some strains [12-13].

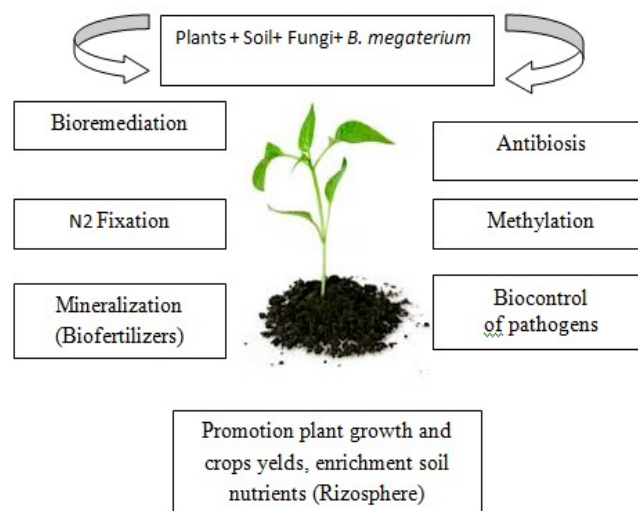


Fig. Important roles of *B. megaterium* on soil, plants and rhizosphere.

Presence of *B. megaterium* in agricultural fields reported to enhance plant growth and crop yield including sugar beet, barley, alfalfa, clover, wheatgrass, perennial ryegrass, soybean and cicer. Production of amino acids, vitamins, indole acetic acid, gibberellic acids, antibiotics induced systemic resistance to plant pathogens, production of siderophore and inhibition of plant ethylene synthesis are reported to be the possible reasons for crop yield increase in addition to nutrient solubilisation [14-15-16-17-18].

Also, it has been reconnu as an endophyte and is a potential agent for the biocontrol of plant diseases and pathogens [19-20] such as *Aspergillus flavus* in peanut [21], *Mycosphaerella graminicola* caused foliar disease of wheat (Septoria Tritici Blotch)

[22], its antagonism against select phytopathogens such as *Rhizoctonia solani*, *Macrophomina phaseolina*, *Sclerotium rolfsii* and *Fusarium oxysporum* [23], biocontrol agent of red-pepper Phytophthora blight Disease by *B. megaterium* KL39 strain [24].

B. megaterium have been advocated as effective and economical bioinoculant to use in the integrated nutrient and pest control system, suppression of bacterial and fungal, nematode pathogens by production of antibiotics, ammoniaetc [25-26].

Environnement

Increase use of fossil fuels (industrial wastes), agricultural Activities (chemical pesticides and insecticides), trading activities, and residences are the causes of environmental pollution and ecosystems

destroy. Microorganisms play a dominant role in transforming pollutants that reach the environment ; *B. megaterium* was used alone or in combination with other *Bacillus* species in eliminating odors and heavy metals from polluted areas, converting wastewater or food waste from livestock into useful resources, improving water quality, balance microbial population and reducing pathogenic bacteria in aquaculture ponds, degrading and absorbing dyes from wastewater, reducing water pollution, decolorizing of the textile dye, and bioremediation of lubricant oil pollution in water and soil ecosystems [27-28-29-30-31].

Medicine and Pharmacy

B. megaterium was considered non-pathogenic specie for human [32-33] and its produces a large variation of proteins [34-35] including penicillin amidase [36] used for the production of penicillins that are used in the treatment of bacterial infections, mainly against Gram positive and some Gram negative bacteria, glucose dehydrogenase used in glucose blood tests in the case of diabetes mellitus or hypoglycemia, enzymes for modifying corticosteroids (steroid hormone), and amino acid dehydrogenases that catalyses the oxidation of D-amino acids into their corresponding oxoacids [37].

Antibiotics, amino acids and vitamins are also produce by these bacteria such as penicillin G Acylase witch is one of the most relevant and widely used biocatalysts for the industrial production of β -lactam semisynthetic antibiotics [38-39-40], vitamin B12 which is essential cofactors for the human enzymes methylmalonyl CoA mutase and methionine synthase and also for normal functioning of the brain, nervous system, blood formation, and the treatment of megaloblastic anemia, pernicious anemia, diabetic neuropathy [41], pyruvate [42], lysine [43] that used in animal and human nutrition to balance diets, exotoxin [44], teichuronic acid [45], and the strain *B. megaterium* QM B1551 was used to produce the antigen for HIV Diagnostic Kits [46].

Industrial applications

The first exploitation of *B. megaterium* for use in industrial processes back to pre-1960. The reasons for intensive use in the laboratory to (1) simples cultural conditions, (2) diversity of carbon and nitrogen sources, (3) no endotoxins found in the cell wall, (4) no alkaline protease secretion, (5) Growth on various media, (6) no pathogenicity [37].

Many primary and secondary metabolites of *B. megaterium* contributed to improving the quality of foods and feeds such as α - and β -amylases used in the baking industry [47-

48], neutral proteases which are used by the leather industry [49], and probiotics used in shrimp culture industry [28]. Production of dextransucrase [50], sucrose [51], carboxy methyl cellulase from *B. megaterium* KU365409 [52].

It also produce poly- γ -glutamic acid, that has potential applications as thickener, cryoprotectant, humectant, drug carrier, biological adhesive, heavy metal absorbent, etc., with biodegradability in the fields of food, cosmetics, medicine, and water treatments [53-54].

High production of biopolymers such as poly (3-hydroxybutyrate) from a wild *B. megaterium* Bolivian strain [55-56], it was 100% biodegradable and are also biocompatible with a wide range of applications in medicine, pharmacy, veterinary and food packaging.

B. megaterium produces other compounds used in the food processing industry [57] such as L-asparaginase from *B. megaterium* H-1 and its application in French fries, and α -amylase and glucose isomerase producing *B. megaterium* BPTK5 isolated from starch processing plant (cassava waste) [58], and tannase enzymes that used as a hydrolysing agent in cleaning up the highly polluting tannin from the effluent of leather industry [59].

Antimicrobial peptides produced by *B. megaterium* were named bacteriocins or

bacteriocin-like substances including megacins [60-61-62]; it is a large proteins (430 kDa) class III, had a bactericidal effect, rapidly digested by proteases in the human digestive tract, and characterized by nontoxicity in animals and humans and their effectiveness against spoilage and pathogenic bacteria [63-64-65-66].

Microbial biotechnology and *B. megaterium*

B. megaterium was extensively studied genetically and is amenable to genetic manipulation [67]. Progress in genetics and the availability of molecular tools such as new transposons, vectors and efficient transformation, an understanding of some of the organization and regulation of many genes and bacteriophages studies make it under investigations ; because (1) desirable cloning host and produces a large variation of enzymes, (2) good cloning host and able to house numerous plasmid vectors while remaining stable due to its unique external proteases, (3) the organism does not have alkaline proteases which allows for recombinant protein synthesis [68-69-70-71-72-73-74].

CONCLUSION

The search of a healthy eating, natural medicine, improve soil quality, reducing ecosystems pollution, and improve foods and feeds quality and bioconservation ; make *B. megaterium* under investigation to

discovery of new features for applied it in others fields.

REFERENCES

- [1] Richardson AE. Making microorganisms mobilize soil phosphorus. In: E. Velázquez and C. Rodríguez-Barrueco (eds.), First international meeting on microbial phosphate solubilization. 2007;85-90.
- [2] Khan KS, Joergensen RG. Changes in microbial biomass and P fractions in biogenic household waste compost amended with inorganic P fertilizers. *Bioresour Technol* 2009a;100:303-309.
- [3] Bhardwaj D, Ansari MW, Sahoo RK, Tuteja N. Biofertilizers function as key player in sustainable agriculture by improving soil fertility, plant tolerance and crop productivity. *Microbial Cell Factories* 2014;13:66.
- [4] Khan AA, Ghulam J, Saleem AM, Naqvi SSM, Mohammad R. Phosphorus solubilizing bacteria: Occurrence, mechanisms and their role in crop production. Published in *J. agric. biol. sci* 2009b;1(1):48-58. A biannual publication of PMAS AridAgriculture University Rawalpindi, Pakistan.
- [5] Plassard C, Robin A, Le Cadre E, Marsden C, Trap J, Herrmann L, Waithaisong K, Lesueur D, Blanchart E, Chapuis-Lardy L, Hinsinger P. Améliorer la biodisponibilité du phosphore: Comment valoriser les compétences des plantes et les mécanismes biologiques du sol? *Innovations Agronomiques* 2015;43:115-138.
- [6] Yazdani M, Bahmanyar MA, Pirdashti H, Esmaili MA. Effect of phosphate solubilization microorganisms (PSM) and plant growth promoting rhizobacteria (PGPR) on yield and yield components of Corn (*Zea mays* L.). *Proc World Acad Science Eng Technol* 2009;37:90-92.
- [7] Idris EES, Bochow H, Ross H, Boriss F. Use of *Bacillus subtilis* as biocontrol agent. Phytohormone action of culture filtrate prepared from plant growth promoting *Bacillus amyloliquefaciens* FZB24, FZB42, FZB45 and *Bacillus subtilis* FZB37. *J Plant Dis Prot* 2004;111:583-597.
- [8] Richardson AE, Barea JM, McNeill AM, Prigent-Combaret C. Acquisition of phosphorus and nitrogen in the rhizosphere and plant growth promotion by

- microorganisms. Plant Soil 2009; 321:305-339.
- [9] Kumar A, Prakash A, Johri BN. *Bacillus* as PGPR in Crop Ecosystem. Chapter 2. In DK Maheshwari (ed.), *Bacteria in Agrobiolgy: Crop Ecosystems*, Springer-Verlag Berlin Heidelberg 2011;37-59.
- [10] Liu ZL, Sinclair JB. Population dynamics of *Bacillus megaterium* strain B153-2-2 in the rhizosphere of soybean. *Phytopathology* 1992;82:1297-1301.
- [11] Cakmakci R, F. Kantar, Algur OF. Sugar beet barley yields in relation to *Bacillus polymyxa* and *Bacillus megaterium* var. *phospaticum* inoculation. *J Plant Nutrition Soil Science* 1999;162:437-442.
- [12] Paul Eldor A. *Soil microbiology, ecology, and biochemistry*. Elsevier Inc (3 ed). Academic Press is an imprint of Elsevier 2007.
- [13] Nahid EA. Phenotypic and genetic variability among three *Bacillus megatherium* isolates. *In viro* evolution of Tri-Calcium Phosphate solubilizing potential and growth pattern. *Journal of American Sciences* 2010;6:111-115.
- [14] Chakraborty U, Chakraborty B, Basnet M. Plant growth promotion and induction of resistance in *Camellia sinensis* by *Bacillus megaterium*. *Journal of Basic Microbiology* 2006;46:186-195.
- [15] Lopez-Bucio J, Campos-Cuevas JC, Hernandez-Calderon E, Velasquez-Becerra C, Farias-Rodriguez R, Macias-Rodriguez LI, Valencia-Cantero E. *Bacillus megaterium* rhizobacteria promote growth and alter root-system architecture through an auxin-and ethylene-independent signaling mechanism in *Arabidopsis thaliana*. *Molecular Plant Microbe Interactions* 2007;20: 207-217.
- [16] Periago PM., Raquel C, Begoña D, Pablo SF, Alfredo P. *Bacillus megaterium* spore germination and growth inhibition by a treatment combining heat with natural antimicrobials. *Food Technol Biotechnol* 2006;44(1):17-23.
- [17] Borriess R, Maheshwari DK (ed.). *Bacteria in Agrobiolgy: Crop Ecosystems*. Chapter 3. Use of Plant-Associated *Bacillus* Strains as Biofertilizers and Biocontrol Agents in Agriculture. Springer-Verlag Berlin Heidelberg 2011;41-76.

- [18] Amalraj Daniel EL, Maiyappan S, John Peter A. *In vivo* and *In vitro* studies of *Bacillus megaterium* var. *phosphaticum* on nutrient mobilization, antagonism and plant growth promoting traits. *Journal of Ecobiotechnology* 2012;4(1):35-42.
- [19] Malanicheva IA, Kozlov DG, Sumarukova IG, Efremenkova OV, Zenkova VA, Katrukha GS, Reznikova MI, Tarasova OD, Sineokii SP, El'-Registan GI. Antimicrobial activity of *Bacillus megaterium* strains. *Microbiology* 2012;81(2):178-185.
- [20] Wiwattanapatapee R, Chumthong A, Pengnoo A, Kanjanamaneesathian M. Preparation and evaluation of *Bacillus megaterium* alginate microcapsules for control of rice sheath blight disease. *World J Microbiol Biotechnol.* 2013;29(8):1487-97.
- [21] Kong Q, Chi C, Yu J et al. The inhibitory effect of *Bacillus megaterium* on aflatoxin and cyclopiazonic acid biosynthetic pathway gene expression in *Aspergillus flavus*. *Applied microbial and cell physiology.* Appl Microbiol Biotechnol 2014;98:5161.
- [22] Kildea S, Ransbotyn V, Khan MR, Fagan B, Leonard G, Mullins E, Doohan FM. *Bacillus megaterium* shows potential for the biocontrol of *Septoria tritici* blotch of wheat. *Biological Control* 2008;47:37-45.
- [23] Islam KZ, Nandi B. Inhibition of some fungal pathogens of host phylloplane by *Bacillus megaterium*. *Zeitschrift Fur Pflanzenkrankheiten Und Pflanzenschutz-Journal of Plant Diseases and Protection* 1985;92:233-240.
- [24] Jung HK, Kim SD. An antifungal antibiotic purified from *Bacillus megaterium* KL39, a biocontrol agent of red-pepper Phytophthora blight disease. *Journal of Microbiology and Biotechnology* 2005;15:1001-1010.
- [25] Abhilash M, Shiva Reddy DM, Mohan BK, Nataraja S, Krishnappa M. Isolation and molecular characterization of *Bacillus megaterium* isolated from different agro climatic zones of Karnataka and its effect on seed germination and plant growth of *Sesamum indicum*. *Research Journal of Pharmaceutical,*

- Biological and Chemical Sciences 2010;1(3):3.
- [26] Reddy SV, Thirumala M, Mahmood SK. Production of PHB and P (3HB-co-3HV) biopolymers by *Bacillus megaterium* strain OU303A isolated from municipal sewage sludge. World J Microbiol Biotechnol 2009;25:391-397.
- [27] Khan JA. Biodegradation of azo dye by moderately halotolerant *Bacillus megaterium* and study of enzyme azoreductase involved in degradation. Advanced Biotech 2011;10:21-27.
- [28] Aftabuddin S, Kashem MA, Kader MA, Sikder MNA, Hakim MA. Use of *Streptomyces fradiae* and *Bacillus megaterium* as probiotics in the experimental culture of tiger shrimp *Penaeus monodon* (Crustacea, Penaeidae). Aquaculture, Aquarium, Conservation and Legislation. International Journal of the Bioflux SocietyAACL Bioflux 2013;6(3):253-267.
- [29] Shah MP, Patel KA, Nair SS, Darji AM. Potential effect of two *Bacillus* spp. on decolorization of azo dye. Journal of Bioremediation and Biodegradation 2013;4:7.
- [30] Maulin PS. Bioremedial application of *Bacillus Megaterium* PMS82 in microbial degradation of acid orange dye. International Journal of Environmental Bioremediation and Biodegradation 2014;2(3):93-99.
- [31] Gopinath SM, Ismail SM., Ashalatha, Aravind G. Bioremediation of lubricant oil pollution in water by *Bacillus megaterium*. International Journal of Innovative Research in Science, Engineering and Technology 2015;4(8):6773-6780.
- [32] Bary DA. Vergleichende morphologie und biologie der pilze. In Mycetozen und bacterien. Wilhelm Engelmann, Leipzig, Germany 1884.
- [33] Priest FG. Systematics and ecology of *Bacillus*. In: *Bacillus subtilis* and other Gram-positive bacteria - Biochemistry, physiology, and molecular genetics. In: Sonenshein AL, Hoch JA, Losick R (eds.) ASM press, American Society for Microbiology, Washington 1993; D.C. ISBN 1-55581-053-5.
- [34] Meinhardt F, Stahl U, Ebeling W. Highly efficient expression of homologous and heterologous

- genes in *Bacillus megaterium*. Appl Microbiol Biotechnol 1989;30:343-350.
- [35] Rygus T, Scheler A, Allmansberger R, Hillen W. Molecular cloning, structure, promoters and regulatory elements for transcription of the *Bacillus megaterium* encoded regulon for xylose utilization. Arch Microbiol 1991;155(6):535-42.
- [36] Yang Y, Biedendieck R, Wang W, Gamer M, Malten M, Jahn D, Deckwer WD. High yield recombinant penicillin G amidase production and export into the growth medium using *Bacillus megaterium*. Microb Cell Fact 2006;5:36.
- [37] Vary PS, Biedendieck R, Fuerch T, Meinhardt F, Rohde M, Deckwer WD, Jahn D. *Bacillus megaterium* from simple soil bacterium to industrial protein production host. Appl Microbiol Biotechnol 2007;76:957-967.
- [38] Pinotti LM, Silva AFS, Silva RG, Giordano RLC. Study of different media for production of penicillin G Acylase from *Bacillus megaterium* ATCC 14945. Applied Biochemistry and Biotechnology 2000;655-677.
- [39] de Souza VR, Silva ACG, Pinotti LM, Selistre Araújo HS, Giordano RLC. Characterization of the penicillin G Acylase from *Bacillus megaterium* ATCC 14945. Brazilian Archives of Biology and Technology 2005;48:105-111.
- [40] Srirangan K, Valerie O, Lamees A, Adam W, Murray MY, Perry C. Biotechnological advances on Penicillin G acylase: Pharmaceutical implications, unique expression mechanism and production strategies. Biotechnology Advances 2013; 31(8):1319-1332.
- [41] Biedendieck R, Malten M, Barg H, Bunk B, Martens JH, Deery E, Leech H, Warren JM, Jahn D. Metabolic engineering of cobalamin (vitamin B12) production in *Bacillus megaterium*. Microb Biotechnol 2010;3(1):24-37.
- [42] Hollmann R, Deckwer WD. Pyruvate formation and suppression in recombinant *Bacillus megaterium* cultivation. Journal of Biotechnology 2004;111(1):89-96.
- [43] Ekwealor IA, Obeta JAN. Studies on lysine production by *Bacillus megaterium*. African Journal of

- Biotechnology 2005;4 (7):633-638.
- [44] Jai S, Ghosh, Sagar S, Barale. Production, isolation and characterization of exotoxin produced by *Bacillus subtilis*, *Bacillus megaterium* and *Proteus vulgaris* and its significance in food poisoning. Int J Pharm Sci Rev Res 2015;35(2):245-249.
- [45] Ivatt RJ, Gilvarg C. Molecular structure of teichuronic acid of *Bacillus megaterium*. Biochemistry 1978;17:3997-4003.
- [46] Bunk B, Biedendieck R, Jahn D, Vary PS. *Bacillus megaterium* and Other *Bacilli*: Industrial applications. 2010.
- [47] Oyeleke SB, Auta SH, Egwim EC. Production and characterization of amylase produced by *Bacillus megaterium* isolated from a local yam peel dumpsite in Minna, Niger State. Journal of Microbiology and Antimicrobials 2010;2(7):88-92.
- [48] Nguyen T, Doan V, Le T. Amylase producing *Bacillus megaterium* T04 isolated in rach lang stream of vietnam. Journal of Applied Pharmaceutical Science 2015;5(10):012-015.
- [49] Neethu SK, Sreeja Devi PS, Arun SN. A review on microbial proteases. International Journal of Advanced Research 2016;4:2048-2053.
- [50] Wang W, Hollmann R, Deckwer WD. Comparative proteomic analysis of high cell density cultivations with two recombinant *Bacillus megaterium* strains for the production of a heterologous dextransucrase. Proteome Sci 2006;5:4-19.
- [51] Biedendieck R, Gamer M, Jaensch L, Meyer S, Rohde M, Deckwer WD, Jahn D. A sucrose inducible promoter system for the intra and extracellular protein production in *Bacillus megaterium*. J Biotechnol 2007;132:426-430.
- [52] Sameen F, Sullia SB, Deshmukh S. Optimization of physicochemical parameters for bacterial carboxy methyl cellulase from *Bacillus megaterium* (KU365409). International Journal of Innovative Research in Technology, Science and Engineering 2016;2(7):13.
- [53] Shimizu K, Nakamura H, Ashiuchi M. Salt inducible bionylon oolymer from *Bacillus megaterium*. Appl Environ Microbiol 2007;73(7):2378-2379.

- [54] Chettri R, Bhutia MO, Tamang JP. Poly- γ -Glutamic Acid (PGA) producing *Bacillus* species isolated from kinema, indian fermented soybean food. *Front Microbiol* 2016;7:971.
- [55] Kulpreecha S, Boonruangthavorn A, Meksiriporn B, Thongchul N. Inexpensive fed-batch cultivation for high poly (3-hydroxybutyrate) production by a new isolate of *Bacillus megaterium*. *J Biosci Bioeng* 2009;107:240-245.
- [56] Rodriguez-Contreras A, Koller M, Miranda-de Sousa Dias M, Calafell-Monfort M, Braunegg G, Marques Calvo MS. High production of poly (3-hydroxybutyrate) from a wild *Bacillus megaterium* Bolivian strain. *Journal of Applied Microbiology* 2013;1-10.
- [57] Zhang S, Xie Y, Zhang C, Bie X, Zhao H, Lu F, Lu Z. Biochemical characterization of a novel l-asparaginase from *Bacillus megaterium* H-1 and its application in French fries. *Food Research International* 2015;77:3527-533.
- [58] Mukesh kumara DJ, Silambarasanb T, Renugac R, Ravi kumara M, Karthigai devic S, Dhandapani R, Kalaichelvana PT. Production, optimization and characterization of α -amylase and glucose isomerase producing *Bacillus megaterium* BPTK5 from cassava waste. *European Journal of Experimental Biology* 2012;2 (3):590-595.
- [59] Archana DT, Kajal S, Lakshmi B. Study on tannase producing *Bacillus megaterium* isolated from tannery effluent. *International Journal of Advanced Research in Biological Sciences* 2016; 3 (7):28-35.
- [60] Donoghue HD. Production of megacins C and Cx: Presumptive evidence of extrachromosomal control. *Antimicrobial Agents and Chemother* 1997;11(1):34-37.
- [61] Brusilow WS, Nelson DL. Improved purification and some properties of megacin Cx, a bacteriocin produced by *Bacillus megaterium*. *Journal of Biology and Chemistry* 1981;256(1): 159-64.
- [62] Von Tersch MA, Carlton BC. Bacteriocin from *Bacillus megaterium* ATCC 19213: comparative studies with megacin A-216. *J Bacteriol* 1983;155:866-871.

- [63] Khalil R, Djadouni F, Elbahloul Y, Omar S. The influence of cultural and physical conditions on the antimicrobial activity of bacteriocin produced by a newly isolated *Bacillus megaterium* 22 strain. Afr J Food Sci 2009a;3:011-022.
- [64] Khalil R, Elbahloul Y, Djadouni F, Omar S. Isolation and partial characterization of a bacteriocin produced by a newly isolated *Bacillus megaterium* 19 strain. Pakistan J Nutr 2009b;8:242-250.
- [65] Abriouel H, Franz Charles MAP, Ben Omar N, Galvez A. Diversity and applications of *Bacillus* bacteriocins. FEMS Microbiol Rev 2011;35:201-232.
- [66] Sumi CD, Yang BW, Hahm YT. Antimicrobial peptides of the genus *Bacillus*: a new era for antibiotics. Can J Microbiol. 2015;61:93-103.
- [67] Eppinger M, Bunk B, Johns MA, Edirisinghe JN, Kutumbaka KK et al. Genome sequences of the biotechnologically important *Bacillus megaterium* strains M B1551 and DSM319. J Bacteriol 2011;193:16 4199-4213.
- [68] Vary PS. Prime time for *Bacillus megaterium*. Microbiology 1994;140:1001.
- [69] Hueck CJ, Kraus A, Schmiedel D, Hillen W. Cloning, expression and functional analyses of the catabolite control protein CcpA from *Bacillus megaterium*. Mol Microbiol 1995;16(5): 855-64.
- [70] Wittchen KD, Meinhardt F. Inactivation of the major extracellular protease from *Bacillus megaterium* DSM319 by gene replacement. Appl Microbiol Biotechnol 1995;42:871-877.
- [71] Wagner A, Elke KS, Wolfgang H. Sugar uptake and carbon catabolite repression in *Bacillus megaterium* strains with inactivated ptsHI. J Mol Microbiol Biotechnol 2000;2(4):587-592.
- [72] Burger S, Tatge H, Hofmann F, Genth H, Just I, Gerhard R. Expression of recombinant *Clostridium difficile* toxin A using the *Bacillus megaterium* system. Biochem Biophys Res Commun 2003;307(3):584-8.
- [73] Wang W, Hollmann R, Furch T, Nimtz M, Malten M, Jahn D, Deckwer WD. Proteome analysis of a recombinant *Bacillus megaterium* strain during

heterologous production of a glucosyl transferase. *Proteome Sci* 2005;3:4.

- [74] Malten M, Biedendieck R, Gamer M, Drews AC, Stammen S, Buchholz K, Dijkhuizen L, Jahn D. A *Bacillus megaterium* plasmid

system for the production, export and one step purification of affinity tagged heterologous levansucrase from the growth medium. *Appl Environ Microbiol* 2006;72(2):1677-9.